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Scientific opinion on the risks for animal and human health related to the presence of glycoalkaloids in feed and food, in particular in potatoes and potato-derived products

EFSA Panel on Contaminants in the Food Chain (CONTAM)

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Abstract

The European Commission asked EFSA for a scientific opinion on the risks for animal and human health related to the presence of glycoalkaloids (GAs) in feed and food. This risk assessment covers edible parts of potato plants and other food plants containing GAs, in particular tomato and aubergine. In humans, acute toxic effects of potato GAs (α-solanine and α-chaconine) include gastrointestinal symptoms such as nausea, vomiting and diarrhoea. For these effects, the CONTAM Panel identified a lowest-observed-adverse-effect level of 1 mg total potato GAs/kg bw per day as a reference point for the risk characterisation following acute exposure. In humans, no evidence of health problems associated with repeated or long-term intake of GAs via potatoes has been identified. No reference point for chronic exposure could be identified from the experimental animal studies. Occurrence data were available only for α-solanine and α-chaconine, mostly for potatoes. The acute dietary exposure to potato GAs was estimated using a probabilistic approach and applying reduction factors for food processing. Due to the limited data available, a margin of exposure (MOE) approach was applied. The MOEs for the younger age groups indicate a health concern for the food consumption surveys with the highest mean exposure, as well as for the P95 exposure in all surveys. For adult age groups, the MOEs indicate a health concern only for the food consumption surveys with the highest P95 exposures. For tomato and aubergine GAs, the risk to human health could not be characterised due to the lack of occurrence data and the limited toxicity data. For horses, farm and companion animals no risk characterisation for potato GAs could be performed due to insufficient data on occurrence in feed and on potential adverse effects of GAs in these species.

Keywords: glycoalkaloids (GAs), solanine, chaconine, potato, margin of exposure (MOE), food, feed

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Summary

The European Commission asked EFSA for a scientific opinion on the risks for animal and human health related to the presence of glycoalkaloids (GAs) in feed and food, in particular in potatoes and potato-derived products. This risk assessment covers edible parts of potato plants and also other food plants containing GAs, in particular tomato and aubergine. Non-edible parts of GA containing plants have not been considered, with the exception of potato sprouts.

GAs are present in many plants of the family of Solanaceae, and contribute to plant resistance against pests and pathogens. GAs are composed of a steroidal aglycone and an oligosaccharide side-chain. In commercial potato cultivars (*S. tuberosum*) the main GAs are α-chaconine and α-solanine, consisting of the aglycone solanidine and chacotriose and solatriose as oligosaccharide side-chains, respectively. The aubergine fruit (*S. melongena*) contains primarily the GAs α-solamargine and α-solasonine, composed of the aglycone solasodine and chacotriose and solatriose, respectively. In tomato fruit (*S. lycopersicum*) α-tomatine and α-dehydrotomatine are the major GAs, with lycotetraose coupled to the aglycones tomatidine and tomatidenol, respectively.

Human risk assessment

In experimental animals, the potato GAs, α-solanine and α-chaconine, show a relatively low oral bioavailability, with differences between species. Hamsters exhibit higher absorption and slower excretion rates for both substances when compared to rats. Due to the limited information, the metabolic profiles of potato GAs in experimental animals could not be characterised.

In humans, α-solanine and α-chaconine are systemically absorbed following ingestion. For both substances, relatively long serum half-lives were reported suggesting a possible accumulation. The blood clearance of the respective aglycone solanidine appears to be slow. Accordingly, levels of solanidine were regularly detected in the blood of human volunteers in several studies, suggesting hydrolysis of GAs. No further information is available on metabolism and excretion of potato GAs in humans.

There are no toxicokinetic data on tomato and aubergine GAs and their aglycones in experimental animals and humans.

In acute oral toxicity studies no adverse effects of α-solanine were observed at doses of 250 mg/kg body weight (bw) per day in rats and 1,000 mg/kg bw per day in mice. Reliable data on other potato GAs or tomato and aubergine GAs and their aglycones are missing.

In repeated oral dose studies on potato GAs, rodents showed non specific effects, such as reduced body weight and relative liver weight with indication of similar potencies of α-solanine and α-chaconine. Hamsters exhibited these symptoms after a 5 days treatment with 100 mg of α-solanine or α-chaconine/kg bw per day, while mice showed these effects after one week of daily treatments with 416.6 mg of α-solanine or 409 mg of α-chaconine/kg bw. Solanidine, however, increased the absolute and relative liver weight at 190.9 mg/kg bw per day in mice, suggesting a different effect of the aglycone compared to the GAs.

The tomato GA, α-tomatine, and its aglycone, tomatidine, exerted no effects in rats, when applied at 20 mg/kg bw per day for a period of 200 day. At higher doses, tomatidine reduced the cholesterol uptake and increased fecal sterol and coprostanol excretion in hamsters and rats. In mice, a 1–2 weeks' treatment with the aubergine GA, α-solasonine, increased the body weight gain at 424.4 mg/kg bw per day, while its aglycone, solasodine, decreased body weight gain and caused gastric gland degeneration and liver toxicity at 160 mg/kg bw per day.

Developmental studies have been performed mainly in hamsters treated with potato GAs and their aglycones for only one day or for a short, very restricted time period during gestation. Outcomes were mainly analysed in late gestational embryos and comprised effects in the central nervous system, predominantly exencephaly, encephalocele and anophthalmia. These malformations occurred at doses of 165 mg/kg bw per day and above for GAs, and of 115 mg/kg bw per day and above for the aglycones. No no-observed-adverse-effect level (NOAEL) or lowest-observed-adverse-effect level (LOAEL) could be identified from these studies. Reduced postnatal survival of pups due to insufficient milk production were reported when pregnant Holtzman rats had been exposed to 3.6 mg of α-solanine/kg bw per day. Studies on the male fertility in dogs have been performed only with the aubergine aglycone, solasodine. Decreased epididymal weight and cauda epididymal epithelial height, and also an epididymal lumen depleted of sperm occurred in dogs after 80 mg/kg bw per day given for 1 month. Similar effects were observed in Rhesus monkeys exposed to 100 mg/kg bw per day for 5 months.

From the limited number of studies available, there was no evidence for genotoxicity of the potato GAs α-solanine and α-chaconine, and the aglycone, solanidine, as well as for the aubergine GA, α-solamargine.

No long-term chronic toxicity/carcinogencity study for potato, tomato or aubergine GAs or for the respective aglycones could be identified.

In humans, acute toxic effects following ingestion of potato GAs include gastrointestinal symptoms of varying severity such as vomiting, diarrhoea and abdominal pain, which may occur from a total potato GAs (potato TGA) intake of 1 mg/kg bw or more. Further symptoms, including drowsiness, apathy, confusion, weakness, vision disturbances, rapid and weak pulse, and low blood pressure may be the consequence of dehydration. In severe cases, paralysis, respiratory insufficiency, cardiac failure, coma and death have been reported. Doses in the range of 3–6 mg potato TGAs/kg bw are considered to be potentially lethal for humans. Results from limited volunteer studies suggest possible differences in the human population with respect to the individual susceptibility towards adverse effects associated with the intake of potato GAs.

Regarding the mode of action, adverse effects of GAs may be due to their ability to complex with membrane 3β-hydroxy sterols, thereby causing disruption and loss of integrity of cell membranes. After oral exposure, these effects may affect the mucosa of the gastrointestinal tract and cause the symptoms observed in intoxicated humans, such as nausea, vomiting and diarrhoea.

GAs inhibit acetylcholinesterase (AChE) and serum butyrylcholinesterase (BuChE) by a reversible, competitive mode of action. The relative potency of inhibition of α-solanine and α-chaconine appear to be similar. The aglycones exert weak or no inhibitory effects. The excess of acetylcholine at the neuronal and neuromuscular junctions upon inhibition of the enzymes might also contribute to the symptoms described for intoxications with GAs.

At high doses, α-tomatine may form a non-absorbable complex with cholesterol and other sterols in the enteral lumen which may impair the absorption of cholesterol. As a consequence, blood cholesterol levels were lowered in rodents.

The CONTAM Panel considered that the use of rodent data on acute toxicity was not appropriate to establish a reference point for acute exposure to potato GAs in humans. The CONTAM Panel selected the LOAEL of 1 mg potato TGA/kg bw per day as the reference point for acute risk characterisation, based on human data from case reports, outbreaks and studies in volunteers. The available data on acute toxicity were considered insufficient to establish a health-based guidance value. Instead, the Panel used the margin of exposure (MOE) approach to assess a possible health concern from acute exposure to potato TGAs via food.

Assuming the main symptoms to be mainly due to local irritation of the gastrointestinal mucosa rather than inhibition of AChE activity, the Panel considered that the possible interindividual variability in toxicodynamics is more relevant than the interindividual variability in toxicokinetics. Accordingly, an MOE higher than 10 indicates that there is no health concern. This MOE of 10 takes into account the extrapolation from a LOAEL to a NOAEL (a factor of 3) and the interindividual variability in toxicodynamics (a factor of 3.2).

The experimental data available for repeated dose toxicity are not sufficient to identify a reference point for chronic exposure to potato GAs. In humans, no evidence of health problems associated with repeated or long-term intake of GAs via potatoes has been identified.

Regarding GAs or aglycones occurring in edible parts of food plants other than *S. tuberosum*, no suitable study for determining a reference point for tomato or aubergine GAs or aglycones was identified.

Occurrence data were only available for α-solanine and α-chaconine, and mostly for 'Main crop potatoes' and 'New potatoes'. Few data were available for processed food. No data on the occurrence of tomato and aubergine GAs and their aglycones were submitted to EFSA.

Since the occurrence data on potato GAs did not cover all the food categories containing potatoes in the Consumption Database, it was decided that the best approach for the exposure assessment would be to use the occurrence data in the raw primary commodities (RPC) (main-crop potatoes and new potatoes) and the RPC Consumption Database. The Panel decided to combine the occurrence of 'New potatoes' with that of 'Main-crop potatoes' and the mean upper bound (UB) occurrence (sum of α-solanine and α-chaconine) for these two groups was 52.0 mg/kg and the P95 occurrence was 117.0 mg/kg. The minimum and maximum reported concentrations were 1.1 mg/kg and 550.3 mg/kg, respectively.

The acute dietary exposure to potato TGAs was estimated using a probabilistic approach, including only days in which there was consumption of main-crop potatoes. As no occurrence data were available for GAs in tomato and aubergine, these foods were not included in the exposure assessment.

Processing of potatoes has been reported to reduce the content of GAs in the final processed product. In general, and according to the literature, the peeling of potatoes reduced the GA content by 25–75%, boiling in water and blanching of peeled potatoes by 5–65%, and frying in oil of peeled potatoes by 20–90%. Microwave and oven baking of unpeeled potatoes may cause a reduction in the GA content by 3–45% and by 20–50%, respectively. No information has been found about the chemical nature of the GA degradation products. For the exposure assessment, reduction factors for the major food processing steps, comprising peeling and heat processing (boiling, frying, baking), were applied to the occurrence data as follows: reduction factors between 0.25 and 0.75 were attributed to the peeling of potatoes, between 0.2 and 0.9 for frying and deep frying, and between 0.05 and 0.65 for all other cooking methods.

Information about the peeling of potatoes was not available in the consumption database, but it was assumed that 90% of the potatoes are consumed as peeled. Where information of the cooking method was not available, a cooking method was randomly attributed to the eating event based on the relative frequency of cooking methods reported.

The mean UB exposure to potato TGAs across surveys ranged from 26.0 µg/kg bw per day in adults to 193.4 µg/kg bw per day in toddlers. The 95th percentile exposure ranged from 88.2 µg/kg bw per day in adults to 617.9 µg/kg bw per day in toddlers (up to 1,057.9 µg/kg bw per day in the upper limit of the 95% confidence interval).

Comparing the LOAEL for potato TGAs of 1 mg/kg bw per day with the acute exposure estimates, the MOEs for the younger age groups indicate a health concern for the food consumption surveys with the highest mean exposure, as well as for the P95 exposure in all surveys. For adult age groups, the MOEs indicate a health concern only for the food consumption surveys with the highest P95 exposures.

The CONTAM Panel calculated the mean percentage of days with potato consumption across surveys per age group on which the potato TGA intake may be below the MOE of 10. The highest number of survey days with intake of potatoes below the MOE of 10 was estimated for toddlers (56%) followed by children (50%). For the other age groups the estimated intake was calculated to be below the MOE of 10 in 23 to 40% of the survey days.

For tomato and aubergine GAs, the risk to human health could not be characterised due to the lack of occurrence data in food and the limited information on the adverse effects in experimental animals and humans.

The CONTAM Panel considered that the impact of the uncertainties on the risk assessment of acute exposure to potato GAs in food is moderate, and that overall, the identified uncertainties may either cause an over- or underestimation of the risk.

Farm animals, horses and companion animals risk assessment

Information on the toxicokinetics of GAs was limited to ruminants, for which the data suggest an extensive conversion of α-solanine and α-chaconine to aglycones in rumen, and a low potential of solanidine to transfer into cows' milk.

No data on the potential adverse effects of potato GAs in horses, companion animals (cats and dogs) or fur animals were identified. Due to an insufficient database on the adverse effects of GAs in ruminants, pigs, poultry, rabbits and fish, an acute reference dose could not be derived.

Potatoes are not grown specifically as feed for livestock, but when supply exceeds market requirements for human consumption whole (raw) potatoes may be used as feed for ruminants and pigs. Some by-products of potato processing and starch extraction are used as feeds for farmed livestock, principally non-ruminants, and for companion animals.

Data on potato GAs in feed were insufficient to perform an exposure assessment.

Thus, no risk characterisation could be performed due to insufficient occurrence data of GAs for feed and the lack of, or limited, data on the adverse effects of GAs in farm animals, horses or companion animals.

Recommendations

The following needs have been identified to improve the risk assessment for humans and reduce the uncertainties:

- Research on the occurrence of GAs and their aglycones and other potentially toxicologically relevant secondary plant metabolites in the potato cultivars available on the market and on new potato cultivars resulting from breeding experiments.
- Occurrence data on GAs and their aglycones in potato processed products, including foods for infants.
- Occurrence data on GAs and their aglycones in tomato and aubergine and products thereof.
- Data on the toxicokinetics of potato, tomato and aubergine GAs and aglycones in experimental animals and humans.

- Data on repeated dose toxicity, including reproductive and developmental toxicity of potato, tomato and aubergine GAs and aglycones in experimental animals.
- Studies in humans linking dietary exposure, biomarkers of exposure and adverse effects.

The following needs have been identified to improve the risk assessment for farm animals, horses and companion animals and reduce the uncertainties:

- Occurrence data on potato GAs and their aglycones in feed.
- Studies on the kinetics and the potential adverse effects from feed material containing GAs of potato GAs in farm animals, horses and companion animals.

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1. Introduction

1.1. Background and Terms of Reference as provided by the requestor

Background

Many plants in the family *Solanaceae* contain glycoalkaloids, and they are considered to be natural toxins. The plant glycoalkaloids are toxic steroidal glycosides and the commonest types found in food plants are α-solanine and α-chaconine. Their natural function is probably to serve as stress metabolites or phytoalexins for the protection of the plant when attacked by insects, fungi, etc.

Amongst the most widely cultivated food crops, aubergines, tomatoes and potatoes are in the *Solanaceae* family, but the levels of glycoalkaloids in tomatoes and aubergines are generally quite low.

The glycoalkaloids of most relevance to food safety are those occurring in the potato. The predominant toxic steroidal glycosides in potato are α-solanine and α-chaconine. They occur in potato tubers, peel, sprouts, berries, leaves and blossoms and their concentration in tubers depends on a number of factors, such as cultivar, maturity and environmental factors. Concentrations of glycoalkaloids are 3 to 10 times greater in the peel than in the flesh. There is considerable variation in glycoalkaloid content among potato cultivars. Storage conditions, especially light and temperature, are mainly responsible for increases in solanine during marketing. Although the glycoalkaloid content can increase in the dark, the rate of formation is only about 20% the rate of formation in light. Increases of solanine in the potato peel are closely associated with greening (synthesis of chlorophyll) of the peel. These biochemical processes are independent of each other, but are both activated by light. Glycoalkaloids are relatively stable in potatoes and levels are not affected to a large extent by boiling, freeze-drying, or dehydration. Microwave cooking has only a limited effect, but cooking at temperatures at or above 170°C is more effective at reducing.

Symptoms associated with glycoalkaloid poisoning from potatoes include a bitter or burning sensation in the mouth and flu-like symptoms such as nausea, vomiting, stomach and abdominal cramps, and diarrhoea. More severe cases of glycoalkaloid poisoning may be accompanied by a variety of neurological effects (i.e. drowsiness, apathy, restlessness, shaking, confusion, weakness, and disturbed vision). There are a few reports of deaths being attributed to glycoalkaloid exposure from the consumption of potatoes, potato leaves, and potato berries.

Potatoes and potato-derived products are listed in the Catalogue of feed materials[2].

Terms of Reference

In accordance with Art. 29 (1) of Regulation (EC) No 178/2002, the European Commission asks the European Food Safety Authority for a scientific opinion on the risks for animal and human health related to the presence of glycoalkaloids in feed and food, in particular in potatoes and potato-derived products.

[2] Commission Regulation (EU) No 6812013 of 16 January 2013 on the Catalogue of feed materials (0J L 29, 30.1.2013, p. 1)

1.2. Interpretation of the Terms of Reference

The CONTAM Panel considered that the opinion should cover edible parts of potato plants and also of other food plants containing glycoalkaloids (GAs), e.g. tomato and aubergine. Non-edible parts of GA containing plants have not been considered, with the exception of potato sprouts. In particular, the CONTAM Panel concluded this Opinion should comprise the:

a) evaluation of the toxicity of GAs in feed and food, in particular in potatoes and potato-derived products, for farm and companion animals and humans considering all relevant toxicological endpoints;

b) evaluation of the alkaloid profile (i.e. composition of the alkaloids and their concentration) of the food and feed samples submitted to EFSA;

c) estimation of the dietary exposure of the European population to GAs in food, in particular in potatoes and potato-derived products, including the consumption patterns of specific groups of the population if appropriate;

d) estimation of the dietary exposure of farm and companion animals to GAs in feed, in particular in potatoes and potato-derived products;

e) assessment of the human health risks for the European population, including specific groups of the population if appropriate, as the consequence of the estimated dietary exposure;

f) assessment of the farm and companion animal health risks in Europe as the consequence of the estimated dietary exposure. Exposure to GAs from weeds containing GA is only addressed in this Opinion in the context of accidental intake by farm animals.

When referring to GAs in potatoes, the term total GAs (TGA) refers to a material comprising α-solanine and α-chaconine as major fraction, with no specification on the occurrence of minor GAs as well as β- and γ-forms of solanine and chaconine. Similarly, when referring to tomato and aubergine, the term TGA refers to the GAs from the corresponding species and forms thereof.

1.3. Supporting information for the assessment

1.3.1. Chemistry

Solanine is one of the first alkaloids that has been isolated from nature, by Desfosses in 1820 (Friedman and McDonald, 1997). In 1861, Zwenger and Kind reported that solanine contains a glycoside sidechain (Zwenger and Kind, 1861). Only in 1954 it was shown that solanine extracted from potato is in fact a mixture of two glycoalkaloids (GAs), α-solanine and α-chaconine, that share the same solanidine aglycone (Kuhn and Löw, 1954). Since then, at least 90 different GAs have been isolated and fully structurally elucidated from over 300 species of the Solanaceae family (Sánchez-Mata et al., 2010; Al Sinani and Eltayeb, 2017). The chemical structures and some physical properties of the most important ones are listed in **Appendix A**.

GAs are composed of a steroidal aglycone and an oligosaccharide side-chain attached to the 3β-hydroxy group of the aglycone (see **Figure 1**). The GAs of relevance can be divided into the (i) solanidane group, with solanidine as the steroid backbone, and the (ii) spirosolane group, with either the solasodine or the tomatidenol/tomatidine backbone. GAs often contain a double bond between C_5 and C_6, but the corresponding 5a, 6-hydrogenated forms are also common and, in some species (e.g. tomato) they constitute the major components. The stereochemistry at carbons C_{22} and C_{25} is well defined: in solanidine the configuration is 22R, 25S, in solasodine it is 22R, 25R, and in tomatidenol/tomatidine the configuration is 22S, 25S.

Further diversification is generated by the composition of the glycoside side-chain. Most GAs contain either a trisaccharide (chacotriose or solatriose) or a tetrasaccharide (lycotetraose) as carbohydrate. In commercial potato cultivars (*Solanum tuberosum*) mostly α-chaconine and α-solanine (composed of the solanidine aglycone and chacotriose and solatriose, respectively) are present (**Figure 1**). Wild *S. tuberosum* varieties may contain a much wider range of GAs. The aubergine fruit (derived from *S. melongena*) contains primarily α-solamargine and α-solasonine (composed of the solasodine aglycone and chacotriose and solatriose, respectively). In tomato fruit (derived from *S. lycopersicum* varieties) α-tomatine and α-dehydrotomatine are the major compounds (composed of the aglycones tomatidine and tomatidenol, respectively, coupled to lycotetraose).

The prefix alpha (α-) refers to the intact glycoside while the prefixes beta (β-), gamma (γ-) and delta (δ-) refer to the corresponding GAs with progressively truncated carbohydrate side-chains due to the action of enzymatic or acidic hydrolysis.

GAs are relatively stable to heat and alkaline conditions. However, acidic hydrolysis using strong acid and elevated temperatures may cleave the glycosidic bonds and liberate the aglycone together with the corresponding reducing sugars (Friedman, 2006). The aglycones, due to their steroid nature are very poorly soluble in water, but the intact GAs are soluble in water.

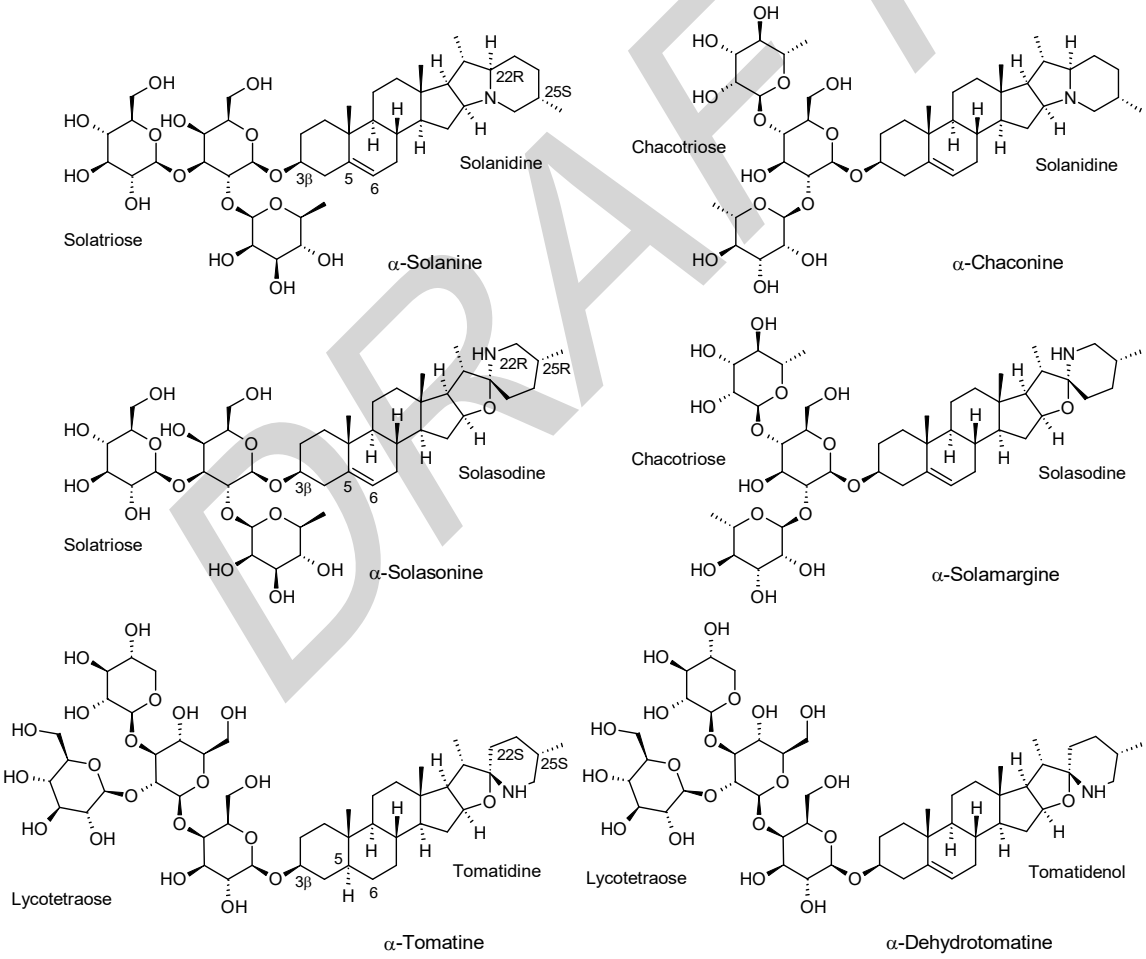


Figure 1. Structure of the most important glycoalkaloids (GAs) found in potato (α-solanine and α-chaconine), aubergine (α-solasonine and α-solamargine) and tomato (α-tomatine and α-dehydrotomatine).

1.3.2. Analytical methods

With older analytical methods (gravimetric, colourimetric) it was often not possible to discriminate the individual GAs. Nowadays most confirmatory methods use liquid chromatography (LC) to separate the intact GAs and detection is performed either by ultraviolet-visible (UV-vis) detection or by mass spectrometry (MS). Assays based on thin layer chromatography (TLC) or immunoassays (i.e. enzyme-linked immunosorbent assay (ELISA)) are also available and can be applied for screening.

For reliable quantification of GAs, reference standards of high purity are required, which are available for the most common ones (α-solanine, α-chaconine, α-tomatine, α-solasodine, α-solamargine, and the corresponding aglycones). For many of the less common GAs (**Appendix A**) no reference standards are currently available.

1.3.2.1. Extraction and clean-up

When analysing the intact GAs, an important aspect is the stability of the compounds. Enzymatic degradation of the glycoside side-chain due to the action of glycosidases present in fresh potato tubers, may occur during extraction when not properly controlled (Friedman and McDonald, 1997). This may result in the underestimation of the GA content, if only the intact GAs are monitored. Two approaches are generally followed to prevent from losses due to enzymatic activity. One approach is to cut or slice the potato tubers, add liquid nitrogen and to lyophilise by freeze-drying. The freeze-dried material is ground and homogenized to a fine powder (Friedman and Dao, 1992). Extraction of the lyophilised material is typically performed with an acidic aqueous solvent (e.g. acetic acid), mixed with a suitable organic solvent (e.g. methanol, acetonitrile). The other approach is to shred the potato tubers in a food processor, add liquid nitrogen and grind to a fine powder (Hellenäs, 1986; AOAC Official Method 997.13, 2000). Extraction of the frozen material is conducted with a mixture of 5% acidic acid to which 0.5% sodium bisulfite ($NaHSO_3$) has been added. For both methods applies that further clean-up is necessary before the extracts can be analysed by high performance LC (HPLC)-based methods. Clean-up is preferably conducted using solid-phase extraction (SPE) cartridges (typically C18-based, but cartridges with NH_2, CN stationary phases have also been used) (Friedman and McDonald, 1997).

1.3.2.2. Detection methods

Gravimetric methods

In gravimetric methods the GAs are measured by alkaline precipitation from potato extracts. GAs are extracted from the tubers using a strong acid, such as sulfuric or hydrochloric acid, that results in hydrolysis of the GAs to the corresponding aglycone. Addition of a surplus of alkaline, e.g. calcium hydroxide, to the solution results in a precipitation of the alkaloids, that can be collected, dried and weighted (Bömer and Mattis, 1924). Gravimetric methods have a limit of quantification (LOQ) of approximately 10 mg total GAs (TGA)/kg. Gravimetric methods have been in general use until the development of colourimetric methods in the 1930s and 1940s. Gravimetric methods have a number of disadvantages, such as losses occurring during precipitation, lengthy procedures, and the fact that other alkaloids may co-precipitate (Lepper, 1949; Friedman and McDonald, 1997).

Colourimetric methods

The first colourimetric method to quantify the content of solanine in potato tubers was developed by Alberti in 1932, using sulfuric acid and formaldehyde to generate a coloured derivative (Alberti, 1932). Many different methods using different reagents have been described in the literature in the following decades. Clement and Verbist (1980) compared nine different colourimetric methods for the analysis of solanidine in potato tubers. They noted that a considerable variation in the results was obtained, which was attributed to unspecific reactions with other plant components with the staining reagents used (resulting in overestimation of the content in some methods) or to losses during extraction (leading to

an underestimation of the content in other methods). It was reported that sensitivity between methods differed up to 20-fold, but limits of detection (LODs)/LOQs were not determined (Clement and Verbist, 1980).

Another important drawback of colourimetric methods is that they require hydrolysis of the glycoside side-chain prior to derivatisation. As a result, only the TGA content can be determined and no information on the individual glycosides is obtained. Since the 1980s colourimetric methods have been largely replaced by HPLC-based methods.

TLC with UV-Vis detection

TLC is often used as a quick, qualitative, screening tool to assess the presence of GAs in plant extracts. However, quantitative methods using high performance TLC (HPTLC)-densitometry have also been described. Bodart et al. (2000) described a method for the determination of α-solanine and α-chaconine in potato extracts by means of HPTLC-densitometry. The GAs extracted from potato were separated on the TLC plate and derivatised to generate chromophoric compounds that were measured at 507 nm with a densitometer. The LOQ for both GAs was 100 ng on plate, corresponding to 40 mg/kg in dehydrated peeled potato powder and to 10 mg/kg in fresh peeled potato tubers. A modified HPTLC method was described by Skarkova et al. (2008), with an LOQ of 5 mg/kg for α-solanine and α-chaconine for peeled, uncooked potatoes.

Enzyme linked immunosorbant assays (ELISA)

A few reports using immunoassays (ELISA) for the quantification of GAs have been published. Morgan et al. (1985) developed an ELISA assay to determine the TGA content in potato tubers and compared their assay with two existing colorimetric methods (Morgan et al., 1985). The sensitivity of the ELISA assay was excellent (0.02 mg/kg) and did not show cross reactivity with tomatidine derivatives. Comparison with a HPLC-ultraviolet (UV) method showed a good linear relationship, when corrected for recovery losses (Hellenäs, 1986).

Stanker et al. (1994) developed an ELISA based on monoclonal antibodies that showed affinity for solanidine, tomatidine and solasodine derivatives. The commercialised version of the ELISA kit was found to work well for the quantification of TGAs in potato tubers, sprouts, potato fries and chips (Friedman et al., 1998). The ELISA displayed good cross reactivity for solanidine derivatives (α-solanine, α-, β-, γ-chaconine), lower sensitivity for α-tomatine and very limited sensitivity for solasodine derivatives. For all investigated products, a good correlation for TGA content was observed with results obtained by HPLC-UV analysis. The LOD for TGAs was estimated at 1 mg/kg.

Gas chromatography (GC) with FID, MS or NPD detection

Analysis of intact GAs by GC-flame ionisation detector (FID) or GC-MS is possible only after derivatisation of the glucoside hydroxyl groups (Herb et al., 1975). However, it is more common to analyse the aglycones after hydrolysis, because derivatisation in that case is not necessary (Friedman and McDonald, 1997; Van Gelder, 1985). Bushway et al. (1984) have developed a GC-nitrogen-phosphorus detector (NPD) method for solanidine in milk, with an LOQ of 0.28 mg/L.

HPLC with UV-VIS detection

Intact GAs as well as the aglycones can be measured by HPLC-UV detector at wavelengths around 200 nm (Houben and Brunt, 1994; Friedman and McDonald, 1997; AOAC Official Method 997.13, 2000). Separation of α-solanine and α-chaconine, as well as the separation of α-tomanine and α-dehydrotomatine, can be achieved on reversed phase C18 chromatographic columns using aqueous phosphate buffers in combination with an organic modifier, typically methanol or acetonitrile (Kozukue et al., 2004). It is possible to separate these GAs by a simple isocratic elution, but for the analysis of a wider range of GAs gradient elution is necessary (Eanes et al., 2008; Maurya et al., 2013). Due to the

fact that GAs do not show significant UV absorption at wavelengths above 215 nm, the analysis needs to be conducted at low, unspecific, wavelengths. It is therefore necessary to include purification steps, e.g. by SPE clean-up, prior to analysis by HPLC-UV (Friedman and McDonald, 1997; Keukens et al., 1994). The LOD is around 2.5 mg/kg for the individual compounds.

LC-MS methods (LC-MS/MS, LC-HRMS)

LC-MS based methods are becoming increasingly popular in the analysis of GAs. The most important advantage of MS detection above UV detection is the much higher selectivity of the MS. Combined with the high sensitivity of modern mass spectrometers, sample preparation and clean-up can often be minimised and simple dilution of extracts may be sufficient. Methods based on LC–tandem mass spectrometry (LC-MS/MS) are often used for targeted analysis of GAs (Jandrić et al., 2011; Hossain et al., 2015; Nara et al., 2019), while methods using LC coupled to high resolution MS (LC-HRMS) or to Orbitrap MS (LC-Orbitrap-MS) are more commonly used for metabolomic studies and exploratory research (Moco et al., 2007; Caprioli et al., 2014). Hossain et al. (2015) reported LOQs for α-solanine and α-chaconine of 2 µg/L in potato extract, corresponding to 40 µg/kg in dehydrated potato powder. Nara et al. (2019) reported an LOQ of 1 and 2 µg/L in whole blood for α-solanine and α-chaconine, respectively.

1.3.3. Sources

Botanical sources of GAs with a steroidal aglycone are mainly found within the dicotyledoneous family Solanaceae (see **Section 1.3.1**), and to a lesser extent in the monocotyledonous family Liliaceae (Roddick, 1996). From the latter family only few intact GAs have been isolated, and their fully structure elucidated, while a large number of steroidal alkaloids have been found and fully identified in extracts, particularly of *Veratrum* and *Fritillaria* species (Liliaceae) following acidic hydroflysis step (Li et al., 2006). In general, the two monocotyledonous genera are not known to be of any use as sources of food, although two species of so-called 'riceroot' (*Fritillaria camschatcensis* [L.] Ker-Gawl. and *F. lanceolata* Pursh) are reported to have been important root foods for the Indians of the Northwest Coast of North America (Turner and Kuhnlein, 1983).

This section focuses on the species of importance of today's food production from the genus *Solanum*, together with a few other *Solanum* species known as possible sources of intoxications in man and farm animals.

The genus *Solanum* L. that contains the GAs as defined in **Section 1.3.1**, include around 1,500 species, and as such the largest genus in the family Solanaceae (Knapp, 2013; Tepe et al., 2016). The genus comprises the potato (*S. tuberosum*), the tomato (*S. lycopersicum* L. – synonyms *Lycopersicon lycopersicum* (L.) H. Karst, and *Lycopersicon esculentum* Mill.), and the aubergine (*S. melongena* L.) (Milner et al., 2011). Less well-known plants are gboma (*S. macrocarpon* L.) and the scarlet aubergine (*S. aethiopicum* L.) (Sánchez-Mata et al., 2010). Other consumed food plants belonging to the genus *Solanum* L. are the pepino (*S. muricatum*, also known as sweet pepino or sweet cucumber) from which the ripe fruit is eaten (Sánchez-Mata et al., 2010), and the wild aubergine (*S. torvum*) from which the fruits, known as susumber or turkey berries, are eaten as vegetables in the traditional Jamaican cuisine (Smith et al., 2008; Karmakar et al., 2015).

Non-food *Solanum* species that produce fruits that might be accidentally ingested are the soda apple (*S. aculeastrum* Dunal, also known as goat apple or poison apple) and the wolf apple (*S. lycocarpum* St. Hil) (Bhat and Pallyath, 2016). The latter is a common bush in parts of Brazil, and the fruits are used in folk medicine, although the unripe fruits are slightly toxic, especially to pregnant women (Schwarz et al., 2007). Bittersweet nightshade (*S. dulcamara*) and black nightshade (*S. nigrum* L.) are common toxic weed plants (Milner et al., 2011).

For the potato (*S. tuberosum*) it is the starchy stem tuber that is eaten. Both sexual and vegetative reproduction occurs in the ancestral wild species and the improved cultivars. Thus, potatoes flower and set true seeds in berries (the fruit) following natural pollination by insects, while the tubers propagate clonally. For the tomato (*S. lycopersicum*), both the mature and the immature (green) fruit are used as food (Martínez-Valverde et al., 2002). The food use of the aubergine (*S. melongena*) concentrates around the mature fruit. For the production the plant is grafted onto other species of Solanum, such as *S. torvum* (Miceli et al., 2014).

Different plant parts contain different concentrations of the individual GAs and TGAs. Furthermore, both the total TGA content and the relative proportions in a plant part (organ), will change as a result of the developmental stage of the organ.

The three main food crops (potatoes, tomatoes and aubergine) are discussed in more detail below.

1.3.3.1. Potatoes

S. tuberosum is the most cultivated tuber of the Solanum species. Potatoes are one of the main staple foods produced worldwide. In 2016, the global production volume was around 375 million tonnes (FAOSTAT, 2018a), which is about half the yearly volume produced of wheat and of rice. In the EU, Germany is the biggest producer of potatoes with over 10 million tonnes yearly, followed by Poland, France, The Netherlands and the UK with production volumes in the range of 5 to 8 million tonnes.

Potato breeding programs have for quite a long time used many primitive forms of cultivated potatoes and their wild relatives as a valuable source of genetic variation. Especially breeding programs concerning disease resistance and environmental tolerance have been successful. Several methods to overcome hybridization barriers have been developed (Machida-Hirano, 2015).

Studies on the occurrence of GAs in commercial (common) potatoes (*S. tuberosum* ssp. *Tuberosum*) measured and reported only on α-chaconine and α-solanine, while the tubers of wild plants of the subspecies andigena have been shown to also contain, e.g. β-chaconine (Distl and Wink, 2009). Both the common potato and other potato species, whether cultivated and consumed or not, may contain GAs other than α-solanine and α-chaconine. However, in general, the tuber concentrations of these other GAs will be low. In case these species are used in breeding, transfer (depending on the crossing methods used) of the biosynthetic pathway for the formation of these other GAs can occur.

According to Milner et al. (2011), the TGAs in different parts of the cultivated potato plant (*S. tuberosum*) will often be found within the concentration ranges given in **Table 1** (see **Figure 2** for the internal structure of a potato tuber). Berries of potatoes have about 10 times the levels of GAs compared to the tubers of commercial potato cultivars (Friedman, 1992).

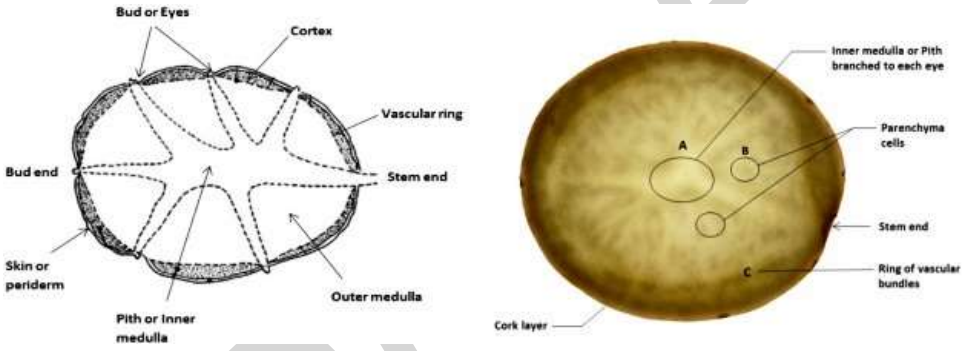
In many regions of the world the potato today is cultivated all year round by using different types of cropping and different cultivars adapted to these, i.e. to different temperature, day length, and growing period among others (Mori et al., 2015). Besides the cultivar characteristics and the type of cropping, also the end use of the given potato matters. Potatoes can be classified into four types, namely for 'table use', 'industrial food processing use(s)', 'starch production', and 'other purposes', including colourful potatoes (Mori et al., 2015). Due to the constant breeding for new cultivars, the GA content ranges given in **Table 1** must be considered as approximate. Concerning potatoes with a food end-use, the TGA content as well as the relative concentration of the different GAs has been reported in several studies. Details of the variation of GAs in different cultivars are discussed in **Section 3.2.2.**

632 **Table 1.** Total glycoalkaloids (TGA) concentration (mg/kg fresh weight (fw)) in various cultivated potato
633 (*S. tuberosum*) plant parts [(a)].

Plant part	Concentration range (mg/kg fw)	Original Reference [(a)]
Flowers	2,150–5,000	(1), (2)
Leaves	230–1,000	(3)
Stems	23–33	(4)
Roots	180–400	(4)
Bitter-tasting tubers	250–800	(5)
Whole tuber	10–150	(6)
Skin (2–3% of tuber)	300–640	(1), (2)
Peel (10–12% of tuber)	150–168	(1), (2)
Cortex	125	(6)
Flesh	12–100	(1), (2)
Pith [(b)]	Not detected	(6)
Sprout	2,000–7,300	(1), (2)

634 (a): From Milner et al. (2011), who collected the information from several research articles given in the review. Original references:
635 (1) Lampit et al. (1943), (2) Van Gelder et al. (1991), (3) Kozukue et al. (1987), (4) Sinden et al. (1984), (5) Zitnak (1961),
636 (6) Kozukue and Mizuno (1989).
637 (b): The pith is the centre of the tuber tissue and may be angular with rays extending to each node (also called eyes, Lee and
638 De Luca, 2019) and is relatively low in starch content (Dean, 1994; Van Denburgh et al., 1986).

639



640

641 **Figure 2.** Internal structure of a potato tuber (from Faridnia et al., 2015[3]).

642 The fact that these GAs are regarded as being important components of plant resistance against pests
643 and pathogens means that breeders work to enhance the GA content in the non-edible parts (Ginzberg
644 et al., 2009). To do so, detailed knowledge of the biosynthetic pathway(s) and the regulation is essential.
645 The major steps in the biosynthesis of α-chaconine and α-solanine have been summarised by Ginzberg
646 et al. (2009) (**Figure 3**). GAs occur in most tissues of the *S. tuberosum* plant, except in the tuber pith
647 (**Table 1**). Concerning the regulation of the biosynthesis and catabolism of the GAs, Sarquis et al.
648 (2000) concluded that this most likely is happening at the tissue or organ level. This is because there is
649 no evidence on their internal transport, and there is a poor correlation between the total content found
650 in the tuber as compared to the foliage of the same cultivar (Sarquis et al., 2000; Ginzberg et al., 2009).

651 The outer layers of the potato tuber (i.e. the periderm), contains the highest concentration of the GAs
652 (**Table 1**), particularly in the 1.5 mm of tissue located under the surface of the tuber (Valkonen et al.,
653 1996). It has been suggested that the biosynthesis of tuber GAs occurs in the phelloderm (Ginzberg et
654 al., 2009), which is the inner part of the potato periderm.

[3] Reprinted from Food research, 77, Faridnia F, Burritt DJ, Bremer PJ and Oey I, Innovative approach to determine the effect pf pulsed electric fields on the microstructure of whole potato tubers: Use of cell viability, microscopic images and ionic leakage measurements, 556-564, 2015, with permission from Elsevier.

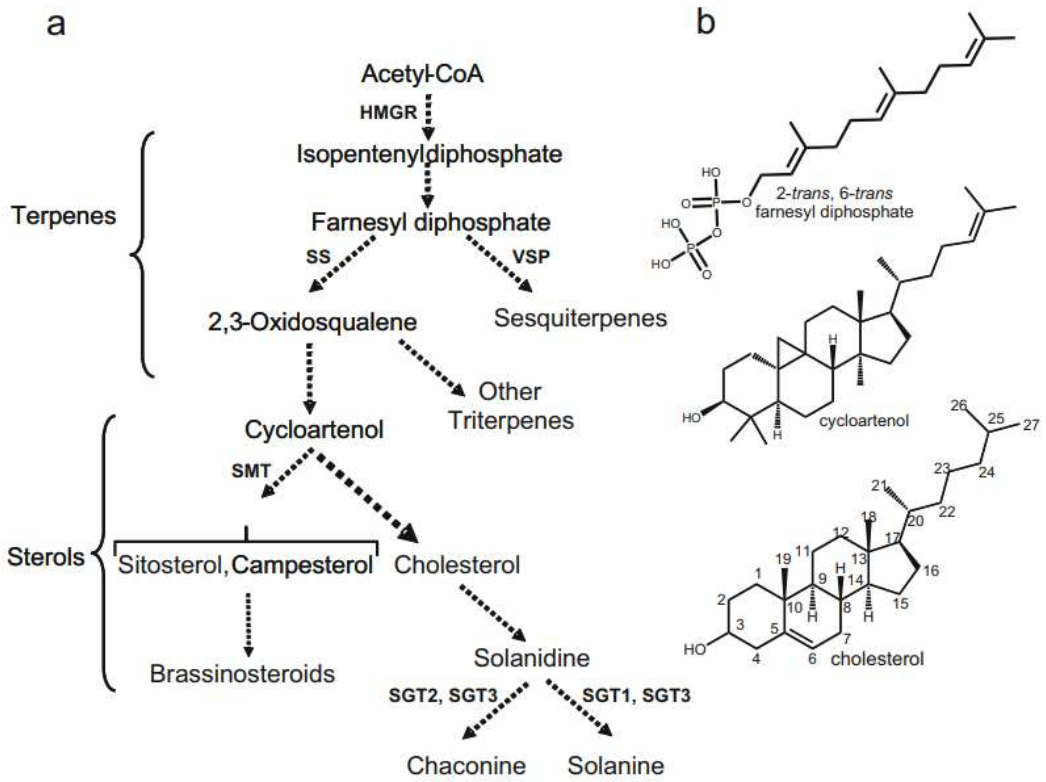


Figure 3. Simplified biosynthetic pathway for steroidal glycoalkaloids (GAs) (from Ginzberg et al., 2009)[4]. (a) The biosynthetic pathway is presented from acetyl-CoA to steroidal GAs with additional pathways included that use the same substrate. *Dashed arrows* indicate multiple enzymatic steps. Terpenes refer to low-molecular weight isoprenoids. (b) Structural formulas for several intermediates of steroidal GA biosynthesis. HMGR: 3-hydroxy-3-methylglutaryl coenzyme A reductase. SS: squalene synthase. VSP: vetispiradiene cyclase. SMT: sterol C24-methyltransferase. SGT: solanidine glycosyltransferase.

[4] Reprinted by permission from Springer: Potato Research, Potato Steroidal Glycoalkaloids: Biosynthesis and Genetic Manipulation, Ginzberg I, Tokuhisa JG and Veilleux RE, 2009.

660 Some reviews, especially in earlier times, have reported relatively small variations for the ratio between
661 the concentrations of α-chaconine and α-solanine in potatoes (peel and flesh). However, later
662 investigations have shown that this ratio may differ to a very great extent, depending on the cultivar as
663 well as the part of the potato. Jin et al. (2018) analysed three different cultivars ('Shepody', 'Russet
664 Burbank' and 'Atlantic') and found the ratio α-chaconine:α-solanine in the whole potato tuber to differ
665 from 0.41 ('Russet Burbank'), to 1.36 ('Shepody') and 3.61 ('Atlantic'). Romanucci et al. (2016)
666 investigated 15 cultivars and reported even higher ratio spans, e.g. in freshly harvested tubers the ratio
667 α-chaconine:α-solanine in the peel varied from <0.06 to 5.7. After 20 days of storage the ratio in the
668 peel was still highly variable, but a general trend towards an increased content of α-chaconine was
669 noted.

670 Several factors have been studied as to whether they could influence the formation of GAs in potatoes.
671 Pre-harvest, factors such as the potato cultivar, the tuber size, climate, damage of the plants by insects,
672 the soil composition, the use of fertilisers, and growth conditions have been reported to potentially
673 influence the content of GAs (e.g. Morris and Petermann, 1985; Gosselin et al., 1988; Mondy and
674 Munshi, 1990; Hlywka et al., 1994; Love et al., 1994; Friedman and McDonald, 1997; Bejarano et al.,
675 2000; Zarzecka and Gugala, 2007; Najm et al., 2012). Some of these factors, such as the potato cultivar,
676 location and farming system, are further discussed in **Section 3.2.2.1.1**.

677 Processing and other post-harvest factors that affect the content of GAs in potatoes are described in
678 **Section 3.2.3**.

679 1.3.3.2. Tomatoes

680 Besides being consumed fresh, tomatoes are used in many processed foods such as canned and sun-
681 dried tomatoes, juices, ketchup, pastes, purees, salads, sauces and soups (Friedman, 2002). In 2016
682 worldwide production of tomatoes was 165 million tonnes (FAOSTAT, 2016). Within the EU, Italy (6.4
683 million tonnes in 2016) and Spain (4.7 million tonnes) are the most important producers, followed by
684 Portugal (1.7 million tonnes) and Greece (1.0 million tonnes). In Europe, tomatoes are grown for the
685 fresh market as well as for processing (tomato paste, juice, peeled tomatoes). Italy and Spain are major
686 producers of processed tomato products.

687 The two most important GAs found in tomato (*S. lycopersicum*) are α-tomatine and α-dehydrotomatine,
688 both of which are found in all parts of the tomato plant (Friedman, 2002). The first discovered GA, the
689 fungitoxic and insect repellent α-tomatine, has been shown to gradually disappear from the fruit with
690 the appearance of lycopene (Sander, 1956, 1958). In addition to α-tomatine and α-dehydrotomatine, a
691 variety of additional GAs have been identified in tomato fruits, including (dehydro)lycopersoside A to H,
692 (dehydro)esculeoside A, B1, B2 and tomatoside A (Moco et al., 2007; Baldina et al., 2016). Some of
693 these other GAs may be present, particularly in the ripe fruits, in higher concentrations than the marker
694 compound α-tomatine (Baldina et al., 2016; Iijima et al., 2009).

695 An example for the variations seen for the two most abundant GAs in the cultivated tomato plant parts
696 is provided by Milner et al. (2011) (**Table 2**). From **Table 2** it follows that the relative α-
697 dehydrotomatine content varies, depending on the plant part, from 6% to 25%. In the (unripe) fruits
698 α-dehydrotomatine is present at approximately 10-fold lower concentrations than α-tomatine.

699 As for the potato plant (see above), the biosynthesis of the tomato GAs (α-tomatine and α-
700 dehydrotomatine) is under separate genetic control in each plant part (Kozukue et al., 2004).

701 **Table 2.** Glycoalkaloid (GAs) concentration (mg/kg fresh weight (fw)) and relative composition of
702 parts of the cultivated tomato (*S. lycopersicum*) plant (Friedman, 2002).

Tomato plant part	α-tomatine (mg/kg fw)	α-dehydrotomatine (mg/kg fw)	α-tomatine (% of total)	α-dehydrotomatine (% of total)
Large immature green fruit	144	14	91	9
Small immature green fruit	465	54	90	10
Root	118	33	77	23
Calyxes	795	62	93	7
Leaves	975	71	93	7
Small stems	896	138	87	13
Large stems	465	142	75	25
Flowers	1,100	190	85	15
Senescent leaves	4,900	330	94	6

703 fw: fresh weight.

704 The TGA concentration in tomato depends, as for potato, on the cultivar and the fruit ripening stage.
705 Eltayeb and Roddick (1984) studied the changes in the α-tomatine content as a result of the fruit growth
706 (e.g. given by the diameter of the berry) and/or the stage of ripening (e.g. defined by the development
707 of the fruit colour: from dark green to full red or yellow colour depending on the cultivar), in
708 commercially grown tomato cultivars and mutants with different ripening characteristics. For three
709 normal-ripening cultivars developing red-, orange- and yellow-ripe fruits, respectively, it was found that
710 the highest concentrations occurred in very young green fruits of around 4 days of age (3,100–4,400
711 mg α-tomatine/kg fw). In subsequent stages of fruit development, the concentration dropped to around
712 700 mg α-tomatine/kg fw at day 10. In the same period the fruit diameter increased from 7 mm at day
713 4 to 20 mm at day 20. By 30 days, when the tomato was approximately 30 mm in diameter, 20 g in
714 weight and still green, α-tomatine concentrations for the three cultivars had decreased to 3–6 mg/kg
715 fw (Eltayeb and Roddick, 1984). Similar results were obtained by Kozukue and Friedman (2003), who
716 reported an inverse relationship between fruit weight and α-tomatine content in green 'Momotaro'
717 tomatoes, with α-tomatine concentrations of 900 mg/kg fw at 1.5–1.9 g of fruit weight, and of 150
718 mg/kg fw at 218.0–220.5 g of fruit weight. Choi et al. (2010) studied the α-tomatine and α-
719 dehydrotomatine content of the tomato cultivar 'Doturakworld' for 60 days, discerning 11 stages of
720 ripeness. The concentrations of α-tomatine and α-dehydrotomatine were highest during stage 1 at day
721 3, amounting to, respectively, 3,600 and 480 mg/kg fw. At growth stage 3 (14 days), α-tomatine and
722 α-dehydrotomatine concentrations had dropped to, 320 and 28 mg/kg fw, respectively. A steady but
723 modest reduction was observed up to stage 7 (47 days), when the tomatoes were full grown, but still
724 green (α-tomatine and α-dehydrotomatine concentrations, 140 and 15 mg/kg fw respectively). When
725 ripening set in at stage 8 (52 days), α-tomatine and α-dehydrotomatine concentrations had dropped
726 below the LOQ (level not indicated) (Choi et al., 2010). It has been shown that during the later ripening
727 stages α-tomatine is converted into esculeoside A and related glycosides (Nohara et al., 2010; Iijima et
728 al., 2009; Fujiwara et al., 2004). The conversion of α-tomatine into esculeoside A is correlated with
729 enhanced levels of ethylene (Iijima et al., 2009). A recent study of Tamasi et al. (2019) revealed that
730 α-tomatine and dehydrotomatidine are primarily located in the locular gel of the tomato fruits, with
731 much lower concentrations in the skin and almost no TGAs present in the pulp.

732 For further details on the concentrations of α-tomatine in tomato and tomato products reported in the
733 literature, see **Section 3.2.2**.

734 #### 1.3.3.3. Aubergine

735 The name aubergine refers to three crops from the genus *Solanum* subgenus *Leptostemonum*
736 (Vorontsova et al., 2013). The three cultivated aubergine species are the Brinjal aubergine (*S.*

737 *melongena* L.), the scarlet aubergine (*S. aethiopicum* L.) and the ghoma aubergine (*S. macrocarpon*).
738 In this opinion the term aubergine alone refers to *S. melongena*, unless stated otherwise.

739 The aubergine is generally cultivated in most tropical and subtropical regions of the world. The two
740 other species (scarlet and ghoma) are less known and mostly grown in sub-Saharan Africa (Taher et
741 al., 2017), not only for their edible fruits, but also for the young leaves (Caruso et al., 2017). The
742 worldwide production of aubergines was 50 million tonnes in 2016 (FAOSTAT, 2018b). Production in
743 the EU is relatively modest: around 0.75 million tonnes were produced in 2016, with Italy being the
744 largest producer (300 ktonnes), followed by Romania and Spain (around 120 ktonnes).

745 The plants are all berry-producing vegetables. The aubergine complex shows a large morphological
746 diversity, including small fruited spiny plants to large fruited non-spiny plants. The long history of
747 breeding has furthermore resulted in a wide range of cultivars with edible fruits of different sizes, shapes
748 and colours (Caruso et al., 2017). Cultivars of aubergine are conventionally grouped into 'Occidental'
749 and 'Oriental'. The first are preferred in North Africa, Europe and the Americas, while the latter mostly
750 are grown in East and South East Asia. They vary from one another with respect to their overall plant
751 morphology and physiology, but also concerning their fruit size, colour and shape (Ceriola et al., 2013).
752 Grafting on rootstocks of related Solanum species (e.g. *S. torvum*, *S. aethiopicum*, *S. macrocarpon*) or
753 tomato (*S. lycopersicum*) hybrids, has become an important area of research to improve the overall
754 quality of the fruits and to increase resistance to soilborne diseases and abiotic stresses (Mennella et
755 al., 2012; Sabatino et al., 2018).

756 For food purposes, aubergines are harvested at an immature stage and have a short storage period.
757 Depending on the cultivar the colour at the unripe stage may vary indeed very much, while the
758 physiologically ripe fruits are brown or yellow (Caruso et al., 2017).

759 Together with α-solamargine, α-solasonine makes up the major content of GAs in this plant species.
760 Both have the steroid solasodine as their aglycone, however, the sugar part in α-solasonine is the
761 trisaccharide solatriose where that of α-solamargine is the trisaccharide chacotriose (see **Figure 1**).
762 While the biosynthesis of α-solasonine starts with the attachment of a D-galactose to the aglycone, the
763 biosynthesis of α-solamargine is initiated with the attachment of a D-glucose. Hence, the expression of
764 solasodine galactosyltransferase 1 (SGT1) specifically reflects the biosynthesis of the GA α-solasonine
765 (Bagheri et al., 2017).

766 Mennella et al. (2012) studied the TGA content (α-solasonine and α-solamargine) in a total of 10
767 different genotypes from two different typologies (D3R and NAS), in commercially unripe, ripe and
768 overripe (physiologically ripe) fruits. In the commercially unripe and ripe stages, the TGA content was
769 relatively low (mean content: 125 and 230 mg/kg dw, respectively), while in the commercially overripe
770 stage a strong increase of the TGA content (mean 1,937 mg/kg dw) was observed. The average ratio
771 between α-solasonine and α-solamargine was approximately 1:2 in the commercially unripe and ripe
772 stage, but changed to 1:1 in the overripe stage (Mennella et al., 2012).

773 Bagheri et al. (2017) compared two different aubergine genotypes, of which one (D1) had 'sweet' fruits
774 and the other (J10) had 'bitter' fruits, with regard to the content of α-solasonine (α-solamargine was
775 not analysed in this study) as well as concerning the expression of SGT1 in the leaves, flower buds,
776 young fruits, mature fruits and physiologically ripe fruits. The levels of α-solasonine closely followed the
777 expression of the SGT1 gene in the same plant tissues. In all tissues the concentration of α-solasonine
778 in the bitter variety was between two and three times higher than in the sweet variety. Concentrations
779 of α-solasonine reported for the D1 and J10 genotypes, respectively, were leaves: 113.3 and 249.3
780 mg/kg dw; flower buds: 135.6 and 478.8 mg/kg dw; young fruits: 61.3 and 160.4 mg/kg dw; mature
781 fruits: 21.6 and 45.6 mg/kg dw and physiologically ripe fruits: 74.7 and 185.0 mg/kg dw (Bagheri et al.,
782 2017).

1.3.4. Previous risk assessments

Previous human risk assessments

The general statement, that a level of 200 mg TGA/kg of unpeeled, uncooked potatoes could be considered as safe, is often found in the scientific literature and can be traced back for decades. Bömer and Mattis (1924) were probably the first to conclude that this TGA level provided sufficient protection to the consumer. They based their statement on the results of a study they conducted on batches of German potatoes from the 1922 harvest that was exceptionally high in solanine content. Batches of potatoes with a TGA content of 257 to 583 mg/kg potatoes reportedly caused intoxications or other health problems, while these effects were not reported for potatoes with levels < 200 mg/kg (concentrations determined with a gravimetric method).

The toxicity of α-solanine and α-chaconine was assessed by the Joint Food and Agricultural Organization/World Health Organisation (FAO/WHO) Expert Committee on Food Additives (JECFA) (JECFA, 1992, 1993). JECFA evaluated the kinetic aspects (absorption, distribution, biotransformation and excretion), toxicity and teratogenicity, primarily in rodents and humans, as available at the time of the evaluation. JECFA concluded that with the available experimental and epidemiological data a safe level of intake could not be determined. The Committee reported that the occurrence of α-solanine and α-chaconine in potatoes is about 20–100 mg/kg, but the levels can increase due to mechanical damage, blight, sprouting, processing and storage. JECFA considered that daily consumption of potatoes containing normal GA levels was of no concern, provided that the potatoes were properly handled and grown.

In 1990, the Swedish National Food Administration carried out an 'Assessment of Health-risks Related to Glycoalkaloids ('Solanine') in Potatoes: A Nordic View', and concluded that "*Based on the present knowledge, a TGA concentration of 200 mg/kg potato for the potato varieties currently available on the Nordic Market appears to be the maximum level which can be accepted. However, because of the small safety margin, efforts should be made to reduce the levels of GA'.* They recommended that the average TGA concentration in new potato varieties should not exceed 100 mg/kg (Nordic Working Group on Food Toxicology and Risk Assessment, 1990).

In 2018, and as a result of an intoxication of the members of a family in Baden-Württemberg (Germany) from potatoes that occurred in 2015, the German Federal Institute for Risk Assessment (BfR) published an assessment on the acute toxicity of potato GAs (BfR, 2018a,b). Jacket potatoes and potatoes boiled in the skin had been consumed before symptoms such as stomach-ache and vomiting had developed. Analytical results showed that the potatoes contained 236 mg TGAs/kg (141 mg α-solanine and 95 mg α-chaconine per kg). Based on available human data in the literature, BfR identified a LOAEL of 1 mg GAs/kg body weight (bw) per day and extrapolated from this by applying an uncertainty factor of 2 to a NOAEL of 0.5 mg GAs/kg bw per day. This uncertainty factor of 2 was considered by BfR to also protect persons with higher susceptibility. In addition, BfR recommended that the intake of GAs should be below the NOAEL of 0.5 mg/kg bw per day to ensure a 'margin of safety' to the NOAEL being >1. To avoid an exceedance of this NOAEL, BfR recommended that the TGA content of potatoes should not be higher than 100 mg/kg fresh weight. With this recommendation, particularly sensitive populations are covered too. In view of significant data gaps, BfR considered its conclusions as preliminary. Since case reports in humans indicate a lethal dose of 3–6 mg/kg bw, whereas the LD_{50} for mice and rats is at least 300-fold higher, BfR pointed to the considerable higher sensitivity of humans compared to rodents.

Previous risk assessments on farm and companion animals

No risk assessments on farm and companion animals have been identified.

1.3.5. Legislation and other standards

In order to protect public health, Article 2 of the Council Regulation (EEC) No 315/93[5] stipulates that, where necessary, maximum tolerances for specific contaminants shall be established. Thus, a number of maximum tolerances for contaminants as well as natural plant toxicants are currently laid down in Commission Regulation (EC) No 1881/2006[6]. However, no maximum levels for GAs in food or feed have been established at EU level under this or under another Regulation.

Some European countries have national legislation or recommendations on the maximum limits of TGAs, mainly in potato and potato products. Hungary has a national regulation of 100 mg/kg as the maximum limit of solanine equivalents of raw, unpeeled potatoes. In Finland, a maximum level for GAs (solanine glycosides, such as α-solanine and α-chaconine) for potatoes of 200 mg/kg[7] exists, as well as in Sweden, where the National Food Administration's Regulations on Certain Foreign Substances in Food [8] established in 2004 a maximum content of TGAs (total content of solanidine GAs, such as α-chaconine and α-solanine) in potatoes, raw and unpeeled, of 200 mg/kg. Also Denmark uses the Nordic guideline of 200 mg GAs/kg for known potatoes varieties, and 100 mg/kg for new potatoes varieties.

In the Netherlands, potato producers agreed to observe a limit of 100 mg/kg fresh weight for new potato cultivars (Bal, 1989, as cited in Essers et al., 1998). In Austria, no maximum levels is laid down, but in the Regulation for marketing of food potatoes there are minimum requirements for marketing of food potatoes to avoid a high content of GAs, such as the potatoes (i) must be free of noticeable green tubers, (ii) for class I potatoes: a slight green colour of not more than 1/8 of the surface is allowed, and (iii) for class II potatoes: a slight green colour, which can be removed by normal peeling is allowed.

In Germany, BfR recommended that the GA content of potatoes should not be higher than 100 mg/kg fresh weight (BfR, 2018a,b).

In Canada, a maximum level of 200 mg/kg for the sum of α-solanine and α-chaconine in fresh potato tubers was established, on the basis that most reported cases of adverse effects associated with potato GA exposure have occurred at concentrations above this value. This value was confirmed in 2014 [9], indicating that it is achievable when good agricultural, manufacturing and storage practices are followed [10]. In the USA, a maximum acceptable TGA content of potato tubers of 20 to 25 mg per 100 g of fresh potato (equivalent to 200 to 250 mg/kg) has been set according to the US-Food and Drug Administration (FDA) poisonous Plant Database [11].

The Organisation for Economic Co-operation and Development (OECD) published in 2002 its 'Consensus Document on Compositional Considerations for New Varieties of Potatoes: Key Food and Feed Nutrients, Anti-Nutrients and Toxicants' [12]. Under the considerations for the assessment of new potato varieties, it indicated that the comparison of the chemical composition of tubers from a modified variety with tubers from the non-modified comparator should include, among others, the determination of GAs. It reported on the '*widely accepted safety limit for the level of TGA in tubers*' of 200 mg/kg fresh weight

[5] Council Regulation (EEC) No 315/93 of 8 February 1993 laying down Community procedures for contaminants in food. *OJ L 37, 13.2.1993, p. 1–3.*
[6] Commission Regulation (EC) No 1881/2006 of 19 December 2006 setting maximum levels for certain contaminants in foodstuffs. *OJ L 364, 20.12.2006, p. 5–24.*
[7] https://www.finlex.fi/fi/laki/alkup/2016/20160880
[8] Föreskrifter om ändring i Livsmedelsverkets föreskrifter (SLVFS 1993:36) om vissa främmande ämnen i livsmedel, LIVSFS 2002:16https://www.livsmedelsverket.se/globalassets/om-oss/lagstiftning/nummerordning---upphord-lagstiftning/2004/livsfs-2004-7-frammande-amnen.pdf
[9] https://www.canada.ca/content/dam/hc-sc/documents/services/food-nutrition/public-involvement-partnerships/proposal-update-maximum-levels-glycoalkaloids-potato-tubers-eng.pdf
[10] In 2017 Health Canada proposed to transfer the existing maximum level for TGA in potato tubers from the List of Maximum Levels for Various Chemical Contaminants in Food to the regulatory List of Contaminants and Other Adultering Substances in Food, thereby removing this maximum level from the first list.
[11] https://www.accessdata.fda.gov/scripts/Plantox/Detail.CFM?ID=6537
[12] https://www.oecd.org/science/biotrack/46815167.pdf

(based on Bömer and Mattis (1924) and Smith et al. (1996)), and concluded that if the analyses of GAs (and other parameters) indicate that a novel variety is within the ranges given in the literature it can be considered equivalent with respect to its overall composition.

2. Data and Methodologies

The CONTAM Panel applied the general principles for the assessment of chemicals in food as described by WHO/International Programe on Chemical Safety (IPCS) (WHO/IPCS, 2009). In addition, EFSA guidances pertaining to risk assessment were followed for the development of the risk assessment when necessary (EFSA, 2005, 2007, 2009, 2010a,b, 2011a,b; EFSA SC, 2011, 2012a,b, 2017a,b,c,d).

2.1. Methodology for data collection, selection of evidence and study appraisal

EFSA outsourced a call for an 'Extensive literature search and selection for relevance of studies related to the chemistry and toxicity of GAs and quinolizidine alkaloids in food and feed' aiming at identifying and evaluating literature related to the present assignment (and to another mandate of the CONTAM Panel on quinolizidine alkaloids which is not further considered here). The call was launched as a reopening competition for a specific contract under multiple framework contract CT/EFSA/AMU/2014/01 Lot 2. The University of Chemistry and Technology Prague was awarded with the contract and a final project report was delivered in November 2017 and was published (University of Chemistry and Technology Prague, 2019). The aim of the assignment was to identify and collect all relevant literature regarding GAs (and quinolizidine alkaloids) in food and feed covering the following areas: (i) chemistry and analysis, (ii) toxicokinetics in experimental animals, farm/companion animals, humans, and from in vitro studies and on biomarkers, (iii) *in vitro* and *in vivo* mode of action of toxicity, (iv) toxicity *in vivo* in laboratory animals, (v) toxicity *in vitro*, (vi) observations in humans, including epidemiological studies, case reports, outbreaks, intervention studies, (vii) adverse effects in farm and companion animals, (viii) occurrence in food, (ix) occurrence in feed and animal exposure and (x) influence of cultivars, maturity, environmental conditions, storage and processing in the concentrations of the alkaloids in the products. Details of the methodology and the results are reported in University of Chemistry and Technology Prague (2019).

In addition to the literature searches outsourced by EFSA, additional complementary searches were performed. A literature search for studies regarding observations in humans was performed in October 2017 (for details see **Appendix B**, **Section B.1**). An extra search to identify studies on the toxicity of GAs was conducted in April 2018 (for details see **Appendix B**, **Section B.2**). The literature searches were performed using Web of Science[13], PubMed[14] and Scopus. The references resulting from the literature search were imported and saved using a software package (EndNote[15]), which allows effective management of references and citations.

Reviews, relevant scientific evaluations and toxicity studies by national or international bodies were also considered for the current risk assessment, i.e. JECFA (JECFA, 1992, 1993). When relevant papers were identified during the risk assessment process (e.g. from other studies or reviews), they were also considered.

[13] Web of Science (WoS), formerly ISI Web of Knowledge, Thomson Reuters. Available at: http://thomsonreuters.com/thomson-reuters-web-of-science/

[14] PubMed, Entrez Global Query Cross-Database Search System, National Center for Biotechnology Information (NCBI), National Library of Medicine (NLM), Department of the National Institutes of Health (NIH), United States Department of Health and Human Services. Available at: http://www.ncbi.nlm.nih.gov/pubmed/

[15] EndNote X5, Thomson Reuters. Available at: http://endnote.com/

In addition, a search for recent reviews was conducted to identify scientific publications dealing with methods of analysis, chemistry, formation in food, exposure and occurrence of GAs in food and feed.

A search for original studies reporting on the occurrence of GAs in food and feed samples was carried out, since it was expected that only limited number of occurrence data of GAs in food and feed were going to be submitted to EFSA (see **Section 2.3.1**),

An update of the literature available in the public domain was done regularly, on 17 December 2018, 3 May 2019 and 14 August 2019. Since that date, the literature was monitored to identify studies relevant for the risk assessment until the time of endorsement.

Any publications were used in the assessment if they were considered as relevant by applying expert judgement. In addition, for the studies on the toxicity of GAs in experimental animals and farm and companion animals the selection criteria in **Table 3** were used. For the sections on repeated dose toxicity and developmental and reproductive effects, animal studies based on i.p. or i.v. application of GAs or aglycones were excluded (see also **Appendix G**).

Table 3. Selection criteria for the studies on the toxicity of glycolkaloids (GAs) in experimental animals and in farm animals, horses and companion animals.

Inclusion criteria:	Exclusion criteria:
• Data described by the author as purified GAs or GA mixtures, and their aglycones. • Edible parts of the plant, as well as their preparations in which the total and/or GA concentration is indicated. • Preparations of potato sprouts in which the total and/or GA concentration is indicated.	• Studies on non-edible parts (e.g. flowers) of the plants. • Studies giving no adequate information on the composition and/or TGA concentration of the used extracts, materials or preparations.

2.2. Food and feed occurrence data submitted to EFSA

2.2.1. Data collection and validation

Following a mandate from the European Commission to EFSA, a call for annual collection of chemical contaminant occurrence data in food, was issued by the former EFSA Dietary and Chemical Monitoring Unit (now DATA Unit)[16] in December 2010 with a closing date of 1st October of each year[17]. European national authorities and similar bodies, research institutions, academia, food business operators and other stakeholders were invited to submit analytical data on GAs in food through this call. The data submitted to EFSA on GAs were provided by national authorities from Germany, Sweden and the Netherlands.

The data submission to EFSA followed the requirements of the EFSA Guidance on Standard Sample Description for Food and Feed (EFSA, 2010a), and the occurrence data were managed following the EFSA standard operational procedures (SOPs) on 'Data collection and validation' and on 'Data analysis of food consumption and occurrence data'.

By the 2nd of September 2019, a total of 1,300 analytical results on GAs in food (representing 651 food samples) were available in the EFSA database. Data received after that date were not included in the dataset used for further evaluation for this opinion. All data submitted to EFSA concerned potato and potato products. No data were submitted concerning GAs in tomato, aubergine and food products containing tomato or aubergine.

[16] From 1 January 2014 onwards, Evidence Management Unit (DATA).
[17] Current call: https://www.efsa.europa.eu/en/consultations/call/190410

In addition, in January-February 2019, a consultation was held with registered EFSA Stakeholders and other stakeholders to inform the work of the CONTAM Panel and its working group in developing this scientific opinion. The aim of the consultation was to collect information which were not readily available in the public domain regarding GAs occurrence data in food and feed, factors that influence the content of GAs and toxicity (see Section on Documentation provided to EFSA). Following this consultation, EFSA invited the stakeholders to submit occurrence data in food and feed within the annual collection of chemical contaminant occurrence data, following the requirements of the EFSA Guidance on Standard Sample Description for Food and Feed (EFSA, 2010a) for inclusion in the risk assessment. As a result, on the 2nd of July 2019 the European Starch Industry Association submitted occurrence data to EFSA concerning 1,728 samples on dietary fibre and potato proteins, pulp, juice, starch and on potatoes used for starch production. See **Section 3.2.1** for further details.

2.2.2. Data analysis

Following the EFSA SOP on 'Data analysis of food consumption and occurrence data' to guarantee an appropriate quality of the data used in the exposure assessment, the initial dataset was carefully evaluated applying several data cleaning and validation steps. Special attention was paid to different parameters such as 'Sampling strategy', 'Reporting unit' and the codification of the different samples under FoodEx classification. The outcome of the data analysis is presented in **Section 3.1.1**.

The left-censored data (results below LOD or below LOQ) were treated by the substitution method as recommended in the 'Principles and Methods for the Risk Assessment of Chemicals in Food' (WHO/IPCS, 2009). The same method is indicated in the EFSA Scientific Report 'Management of left-censored data in dietary exposure assessment of chemical substances' (EFSA, 2010b) as an option in the treatment of left-censored data. The guidance suggests that the lower bound (LB) and upper bound (UB) approach should be used for chemicals likely to be present in the food (e.g. naturally occurring contaminants, nutrients and mycotoxins). The LB is obtained by assigning a value of zero (minimum possible value) to all samples reported as lower than the LOD (<LOD) or LOQ (<LOQ). The UB is obtained by assigning the numerical value of LOD to values reported as <LOD and LOQ to values reported as <LOQ (maximum possible value), depending on whether LOD or LOQ is reported by the laboratory.

2.3. Food and feed consumption data

2.3.1. Food consumption data

The EFSA Comprehensive European Food Consumption Database (Comprehensive Database) provides a compilation of existing national information on food consumption at individual level. Details on how the Comprehensive Database is used are published in the Guidance of EFSA (EFSA, 2011a). Consumption data were collected using single or repeated 24- or 48-h dietary recalls or dietary records covering from three to seven days per subject. Subjects in the Comprehensive Database are classified according to the following sub-populations:

- Infants: < 12 months old
- Toddlers: ≥12 months to < 36 months old
- Other children: ≥36 months to < 10 years old
- Adolescents: ≥10 years to < 18 years old
- Adults: ≥18 years to < 65 years old
- Elderly: ≥65 years to < 75 years old
- Very elderly: ≥75 years old.

Two additional surveys provided information on specific population groups: 'Pregnant women' (≥15 years to ≤45 years old; Latvia) and 'Lactating women' (≥28 years to ≤39 years old; Greece).

Owing to the differences in the methods used for data collection, direct country-to-country comparisons can be misleading. Overall, the food consumption data gathered by EFSA in the Comprehensive Database are the most complete and detailed data currently available in the EU.

Some of these data, however, cannot be used in exposure assessments if the relevant occurrence data are reported at the level of the raw primary commodities (RPCs). The RPC model developed by EFSA in 2019 aimed to bridge this gap by transforming food consumption data from the Comprehensive Database for composite foods (foods consisting of multiple components) and RPC derivatives (single-component foods which have been physically changed by processing) into their corresponding quantities of RPCs. Details on how the RPC database was developed are published in a dedicated technical report (EFSA, 2019). The RPC Consumption Database, based on the Comprehensive Database version published on the 31 March 2018, contains 51 dietary surveys from 23 different countries. These surveys cover a total of 94,532 subjects and 26,573,088 RPC consumption records.

The use of the RPC Consumption Database was considered as the most appropriate approach for the current assessment for which occurrence data in the RPC main-crop potato were available (see **Section 2.5**). As no occurrence data were available for GAs in tomato and aubergine, these foods were not included in the consumption and exposure assessment.

Detailed information on the different dietary surveys used in this report for the acute exposure assessment is shown in **Annex A1**, including the number of subjects and days available for each age class.

2.3.2. Feed consumption data

Of the large number of species within the Solanaceae family, only potatoes and by-products derived from them are used as feed for livestock. Due to the high cost of growing the crop, and the value of potatoes as food for human consumption, they are not usually grown as a feed for livestock, and whole (raw) potatoes are only used as feed when production is in surplus to requirements as food, or when they are discarded because they do not meet market requirements because of their size, appearance, diseases or physical damage. As a result, the use of raw tubers as livestock feed tends to be opportunistic, and the CONTAM Panel have not identified any data on quantities used in this way.

Feed grade potato tubers are mainly used as feeds for ruminants but may also be fed to horses. They are highly palatable and nutritious and dairy cows will typically consume up to 25–30 kg of potatoes (Crawshaw, 2001). For growing cattle, up to 30% of the diet (dry matter) may be fed (Fiems et al., 2013).

Raw potatoes are relatively unpalatable, and the starch indigestible for pigs and poultry, and therefore it is recommended that they are cooked or steamed before being fed (Whittemore, 1977). They may be included at up to 15% of the diet dry matter for finishing pigs and sows (Ewing, 2002), and up to 50% of the dietary dry matter for growing and finishing pigs (Pond and Maner, 1984). Inclusion of cooked potatoes in poultry grower diets at up to 20% have been recommended, while for layers a dietary inclusion rate of 15% potato meal did not reduce egg production (Whittemore, 1977).

In addition to raw potatoes, the EU Catalogue of Feed Materials [18] lists fourteen different by-products from potato processing industries, including cuttings, scrapings, flakes and pulp. The suitability of these as feeds for different livestock depends on the degree of processing to which they have been exposed, and the nutritional requirements of the livestock. Steamed potato peel is a highly nutritious and palatable feed for pigs, and its availability has increased in recent years in line with the increase in the production of French fries. Low-GA potato protein concentrate is often added to low protein pig starter diets to

[18] EU Regulation 68/2013

reduce the incidence of diarrhoea in newly weaned pigs (Stein, 2002). The use of potato protein concentrate in poultry diets is generally limited, with some research reporting an increase in necrotic enteritis in broilers. Potato protein concentrate was used as an alternative to fishmeal in the diets of rainbow trout, with limited success (Tusche et al., 2011), and has been included in specialist feeds for dogs [19]. However, information on amounts of this or other potato by-products used as animal feeds within the EU is not publicly available. Since the highest concentrations of GAs are found in the skin (Milner et al., 2011), and these are removed at an early stage in the processing, levels of GAs in by-products such as potato starch and potato protein concentrate are likely to be low.

2.4. Food classification

Consumption data were classified according to the FoodEx classification system (EFSA, 2011b). FoodEx is a food classification system developed by EFSA in 2009 with the objective of simplifying the linkage between occurrence and food consumption data when assessing the exposure to hazardous substances. It contains 20 main food categories (first level), which are further divided into subgroups having 140 items at the second level, 1,261 items at the third level and reaching about 1,800 end-points (food names or generic food names) at the fourth level.

In 2011, a new version of FoodEx, named FoodEx2 has been developed and is described in the scientific document "Report on the development of a Food Classification and Description System for exposure assessment and guidance on its implementation and use" (EFSA, 2011b). The last release of FoodEx2 complements the previous hierarchical classification system of basic codes with more detailed food levels and gives the possibility of reporting additional information through the use of facets and facet descriptors (EFSA, 2015).

For this opinion the amount consumed for each reported FoodEx code were converted to the equivalent amount of RPC (main-crop potatoes) by means of the RPC model (EFSA, 2019).

2.5. Methodology for Exposure assessment

Human exposure assessment

The CONTAM Panel estimated the acute exposure to GAs (see **Section 3.1.6**).

Since available occurrence data did not cover all the food categories containing potatoes in the Consumption Database (see **Section 3.2.1**), it was decided that the best approach for the exposure assessment would be to use the occurrence values in the RPC (main-crop potatoes and new potatoes) and the RPC Consumption Database (see **Section 2.3.1**).

The acute dietary exposure to GAs was estimated by using a probabilistic approach (EFSA, 2011b). Acute dietary exposure to GAs was calculated on a per day basis. Acute exposure was assessed for each reporting day by multiplying the total daily consumption amount of main-crop potatoes by one occurrence level randomly drawn among the individual results available. For the calculations of exposure, the UB values for the occurrence were used as considered more adequate to represent acute exposure (in this assessment the LB and UB of the analysed samples do not differ substantially as only two analytical results for α-solanine and 10 results for α-chaconine are left censored in the final dataset).

Since peeling of potatoes and heat processing decrease the content of GAs (see **Section 3.2.4**), the CONTAM Panel applied reductions factors to estimate the GA levels in the final food consumed.

Information about the peeling was not available in the RPC Consumption Database, thus the CONTAM Panel assumed that in 90% of all eating events potatoes were consumed peeled. A probabilistic

[19] Source: www.4petsusa.com

approach was used to assign to each eating event the 'peeled' vs 'unpeeled' status. Based on studies in the literature (see **Section 3.2.4**), a reduction factor between 0.25 and 0.75 was attributed to the peeling of potatoes.

Information about the heat processing methods was only available in a limited number of cases. A probabilistic approach was also used to assign to each eating event one of the heat processing methods, based on the proportion of each cooking method within the events for which this information was available (frying: 23%, deep frying: 12%, cooking in water: 47%, drying, flaking and roasting: 5% each, and stewing: 2%). Based on studies in the literature (see **Section 3.2.4**), reduction factors between 0.2 and 0.9 were attributed to frying and deep frying, and between 0.05 and 0.65 for all other cooking methods.

The reduction factors for peeling and heat processing assigned to each eating event were extracted from a normal distribution, where the minimum and maximum allowed value were the ranges described above. Results were compared with an exposure assessment scenario where reduction factors were extracted from an uniform distribution rather than a normal distribution and the exposure outcome was very similar (see **ANNEX A**).

Exposures from the different eating events of one day (for the considered subject) were summed and divided by the individual's body weight to obtain the total exposure per kg bw per day.

Consumption of main-crop potatoes linked to the consumption of alcholic beverages (vodka and spirits) was not taken into consideration as the CONTAM Panel considered the transfer of GAs from the potatoes during the distillation and refining process to be negligible.

The mean daily acute UB exposure was then calculated per survey and age class.

This process was iterated 1,000 times. The mean of the 1,000 mean daily acute UB exposures per survey and per age class was then calculated. The 95% confidence interval (CI) defined as the interval between the 2.5th and 97.5th percentiles obtained from the 1,000 iterations was also determined to indicate the uncertainty around the mean value.

In **Table A1** of **Annex A** the number of available subjects for each age class used in the acute exposure assessment is provided.

Animal exposure assessment

Due to insufficient data on levels of GAs in either potatoes or potato by-products used as animal feeds, and lack of data with which to calculate reference points for any of the farm animal species (see **Section 3.1.4** and **3.1.7**), no estimates of exposure to GAs have been made.

2.6. Methodology for Risk characterisation

The CONTAM Panel applied the general principles of the risk characterisation process for chemicals in food as described by WHO/IPCS (2009) and the relevant EFSA guidance documents.

3. Assessment

3.1. Hazard identification and characterisation

3.1.1. Toxicokinetics

3.1.1.1. Experimental animals

The studies identified on the toxicokinetics of GAs in experimental animals are described below, and details are given in **Appendix C**.

3.1.1.1.1. α-Solanine

Nishie et al. (1971) studied the metabolic fate and distribution of α-solanine in male Fisher rats. [^{3}H]-α-solanine was administered by gavage at 5 mg/kg bw and i.p. injection at 5–25 mg/kg bw to an unreported number of rats. Following oral exposure, fecal elimination of radioactivity reached 84% of the administered dose by day 4, with urinary excretion accounting for less than 10%. 24 h after the oral exposure, radioactivity distributed mainly in the liver (about 1.5% of the administered dose), followed by blood, kidney and lungs. Following i.p. injection of 5–15 mg/kg bw of labelled α-solanine, radioactivity was eliminated at similar rates in urine and feces (approximately 15–20% of the administered doses by 24 h). Decreased urinary excretion and complete absence of fecal elimination were observed at the highest dose administered (25 mg/kg bw), with a corresponding increase in the radioactivity retention in the gastrointestinal tract, similarly to what reported by Norred et al. (1976) for α-chaconine (see **Section 3.1.1.1.2**). Lesions of increasing severity were observed in the peritoneal cavities of animals exposed to ≥10 mg/kg bw. A metabolic study was performed using TLC. Following oral exposure, 65% of the radioactivity present in feces was identified as solanidine, whereas only 6% of the dose excreted in urine was identified as solanidine, with two other non-identified metabolites accounting for 80% and 13% of the radioactivity excreted in urine.

Groen et al. (1993) studied the toxicokinetics of [^{3}H]-α-solanine in male rats and hamsters. [^{3}H]-labelled α-solanine was administered via i.v. injection or via gavage at a dose of 54 and 170 µg/kg bw, respectively. Following i.v. administration a similar toxicokinetics of radioactivity in blood and plasma was observed in rats and hamsters. Plasma levels of total radioactivity and unchanged α-solanine showed three-phase profiles (see **Table 4**). Compared to i.v. treated animals, the oral bioavailability of α-solanine and metabolites (based on total radioactivity) in animals treated by gavage was 29% and 57% in rats and hamsters, respectively. The oral bioavailability for the parent compound α-solanine was 1.6% and 3.2% in rats and hamsters, respectively. Plasma clearance of unchanged α-solanine following oral administration was significantly faster in rats ($t_{1/2}$ = 7.79 ± 0.83 h) than in hamster ($t_{1/2}$ = 19.7 ± 4.2 h), however similar plasma clearance was observed for total radioactivity in the two species (82.2 ± 6.1 h in rats, and 94 ± 33 h in hamsters). Excretion of total radioactivity in urine and feces was significantly faster in rats than in hamsters treated by gavage. Seven days after the oral treatment, total excretion of the administered dose was 89% in rats, with approximately 3% in urine and 86% in feces; in hamsters, total excretion was 39%, with approximately 10% in urine and 29% in feces. Macroscopic observation performed at the necropsy of the animals sacrificed 7 days after the treatment revealed damages in the intestinal walls of some hamsters following oral treatment.

Table 4. Plasma half-lives (tri-phase elimination) of total radioactivity and unchanged α-solanine following i.v. injection of 54 mg/kg bw [^{3}H]-α-solanine in rats and hamsters (Groen et al., 1993).

		$T_{1/2\alpha}$ (h)	$T_{1/2\beta}$ (h)	$T_{1/2\tau}$ (h)
Total radioactivity	Rat	1.4 ± 1.6	8.9 ± 2.2	61 ± 13
	Hamster	1.5 ± 0.4	7.7 ± 3.0	62 ± 22
Unchanged α-solanine	Rat	1.9 ± 1.4	9.9 ± 0.8	32 ± 6.5
	Hamster	2.1 ± 1.4	8.9 ± 2.8	39 ± 13

3.1.1.1.2. α-Chaconine

Norred et al. (1976) studied the toxicokinetics of [^{3}H]-α-chaconine (tritiated in the aglycone moiety at carbon atoms adjacent to the double bond and to the nitrogen atom) in male Sprague-Dawley rats (200–300 g bw). Rats were dosed either by gavage (5 mg/kg bw) or i.p. injection (5–25 mg/kg bw) and housed in metabolic cages up to at least 100 h after treatment. Following oral administration of 5 mg/kg bw, fecal elimination of 60% and 80% of the administered radioactivity dose was observed after 24 and 48 h, respectively. Urinary excretion reached a plateau of 10% of the administered radioactivity dose by 24 h. Maximum levels in tissues were measured after 6–12 h, with higher levels in liver (about 1.3% of the radioactivity dose) and blood (about 0.2%). The radioactivity recovery considering the sum of fecal elimination, urinary excretion and gastrointestinal content, ranged approximately from 85 to 100% across the selected timepoints. Following i.p. injection of 5–10 mg/kg bw, elimination took place via the urine (10–15% of the administered radioactivity dose excreted 24 h after treatment) and the feces (<10%). However, reduced urinary excretion was observed at the highest dose administered (25 mg/kg bw); complete absence of fecal elimination was also observed at 15 and 25 mg/kg bw with a corresponding increase in the radioactivity detected in the gastrointestinal tract. A metabolic study was performed using TLC, which the CONTAM Panel considers to be of limited value. Following oral treatment with 5 mg/kg bw, about 63% and 16% of the radioactivity present in feces and urine was extracted with ether, respectively. TLC analysis of the feces and urine extracts revealed the presence of a main metabolite with the same retention factor of solanidine. About 25% of radioactivity detected in feces was attributable to the parent compound.

The distribution of radioactivity in subcellular fractions of hepatocytes in female Swiss-Webster mice exposed to α-chaconine was investigated by Sharma et al. (1983). Following oral administration of 10 mg/kg [^{3}H]-α-chaconine, mice were sacrificed at 3, 6, 14, 72 and 120 h after exposure and liver tissue and blood were sampled. The distribution of radioactivity in subcellular fractions of hepatocytes was studied. Higher radioactivity levels in liver homogenate were measured from 3 until 14 h after the exposure, with the nuclear subcellular fraction containing the highest levels, followed by mitochondria and microsomes. In a second experiment, mice were orally exposed to different doses (1, 3 or 10 mg/kg) and sacrificed after 14 h. The radioactivity distribution in liver homogenate and cellular subcellular fractions was observed to increase consistently with the administered dose. Finally, mouse pre-treatment with the cytochrome P450 inducer dibenzo[a]pyrene resulted in an increased radioactivity in all the liver subcellular fractions. However, lower levels were measured in the subcellular fractions, possibly suggesting an increased metabolism of α-chaconine due to enzymatic induction by dibenzo[a]pyrene.

Alozie et al. (1979a) studied the toxicokinetics of [^{3}H]-α-chaconine in male Golden hamsters administered a dose of 10 mg/kg bw by gavage or i.p. injection. Hamsters were sacrificed at regular time intervals (3 animals per time point) up to 168 h after treatment and radioactivity was measured in different tissues. Urine and feces were collected and measured for radioactivity. Subcellular distribution of labelled α-chaconine was also studied in various tissues. After the oral exposure, distribution of

radioactivity in various tissues was observed at the different timepoints, in particular higher levels were observed in lungs, liver, and skeletal muscles. Radioactivity levels remained high in the intestine throughout the oral study and also relatively high in the i.p. study, possibly suggesting high biliary excretion. Following oral exposure, the excretion of radioactivity occurred mainly via the urine reaching about 21% of the administered dose after 168 h, whereas only a minor fraction was detected in feces. The study of subcellular distribution showed α-chaconine or its metabolites to be distributed mainly in the nuclear, microsomal and soluble cellular fractions in various tissues including brain, heart and liver. The residual radioactivity present in the cellular subfractions following a wash procedure with methanol suggested that a fraction of α-chaconine (or α-chaconine metabolites) could bind to macromolecules (type not specified) in these tissues.

In a follow-up publication, Alozie et al. (1979b) studied the metabolic fate of α-chaconine in Golden hamsters following oral exposure to 10 mg/kg bw [^{3}H]- α-chaconine. Urine and feces were collected at different time points up to 168 h after exposure and subsequently extracted with water or chloroform. These extracts were analysed by TLC. In urine, initially unaltered α-chaconine appeared to be the only excreted substance (100% of detected radioactivity at the 12 h timepoint, corresponding to 0.25% of the administered dose), showing a declining trend towards the end of the study. Solanidine appeared in urine 24 h after the exposure and reached a maximum level after 72 h. Four other metabolites could be detected but not identified, two of them representing the major peaks at 168 h. The total excretion in feces represented only a minor amount of the administered dose (see Alozie et al., 1979a) and was mainly constituted by the parent compound.

3.1.1.2. Humans

The studies identified on the toxicokinetics of GAs in humans are described below, and details are given in **Appendix C**.

3.1.1.2.1. Mixtures of α-solanine and α-chaconine

The kinetics of α-solanine and α-chaconine in human volunteers was studied by Hellenäs et al. (1992). Seven healthy male adults were administered a peeled mashed potato meal containing 82 mg/kg α-solanine and 118 mg/kg α-chaconine. The study subjects were given a portion adjusted to reach a dose of 1 mg TGAs/kg bw. Blood was sampled at regular intervals prior the exposure and up to 25 h following the exposure and the content of α-solanine, α-chaconine and solanidine were determined via HPLC analysis. Average serum C_{max} for α-solanine and α-chaconine were 7.7 and 14.4 ng/mL, with average T_{max} of 5.1 and 6 h, respectively. Solanidine reached the maximum level after 8 h (ranging from 1.0 to 4.8 ng/mL across subjects) and remained apparently constant until the end of the study. Average apparent biological half-lives were calculated to be approximately 11 and 19 h for α-solanine and α-chaconine, respectively, with low variability across subjects. The CONTAM Panel noted that the reliability of the calculated half-lives was limited by the short observation period (25 h). Adverse/undesirable effects were observed in most of the study volunteers, as described in **Section 3.1.3**.

Mensinga et al. (2005) studied the pharmacokinetics and toxicity of α-solanine and α-chaconine in 6 male and 8 female healthy volunteers treated with escalating doses of the two GAs either via solutions (0.30, 0.50 or 0.70 mg TGA/kg bw) or in a form of a mashed potato meal (0.95, 1.10 or 1.25 mg total TGA/kg bw). Both the solution and the potato meals contained approximately equal amounts of the two GAs. Blood was sampled from all subjects at various timepoints as from 1 h before treatment till 96 h after treatment and serum GA levels were measured by HPLC analysis. Both α-solanine and α-chaconine showed similar t_{max} in serum ranging from 4 to 8 h without correlation with the administered doses. Maximum serum concentrations (C_{max}) showed strong positive correlations for α-solanine and α-chaconine both for administration via solution (correlation coefficient 0.98) and via potato meal (0.97)

(see **Table 5**). A similar result was obtained for the 0–24 h Areas Under the Curve (AUC_{0-24}), whereas a significant correlation was observed only in subjects treated with mashed potatoes for the AUC from point zero to infinity ($AUC_{0-\infty}$). Serum half-lives showed a high inter-individual variability and were generally longer for α-chaconine (27–84 h, average of 44 h) than for α-solanine (5–42 h, average of 21 h). One volunteer exposed to 1.25 mg/kg bw experienced adverse effects as described in **Section 3.1.3.1.2**.

Table 5. Pharmacokinetic parameters (ranges) of α-solanine and α-chaconine in human volunteers (adapted from Mensinga et al., 2005).

	GAs solution			**GAs in mashed potatoes**		
Dose (mg/kg bw)	0.30	0.50	0.70	0.95	1.10	1.25
Number of subjects	2	2	2	3	3	2
			α-Chaconine			
C_{max} (µg/L)	2.8–3.2	3.2–4.5	3.3–6.3	5.2–7.4	2.9–7.8	6.3–9.7
T_{max} (h)	5–6	5–6	4-5	5–7	5-8	7–12
$T_{1/2}$ (h)	32–37	49–84	29–39	27–49	49–60	37–45
$AUC_{0-\infty}$ (µg h/L)	117–130	148–242	128–227	237–345	168–316	299–419
			α-Solanine			
C_{max} (µg/L)	2.9–3.9	3.5–4.7	3.1–6.5	7.0–10.7	3.8–10.6	8.0–11.8
T_{max} (h)	5–6	5–6	4–4	5–5	5–8	7–8
$T_{1/2}$ (h)	17–35	5–28	8–17	14–18	1–-42	18–19
$AUC_{0-\infty}$ (µg h/L)	84–89	47–110	50–86	164–163	111–200	204–256

3.1.1.2.2. Solanidine

Claringbold et al. (1982) administered an unreported dose of [^{3}H]-solanidine by a single i.v. injection in two male and one female volunteers. Blood was sampled at various times and urine and feces were collected. The decrease of radioactivity in plasma followed a three-phase trend and levels were detected up to 150 h after the treatment. The authors estimated that in blood, about 70% of radioactivity detected was retained by erythrocytes, which could explain the slow clearance from plasma. After 24 h only 3 to 7% of the administered radioactivity was excreted in urine and feces.

The same authors also studied the presence of solanidine in five post-mortem liver samples. The extraction procedure from liver homogenates identified four solanidine containing fractions (free, glucuronide, glycoside and a residual fraction obtained following acid hydrolysis). In two out of five liver samples, no solanidine was detected in any of the four fractions. In another sample, no free solanidine was found and solanidine was observed mainly as the glucuronide (70%). In the two remaining samples, free solanidine accounted for 25 and 15%, respectively. In their acid hydrolysed fraction, solanidine accounted for 42 and 60%, respectively.

3.1.1.2.3. Biomarkers of exposure

Three studies were identified in which a radioimmunoassay method was developed to quantify solanidine in human plasma. In the first study, Matthew et al. (1983) described the method and reported the results on 34 human donors. One limitation of this method was that the rabbit immune serum developed had a 100% cross-reactivity with α-solanine, α-chaconine and demissidine. However, a preliminary extraction step using chloroform allowed the specific determination of solanidine. The

method was applied on plasma of 34 human donors (7 males and 27 females), randomly selected from a hospital clinic. All plasma samples showed quantifiable levels of solanidine, with mean levels of 1.56 ± 1.17 and 1.20 ± 0.93 ng/mL calculated for males and females, respectively.

The assay developed by Matthew et al. (1983) was subsequently applied by Harvey et al. (1985a) to study the levels of solanidine in 57 volunteers (30 males and 27 females) during winter. The consumption of potatoes and potato products of 33 subjects was recorded for one month. Two male volunteers maintained a potato-free diet for several weeks while their serum was regularly analysed for solanidine levels. The mean levels of solanidine were 10.8 ± 5.4 ng/mL and 7.9 ± 4.3 ng/mL in males and females, respectively. A positive correlation between consumption of potatoes and potato products and serum solanidine levels was found both in male and female subjects in the subgroup that recorded their diets. Avoidance of potato consumption corresponded to a drop in the serum levels of solanidine, reaching minimum levels (lower than the assay sensitivity of 0.5 ng/mL) three weeks after the cessation of potato intake.

In a subsequent study, Harvey et al. (1985b) adapted the radioimmunoassay method to quantify both solanidine and the TGA levels in serum and saliva. Two groups of volunteers (18 males and 15 females from UK, and 5 males and 5 females from Sweden) were analysed for levels of GAs during summer. All the subjects maintained their standard diet during the study, with the exclusion of three Swedish volunteers, who had daily consumption of two potato cultivars known to be high in GAs for a week before serum and saliva sampling. Mean levels of solanidine and TGAs (all subjects) were respectively 14.5 (5.8 ng solanidine/mL) and 38.5 nmol/L in serum, and 3.2 (1.3 ng solanidine/mL) and 3.7 nmol/L in saliva. No marked differences were observed among the UK and Swedish subjects following standard diets. Significant correlations were shown between serum and salivary levels both for solanidine and TGAs, and between serum solanidine and serum TGA levels. The three subjects consuming potato varieties high in GA levels had substantially higher serum and salivary levels compared to the other Swedish subjects (e.g. in serum solanidine mean levels were 10.5 nmol/L (4.2 ng solanidine/mL) in the 7 subjects following standard diets, and 80.8 nmol/L (32.3 ng solanidine/mL) in the 3 subjects with high-alkaloid diets).

3.1.1.3. Farm animals, horses and companion animals

Only scant information is available on the toxicokinetics of GAs in ruminants.

King and McQueen (1981) studied the effect of rumen on α-solanine and α-chaconine *in vitro*. Rumen contents were collected from a rumen-fistulated cow before the morning feeding and processed to remove feed particles. A mixture of α-solanine and α-chaconine was isolated from potato blossom and added to the rumen inoculum at a concentration of 1 mg/mL. At different timepoints up to 48 h, chloroform extracts were analysed by GC-NPD. The analyses revealed the formation of solanidine, that reached a maximum concentration at 12 h, when a subsequent conversion to a 5,6-dihydro derivative of solanidine, identified as 5β-solanidan-3β-ol, was observed. By 24 h the hydrolysis of the GA mixture reached a plateau and the sum of the 2 aglycones recovered from the rumen inoculum were on average 83% of the test concentration (approximately 30% constituted by solanidine and 50% by its 5,6-dihydro derivative).

Bushway et al. (1984) studied the potential transfer of GAs into cow's milk. Three samples of potato meal were analysed for their GA content and incorporated at 10 or 20% (w/w) in bovine feed ration, corresponding to 2.25 and 4.5 kg potato meal per day, respectively. The mean contents of α-chaconine and α-solanine in the potato meal samples were 515 and 285 mg/kg dw, respectively. This resulted in estimated daily exposure of 3.6 and 2.0 mg/kg bw for α-chaconine and α-solanine, respectively, at the highest level of potato meal intake, and of 1.78 and 0.99 mg/kg bw, respectively, at the lower level of

dietary inclusion[20]. Considering the evidence from the King and McQueen (1981) study, that GAs are almost completely converted to aglycones in rumen, milk samples from dairy cows fed with spiked bovine feed ration were analysed by GC-NPD for the presence of solanidine. No solanidine was detected in any milk sample (LOD: 0.14 mg/L).

3.1.1.4. Summary on toxicokinetics

Overall, limited conclusions on the toxicokinetics profiles of α-solanine and α-chaconine can be drawn based on the available evidence

Limited information is available from toxicokinetic studies in experimental animals. Both α-solanine and α-chaconine showed a relatively low bioavailability in experimental animals. Different toxicokinetic profile were observed in rats and hamsters. In hamsters there is indication for higher absorption and slower excretion rates for both substances in comparison with rats. In particular in rats, following oral exposure to labelled α-solanine or α-chaconine, radioactivity was excreted mainly via the feces (about 60% in 24 h and more than 80% after 4 days) as solanidine. Urinary excretion accounted for about 3–10% of the doses for α-solanine and 10% α-chaconine. Conversely, in hamsters, a lower fecal excretion was observed, with urinary excretion reaching about 10% for α-solanine and 21% for α-chaconine 7 days after the exposure. Information on the metabolic profiles of α-solanine and α-chaconine were mainly obtained via TLC techniques and are of low reliability.

The blood kinetics profiles of α-solanine and α-chaconine in humans were relatively well studied in two volunteer studies, using HPLC methods and showing comparable results. In volunteers exposed to mixtures of α-solanine and α-chaconine in the range of 0.3–1.25 mg TGA/kg bw via consumption of mashed potato meals, peak levels in serum were reached approximately 6–8 h after exposure, independently from the doses. Both substances showed long serum half-lives, suggesting a possible accumulation. α-Chaconine showed longer serum half-life than α-solanine (44 and 21 h, respectively). In addition, in one study the blood clearance of α-solanine and α-chaconine corresponded to the increase in solanidine levels, which reached a maximum level after 8 h and remained apparently constant up to 24 h following exposure. No further information on metabolism and no information on excretion was available in these studies. A volunteer study using tritiated solanidine support the evidence of slow clearance from plasma associated with a high retention of radioactivity in erythrocytes, with a corresponding slow excretion in urine and feces. In a separate experiment, solanidine was detected in 3 out of 5 post-mortem liver samples, either in the free form or in conjugated form (e.g. as glucuronide-conjugate). Levels of solanidine were regularly detected in the blood of human volunteers in a series of studies using a radioimmuno assay for the detection of solanidine or TGAs. In particular, in one experiment solanidine could be detected in serum of two volunteers up to three weeks after they started a potato-free diet. Overall, these studies provide some limited information on the kinetics of potato GAs, mainly suggesting their hydrolysis to solanidine following absorption and the slow clearance of solanidine from blood.

Only scant information is available on the toxicokinetics profile of potato GAs in ruminants. In an *in vitro* study, α-solanine and α-chaconine were shown to be extensively converted to solanidine and subsequently to a 5,6-dihydro derivative of solanidine, identified as 5β-solanidan-3β-ol. Another study showed no detection of solanidine in cows' milk upon consumption of feed spiked with a potato meal contaminated with α-solanine or α-chaconine, suggesting a low potential to transfer into milk for GA. No information was retrieved for other farm animals, horses or companion animals.

Toxicokinetic data on tomato and aubergine GAs and their aglycones could not be identified, neither for experimental animals nor for humans.

[20] Assuming 650 kg bw.

3.1.2. Toxicity in experimental animals

The selection criteria for studies relevant to inform the toxicity in experimental animals are described in **Section 2.1**.

In the descriptions below the doses are reported as mg/kg bw per day. If the doses were reported on molar basis in the original study, these were converted using the molecular weights given in **Appendix A.** Due to the weight of the oligosaccharide in the GAs, the aglycones have a lower molecular weight by a factor of ~ 2. This should be considered when comparing the dose-response relationships of GAs and aglycones at doses given in mg/kg bw per day.

3.1.2.1. Acute toxicity studies

3.1.2.1.1. GAs from edible parts of S. tuberosum

Regarding acute oral toxicity of potato GAs (**Table 6**), a single oral application of 250 mg/kg bw of α-solanine did not affect serum levels of liver transaminases or serum cholinesterase activity in rats (Dalvi et al., 1985). Mice showed no lethality at a dose of 1,000 mg/kg bw (Nishie et al., 1971).

An LD_{50} value of 590 mg/kg bw was identified for α-solanine in male and female rats (Gull, 1960). However, it is uncertain whether pure α-solanine was tested, as it appears likely that the standard (an extract of potato sprouts, purified by crystallisation) contained an unknown amount of α-chaconine. Therefore, the study cannot be used for risk assessment. Diarrhoea was reported to be the most frequent symptom at lower doses of the extract.

When compared to oral toxicity, considerably lower doses of potato GA, aglycones or mixtures caused symptoms or were lethal when applied i.p. or i.v. (see **Table 7**). For example, LD_{50} values for α-solanine ranged from 32.3–42 mg/kg bw in mice to 67 mg/kg bw in rats. Reported symptoms were diarrhoea and reduced spontaneous motor activity (Gull, 1960). The LD_{50} of α-chaconine, applied i.p., appears to be similar, i.e. 19.2 or 32.3 mg/kg bw in mice and 84 mg/kg bw in rats (Nishie et al., 1975; Sharma et al., 1979; Chaube and Swinyard, 1976). Rabbits showed a similar sensitivity as rats and mice towards α-solanine and α-chaconine, i.e. lethality occurred when α-solanine was applied at an i.v. dose of 10 mg/kg kg bw, and at i.p. doses of either 20 or 40 mg/kg bw or when animals received α-chaconine at an i.p. dose of 50 mg/kg bw (Nishie et al., 1971, 1975). Swinyard et al. (1973) reported an LD_{50} value of < 40 mg of α-solanine per kg bw for rhesus monkeys (administered i.p.).

Regarding i.p. application of defined mixtures of α-solanine and α/ß-chaconine, Syrian hamsters were relatively sensitive showing an LD_{50} value between 10 and 25 mg/kg bw, while in rats the LD_{50} value was 60 or 75 mg/kg bw (Gull, 1960; Phillips et al., 1996; Chaube and Swinyard, 1976).

The aglycone solanidine was tested in mice only at a single i.p. dose of 500 mg/kg bw, which exerted no lethal effects (Nishie et al., 1971).

3.1.2.1.2. GAs from edible parts of food plants other than S. tuberosum

The studies identified are reported in **Table 8** and **9**.

Wilson et al. (1961) reported on 100% lethality in rats after applying 900–1,000 mg/kg bw in rats. Histologic analyses revealed an acute mucosal erosion of the stomach associated with a mild acute inflammation in the submucosa.

Regarding mixtures, Baker et al. (1989) treated hamsters with water suspensions of lyophilised seeds of aubergine containing 160 mg TGA/kg dry weight (dw). The authors reported that solasodine was the only aglycone detectable. A dose of 6,316 mg dw (approximately 1 mg TGA/kg bw) caused haemorrhagic gastritis in hamsters.

When compared to oral toxicity, considerably lower doses of tomato GAs were lethal when applied i.p. or i.v. The LD_{50} values for α-tomatine in mice ranged from 18 mg/kg bw (i.v.) to 32.4 mg/kg bw (i.p.) (Sackman et al., 1959; Wilson et al., 1961; Nishie et al., 1975). After s.c. administration, the LD_{50} was raised to >1,000 mg/kg bw (Sackman et al., 1959). Rabbits appear to be less sensitive than mice, since a single i.p. dose of α-tomatine at 100 mg/kg bw did not cause death of the animals (Nishie et al., 1975).

The LD_{50} value of α-solamargine was determined to be 42 mg/kg bw, when applied i.p. to rats (Al Chami et al., 2003).

3.1.2.1.3. Summary on acute toxicity studies

Regarding oral exposure to α-solanine, an LD_{50} of 590 mg/kg bw was identified for rats, while mice showed no lethality at a dose of 1,000 mg/kg bw. When compared to gavage, considerably lower i.p. doses elicited effects, e.g. in mice, the LD_{50} was 40–42 mg/kg i.p., and in rats increased liver transaminases in serum were reported at 20 mg/kg bw i.p. Likewise, the aglycone solanidine exerted no significant effect in mice at 500 mg/kg bw i.p.

Relative potency of toxicity of different GAs can be deduced after i.p. application and appear to be similar, i.e. in rats the LD_{50} was 67 mg/kg bw for α-solanine and 84 mg/kg bw for α-chaconine, and in mice it was 32.3–42 mg/kg bw for α-solanine and 19–32.3 mg/kg bw for α-chaconine. Lethality occurred in New-Zealand rabbits starting at 40 mg/kg bw of α-solanine and at 50 mg/kg bw for α-chaconine. In mice, the aglycone solanidine exerted no effect when applied at 500 mg/kg bw i.p., contrasting to the pronounced toxicity of α-solanine and α-chaconine after the identical route of exposure.

Lethal doses of α-tomatine were approximately 900–1,000 mg/kg bw p.o. in rats. Acute lesions comprised erosion, and also inflammation occurred in the stomach. When applied i.v. or i.p., LD_{50} values of 18 to 32.4 mg/kg bw were found in rodents. Again, the parenteral route was found to increase the toxicity of GAs from tomato.

1404 **Table 6.** Acute toxicity in experimental animals: oral administration of glycoalkaloids (GAs) (α-solanine) or mixtures, from edible parts of *S. tuberosum*.

Test compounds	Species Dose route Doses	Observed effects	Highest dose with no effect (mg/kg bw)	Lowest dose with effect (mg/kg bw)	LD_{50} (mg/kg bw)	Reference
α-Solanine [(a)]	Sprague-Dawley rats (M) No/sex/group: 5 Single gavage: 0, 20 or 250 mg/kg bw	-	250			Dalvi et al. (1985)
α-Solanine [(a)]	Albino mice (M) No/sex/group: n.r. Single oral application: 0 or 1,000 mg/kg bw	-	1,000			Nishie et al. (1971)
Mixture of α-solanine and unknown proportion of α-chaconine [(b)]	White rats (M/F) No/sex/group: 9 Single gavage: 300, 600 or 1,200 mg/kg bw	Diarrhoea Lethality		300	 590	Gull (1960)

1405 F: females. M: males. n.r.: not reported.
1406 (a): Chemical standard obtained from a commercial supplier, with high purity (95% pure or higher).
1407 (b): Poorly characterised chemical standard (e.g. purity or composition not defined).

1408 **Table 7.** Acute toxicity in experimental animals: non-oral administration of glycoalkaloids (GAs) (α-solanine, α-chaconine), mixtures, and the aglycone
1409 (solanidine), from edible parts of *S. tuberosum*.

Test compound	Species/ dose route	Observed effects	Highest dose with no effect (mg/kg bw)	Lowest dose with effect (mg/kg bw)	LD_{50} (mg/kg bw)	Reference
α-Solanine [a]	Rats (M/F) / i.p.	↓ Hepatic glycogen content		5		Satoh et al. (1967)
α-Solanine [a]	Rats (M) / i.p.	↑ Serum liver transaminases ↓ Serum cholinesterase activity		20		Dalvi et al. (1985)
α-Solanine [b]	Wistar rats (F) / i.p.	Lethality			67	Chaube and Swinyard (1976)
α-Solanine [a]	Albino mice (M) / i.p.	Lethality ↓ Spontaneous motor activity	5	10	42	Nishie et al. (1971)
α-Solanine [a]	Balb/c mice (F) / i.p.	Lethality	10	20	40	Mohsenikia et al. (2013)
α-Solanine [a]	Swiss-Webster mice (M) / i.p.	Lethality			32.3	Patil et al. (1972)
α-Solanine [b]	Swiss-Webster mice (M) / i.p.	Lethality			32.3	Sharma et al. (1979)
α-Solanine [a]	Swiss-Webster mice (M) / i.p.	Lethality			34.5	Nishie et al. (1975)
α-Solanine [a]	Balb-c mice (M) / i.p.		40			Khodayari et al. (2013)
α-Solanine [a]	Rabbits (sex n.r.) / i.p. or i.v.	EEG alterations (i.p.) ↑ Respiratory rate, lower blood pressure (i.v.) Lethality (i.p.) Lethality (i.v.)	< 15	> 15 2 20 10		Nishie et al. (1971)

α-Solanine [a]	White New-Zealand rabbits (sex n.r.) / i.p.	Lethality		40		Nishie et al. (1975)
α-Solanine [a]	Rhesus monkey (sex n.r.) / i.p.	Lethality (100%)		40 2x20	< 40	Swinyard et al. (1973)
α-Chaconine [b]	Swiss-Webster mice (M) / i.p.	Lethality			32.3	Nishie et al. (1975)
α-Chaconine [a]	Swiss-Webster mice (M) / i.p.	Lethality			19.2	Sharma et al. (1979)
α-Chaconine [b]	Wistar rats (F) / i.p.	Lethality			84	Chaube and Swinyard (1976)
α-Chaconine [b]	Sprague-Dawley rats (M) / i.p.	Altered EEG of occipital cortex Bradycardia	 10	10 20		Aldous et al. (1980)
α-Chaconine [b]	White New-Zealand rabbits (sex n.r.) / i.p.	Lethality		50		Nishie et al. (1975)
Mixture of α-solanine and unknown proportion of α-chaconine [c]	Rats (M/F) / i.p.				75	Gull (1960)
Mixture (a) α-solanine: α-chaconine (1:1)	Syrian hamster (M) / i.p.	Lethality	10	25	> 10 < 25	Phillips et al. (1996)
Mixture (d) α-solanine, α-chaconine and ß-chaconine comprised ~20% of the extract at a ratio of 1:2:0.3, respectively	Wistar rats (F) / i.p.				60	Chaube and Swinyard (1976)
Solanidine [a]	Albino mice (M) / i.p.	-	500			Nishie et al. (1971)

EEG: electroencephalogram. F: female. M: male. n.r.: not reported
(a): Chemical standard obtained from a commercial supplier, with high purity (95% pure or higher).
(b): Chemical standard obtained by in-house isolation from plant material, and checked for purity (95% pure or higher).
(c): Poorly characterised chemical standard (e. g. purity or composition not defined).
(d): A non- or only partly purified extract or material for which the glycoalkaloid composition and concentration is given.

1415 **Table 8.** Acute toxicity in experimental animals: oral administration of glycoalkaloids (GAs) (α-tomatine) from edible parts of *S. lycopersicum* and extracts
1416 from edible parts of *S. melongena*.

Substance (purity)	Species (sex) Dose route Doses	Observed effects	Highest dose with no effect (mg/kg bw)	Lowest dose with effect (mg/kg bw)	LD_{50} (mg/kg bw)	Reference
α-Tomatine [(a)]	Rats (M/F) No/sex/group: n.r. Oral dose of 900–1,000 mg/kg bw given within 24 h	100% lethality		900–1,000		Wilson et al. (1961)
Water suspensions of ground lyophilised fruits of ripe *S. melongena* [(b)]	Syrian hamster (sex n.r.) No/sex/group: 2 Single oral application: 6,316 mg suspension/kg bw TGA content of dried matter: 0.016% (corresponding to 1.0 mg TGA/kg bw)	Haemorrhagic gastritis		1		Baker et al. (1989)

1417 F: females. M: males. n.r.: not reported
1418 (a): Chemical standard obtained by in-house isolation from plant material, may contain α-dehydrotomatine as an impurity.
1419 (b): A non- or only partly purified extract or material for which the GA composition and concentration is given.

1420

Table 9. Acute toxicity in experimental animals: non-oral administration of glycoalkaloids (GAs) (α-tomatine, α-solamargine) from edible parts of *S. lycopersicum* or *S. melongena*.

Substance (Purity)	Species (sex) Dose route	Observed effects	Highest dose with no effect (mg/kg bw)	Lowest dose with effect (mg/kg bw)	LD_{50} (mg/kg bw)	Reference
α-Tomatine [b]	Mice (sex n.r) i.v.	Lethality			18	Wilson et al. (1961)
α-Tomatine [a]	Swiss-Webster mice (M) i.p.	Lethality			32.4	Nishie et al. (1975)
α-Tomatine [b]	Mice (sex n.r) i.p.	Lethality			25	Sackmann et al. (1959)
α-Tomatine [b]	Mice (sex n.r) s.c.	Lethality			> 1,000	Sackmann et al. (1959)
α-Tomatine [a]	White New-Zealand rabbits (sex n.r.) i.p.	-	100			Nishie et al. (1975)
α-Solamargine [c]	Wistar rats (M) i.p.	Lethality			42	Al Chami et al. (2003)

M: male. n.r.: not reported
(a): Chemical standard obtained from a commercial supplier, may contain α-dehydrotomatine as an impurity.
(b): Chemical standard obtained by in-house isolation from plant material, may contain α-dehydrotomatine as an impurity.
(c): Poorly characterised chemical standard (e. g. purity or composition not defined).

3.1.2.2. Repeated dose toxicity studies

3.1.2.2.1. GAs and aglycones from edible parts of S. tuberosum

The studies on effects following repeated oral doses are summarised in **Table 10**, **11** and **12**. The no-observed-effect level (NOEL) (or lowest-obserserved-effect level, LOEL) and NOAEL (LOAEL) values indicated, have been derived by the CONTAM Panel, unless stated otherwise.

α-Solanine

α-Solanine reduced body weight gain and the absolute liver weight when given at 416.6 mg/kg bw per day (the only dose administered) for one week to Swiss Webster mice (Friedman et al., 1996). The food intake remained unaffected.

A lower dose (180 mg α-solanine/kg bw per day) reduced the growth rate in rats treated over a period of 5 weeks which was not due to an altered food intake of the animals. No further compound-related symptoms were reported (Gull, 1960). Based on the lower body weight at 180 mg/kg bw per day, a NOAEL of 90 mg α-solanine/kg bw per day was derived.

Hamsters exhibited lethality in 50% of the animals and reduced growth and relative liver weights when exposed daily to 100 mg α-solanine/kg bw (the only dose administered) for 5 days. The lowered body weight gain might have been caused by a decreased food intake by the animals. Serum glucose, serum creatine, red blood cell (RBC), haemoglobin and haematocrit were elevated (Langkilde et al., 2008).

α-Chaconine

α-Chaconine lowered body and liver weights at 409 mg/kg bw per day (the only dose administered) in mice over a period of 7 days (Friedman et al., 1996). The food intake by the animals was unaffected.

In hamsters, 100 mg/kg bw per day (the only dose administered) for 5 days induced lethality (in 25% of the animals) and reduced body weight gain and relative liver weights despite of an unaltered food intake (Langkilde et al., 2008). Serum glucose, serum creatine, RBC, haemoglobin and haematocrit were elevated.

Solanidine

The aglycone solanidine was administered to mice for up to 28 days at doses ranging from 95.5 to 381.8 mg/kg bw per day (Friedman et al., 1996, 2003a). Adverse effects were seen at 190.9 mg/kg bw per day, such as decreased body weight gain and increased absolute and relative liver weights (Friedman et al., 1996). Feed intake was lowered at a daily dose of 381.8 mg/kg bw. Based on increases in absolute and relative liver weights at day 14 and 28 of treatment, a NOAEL of 95.5 mg solanidine/kg bw per day was derived.

Mixtures of GAs

Friedman (1992) fed mice potatoes containing 64 mg TGAs (20 mg α-solanine and 37 mg α-chaconine)/kg fresh weight. α-Solanine and α-chaconine were quantified by HPLC-UV, while TGA were determined by a colourimetric method (possibly detecting other GAs as well, see **Section 1.3.2)**. The potatoes were admixed as powder at high concentrations (up to 40%) to the diet presumably causing dietary imbalances of macro- and micronutrients. Increased absolute and relative weights of pancreas were observed at the highest TGA dose of 23.2 mg/kg bw per day. The CONTAM Panel agrees with the assumption of the authors that the effects on the pancreas may be due to presence of trypsin inhibitor activity in the potato powder and not caused by GAs. Consequently, no NOAEL was derived for this study.

For a period of 3 days, Phillips et al. (1996) treated male hamsters orally with doses up to 50 mg TGA/kg bw per day at a ratio of α-solanine: α-chaconine of approximately 1:1. No adverse effects could be observed. However, the study was conducted with a low number of animals per treatment group.

In a series of experiments, Langkilde et al. (2008, 2009, 2012) tested defined mixtures of α-solanine and α-chaconine in female hamsters. At a ratio α-solanine:α-chaconine of 1:2.5, gavage of 75 mg TGA/kg bw per day induced lethality (in 25% of the animals), lowered food intake, decreased the relative liver weight and increased hemoglobin and haematocrit within 5 days of treatment (Langkilde et al., 2008). At gavage of 100 mg TGA/kg bw per day, lethality was seen again in 25% of the animals, body weight gain was lowered and the relative weight of adrenals, serum creatine and RBC were elevated (Langkilde et al., 2008). The LOAEL in this study was 75 mg TGAs/kg bw per day based on the lethality and the decreased relative liver weight in female hamsters.

An α-solanine:α-chaconine ratio of 1:3.7 with a dose of 33 mg TGA/kg bw, applied by daily gavage, raised white blood cells (WBC), RBC and serum aspartate aminotransferase (AST), and decreased albumin, proteins and urea in blood within 4 weeks (Langkilde et al., 2009). When the ratio α-solanine:α-chaconine was 1:70 at a dose of 33.3 mg TGAs/kg bw per day, death occurred in one of the four treated animals 2 h after gavage. Weight of adrenal glands, AST, WBC and RBC were affected (Langkilde et al., 2009). Thus, the NOAEL was 10 mg TGAs/kg bw per day irrespective of whether α-solanine and α-chaconine were applied at a 1:3.7- or 1:70-ratio.

Langkilde et al. (2012) investigated the putative effects of a wildtype ('Desiree') and a genetically modified potato line (SGT 9-2). While the wildtype potato contained α-solanine:α-chaconine at a ratio of 1:3.1, the SGT 9-2 line exhibited a ratio of 1:25.5–26 and a considerably lower TGA content (determined by HPLC-UV). All animals of the study received diets, which were identical in the concentrations of minerals, vitamins, sucrose/dextrin, soybean oil and cellulose but contained 0%, 20%, 40% or 60% freeze-dried powder of control or SGT 9-2 potatoes (trypsin inhibitor activity of the powders was not excluded). The variable proportions of potato powder in the diets were compensated to some extent by adjusted contents in caseinate and corn/potato starch. Since the TGA content of the SGT 9-2 line was considerably lower than in the controls, commercially available α-solanine and α-chaconine were added to the diets with SGT 9-2 in order to treat animals with TGA doses as occurring in control potatoes. However, the ratio between the two GAs was kept as being typical for the SGT 9-2 line. In addition, one of the diets (40% powder of control potatoes) was spiked with commercially available α-solanine and α-chaconine to elevate the TGA dose to 33 mg/kg bw per day. When the relative proportion of α-solanine:α-chaconine was 1:3.1, a daily TGA dose of 33 mg/kg bw exerted no effect over a period of 90 days. At a ratio of 1:25.5–26 again no effects were seen at TGA doses of up to 20.6 mg/kg bw per day (Langkilde et al., 2012). In any case, the NOEL in this study was 33 mg TGA/kg bw per day at a ratio α-solanine:α-chaconine of 1:3.1 (the highest dose tested).

In contrast to data published in 2009, Langkilde et al. (2012) did not find adverse effects when hamsters were exposed to 33 mg TGAs/kg bw per day for 90 days with a ratio of α-solanine:α-chaconine of approximately 1:3.1. In this study the NOEL for TGAs was 33 mg/kg bw per day which is considerably higher than in the study appearing in 2009. The different outcome may be based on the fact, that in the study of 2009 purified standards were applied to the animals as bolus by gavage, whereas in the study published in 2012, freeze-dried potato powder, partly spiked with commercial standards, were administered via the diet.

Preparations based on edible parts of *S. tuberosum and sprouts*

In some experiments the effects of plant parts of potatoes were studied. Tuśnio et al. (2013a) used potato sprout meal with 41.85 g TGAs/kg admixed to the diet to provide daily doses of up to 36 mg TGAs/kg bw to rats (no information on the ratio of α-solanine:α-chaconine provided). The potato sprout meal did not exhibit any trypsin inhibitor activity. This treatment did not elicit any effect when applied over a period of 28 days (Tuśnio et al., 2013a). Thus, the NOEL was 36 mg TGAs/kg bw per day (the highest dose tested).

In a second experiment, commercial potato protein concentrates were applied, which contained 677–3,185 mg TGAs/kg (Tuśnio et al., 2013a) and which did exhibit trypsin inhibitor activity. Again, the ratio of α-solanine:α-chaconine was not given and might have been variable. TGA doses of 13.9–52.7 mg/kg bw per day affected the gastrointestinal tract and caused decreased relative stomach weight, increased relative weight of the intestine and elevated serum alanine aminotransferase (ALT) levels. The study authors speculated that these alterations were rather due to trypsin inhibitor activity than the GA content in the diet (Tuśnio et al., 2013a). Consequently, no NOAEL or LOAEL could be derived.

Azim and colleagues performed three studies in rabbits and compared the effects of green or ripe potatoes, fed as chow (Azim et al., 1982, 1983, 1984). In all studies, the GA content in potatoes was determined but no ratio of α-solanine:α-chaconine was given. Furthermore, the studies lack negative controls. Generally, decreased body weight and increased relative liver weights were reported. However, the data are difficult to interpret since untreated controls are missing. No NOEL or LOEL could be derived.

Swinyard et al. (1973) used potato strain B5141-6, containing a mean TGA content of 260 mg/kg fw (range: 150–350 mg/kg fw). α-Solanine accounted for 25% of the TGA content, while the remaining GAs were not specified, but probably contained α-chaconine. The tubers were diced and admixed to the diet. By this procedure Rhesus monkeys were treated with approximately 3.47 mg TGAs/kg bw per day for 25 days, which did not elicit any significant effect on food consumption and body weight. However, the study lacks an untreated control group. Therefore, no NOAEL could be derived.

Summary of repeated dose toxicity studies with GAs and aglycones from edible parts of *S. tuberosum* and sprouts

To conclude, α-solanine and α-chaconine reduced body and liver weights in mice after one week at 416.6 and 409 mg/kg bw per day, respectively. A NOAEL was set for α-solanine at 90 mg/kg bw per day based on reduced growth of rats, treated for 5 weeks. In hamster, body and relative liver weights were lowered by 100 mg/kg bw of either α-solanine or α-chaconine, applied for 5 days. The aglycone solanidine caused adverse effects in mice treated with 190.9 mg/kg bw per day up to 4 weeks; the NOAEL was 95.5 mg/kg bw per day.

Mixtures of α-solanine and α-chaconine were applied to hamsters over 90 days (Langkilde et al., 2012). The NOEL was 20.6 mg/kg bw per day (α-solanine:α-chaconine ratio of 1:25.5–26) and 33 mg/kg bw per day (α-solanine:α-chaconine ratio of 1:3.1) (the highest doses tested). The NOEL of 33 mg/kg bw per day is identical to the LOAEL reported in another study, in which 33 mg/kg (α-solanine:α-chaconine ratio of 1:3.7), applied via gavage for 4 weeks, caused gastrointestinal effects in hamsters (Langkilde et al., 2009). The different outcome at the identical dose may be due to the bolus effect of gavage in the 2009 study and the matrix effects of the basal chow in the 2012 study.

Regarding edible parts of potato and sprouts, the NOEL was 36 mg TGA/kg bw per day, fed to rats for 28 days (the highest dose tested). No effects were seen in monkeys at a daily dose of 3.47 mg TGA/kg bw, applied for 25 days.

3.1.2.2.2. GAs and aglycones from edible parts of food plants other than S. tuberosum

The studies identified are reported in **Table 13**.

α-Tomatine

α-Tomatine was tested in rats at doses of 1.25 to 250 mg/kg bw per day over periods of 5 to 200 days. No effects were reported at 250 mg/kg bw per day when given for 5 days, and at 20 mg/kg bw applied over a period of 200 days (Wilson et al., 1961). A NOAEL of 20 mg/kg bw per day for rats was identified from this study (the highest dose applied in this study). A dose of 600 mg/kg bw caused decreased intrahepatic cholesterol after 7 days of administration (Cayen et al., 1971). Based on the lower hepatic cholesterol concentration, the LOEL was 600 mg α-tomatine/kg bw per day in male rats. When the effects on the cholesterol metabolism were studied in rats in more detail, Cayen et al. (1971) reported on reduced uptake of dietary cholesterol by the liver, an elevated hepatic and intestinal cholesterol synthesis and increased fecal sterol excretion without affecting bile acid excretion at 1,200 mg α-tomatine/kg bw per day for 2 weeks (see also **Section 3.1.2.9**).

In mice, 496 mg α-tomatine/kg bw per day (the only dose administered) for one week did not affect body or liver weights (Friedman et al., 1996).

When hamsters were treated with 30.5 mg/kg bw per day for 3 weeks, the LOEL was 30.5 mg α-tomatine/kg bw per day, based on the elevated fecal sterol and coprostanol excretion (Friedman et al., 2000a) (see also **Section 3.1.2.9**).

α-Solasonine

α-Solasonine increased the body weight gain in mice at 424.4 mg/kg bw per day (the only dose administered) for 7 days despite of an unaltered food intake by the animals. No further effects were reported (Friedman et al., 1996).

Tomatidine

Wilson et al. (1961) obtained the aglycone tomatidine by hydrolysis, as described by Fontaine et al. (1948). When given to rats daily at 1.25 to 20 mg/kg bw for 200 days, tomatidine exerted no significant effects (Wilson et al., 1961). The NOEL in this study was 20 mg tomatidine/kg bw per day for rats (the highest dose tested). Two studies in mice reported an elevated absolute and/or relative liver weight at 199.5 mg/kg bw per day, when applied for 7 or 14 days (Friedman et al., 1996, 2003). Further observations were decreased feed intake (day 7 at 399 mg/kg bw per day) and reduced body weight gain (day 14 at 199.5 mg/kg bw per day) (Friedman et al., 1996, 2003). The LOAEL was 199.5 mg/kg bw per day, based on the reduced body weight gain and elevated absolute and relative liver weight in female mice.

Solasodine

The aglycone solasodine was tested in Swiss-Webster mice at doses of up to 397.2 mg/kg bw per day (Friedman, 1992; Friedman et al., 1996, 2003). In the first study, Friedman (1992) reported on decreased body weight gain (despite of unaltered food intake), gastric gland degeneration, cholangiohepatitis, elevated relative liver weight and increased serum glutamic oxaloacetic transaminase (GOT), glutamic pyruvic transaminase (GPT) and bilirubin, when animals received 160 mg/kg bw per day for 7 and/or 14 days. This indicates that the gastrointestinal tract and the liver were affected severely by this compound. The NOAEL was 80 mg/kg bw per day, based on the effects on body weight and on the gastrointestinal tract at 160 mg/kg bw per day in mice. In the two subsequent studies, the

same study authors administered partly higher doses than in the study published in 1992 and focused on compound effects on body and liver weights (Friedman et al., 1996, 2003). Decreased body weight gain (despite of unaltered food intake) and increased relative and/or absolute liver weights were induced at 198.6 mg solasodine/kg bw per day (Friedman et al., 2003). At 397.2 mg/kg bw per day feed intake was reduced at day 7 of treatment (Friedman et al., 1996). Due to the high level and the limited dose ranges applied, these studies were not used to identify a NOAEL.

Friedman et al. (1996) compared the relative potencies of equimolar doses of the aglycones with regard to the increase in liver weight in mice and found that solanidine, solasodine and tomatidine were nearly equal (see also **Table 10**). This was confirmed by Friedman et al. (2003). In contrast, equimolar doses of the GA α-solasonine decreased insignificantly the liver weight (Friedman et al., 1996). Wilson et al (1961) applied α-tomatine and tomatidine at 20 mg/kg bw per day for a period of 200 days to rats. When considering the different molecular weights of the compounds the α-tomatine dose was 20 µmol/kg bw per day and the tomatidine dose 50 µmol/kg bw per day. Nevertheless, tomatidine did not elicit any effect, as was true also for α-tomatine.

Summary of repeated dose toxicity studies with GAs and aglycones from edible parts of food plants other than S*. tuberosum*

To conclude, α-tomatine exerted no effects in mice receiving 496 mg/kg bw per day for 1 week. A NOEL of 20 mg/kg bw per day (the highest dose tested) was identified for rats, treated for 200 days. In hamsters, the LOEL was 30.5 mg/kg bw per day, based on an increased fecal sterol and coprostanol excretion. α-Solasonine increased the body weight gain in mice when treated daily with 424.4 mg/kg bw for 1 week (the only dose applied). For tomatidine, a LOAEL of 199.5 mg/kg bw per day was derived in mice, and a NOEL of 20 mg/kg bw per day (the highest dose applied) in rats. For solasodine, a NOAEL of 80 mg/kg bw per day was identified in mice, based on decreased body weight gain, gastric gland degeneration, and liver toxicity at higher doses.

1621 **Table 10.** Repeated dose oral toxicity studies with glycoalkaloids (GAs) (α-solanine and α-chaconine) and the aglycone (solanidine) from edible parts of *S.*
1622 *tuberosum*.

Test compound	Species/Experimental design and doses	Most sensitive endpoint	Highest dose with no effect (mg/kg bw/day)	Lowest dose with effect (mg/kg bw/day)	Reference
α-Solanine [(b)]	**Swiss-Webster mice** (F) No/sex/group: 8 Diet: 0 or 2.4 mmol/kg diet (equivalent to 0 or 416.6 mg/kg bw per day [(d)]) Duration: 7 days	↓ bw gain ↓ absolute liver weight		416.6 416.6	Friedman et al. (1996)
α-Solanine [(c)]	**White rats** (M) No/sex/dose: 5–10 Diet: 0, 20, 50, 100, 250, 500, 1,000, 2,000 mg/kg diet (equivalent to 0, 1.8, 4.5, 9, 22.5, 45, 90, or 180 mg/kg bw per day [(d)]) Duration: 5 weeks	↓ growth rate	90	180	Gull (1960)
α-Solanine [(a)]	**Golden Syrian hamster** (F) No/sex/group: 4–8 Gavage: 0 or 100 mg/kg bw per day Duration: 5 days	Lethality (50% of animals) ↓ food intake and bw gain ↓ relative liver weight ↑ RBC, HGB, HCT ↑ serum glucose, creatine ↓ serum alk phosphatase, albumin		100 100 100 100 100 100	Langkilde et al. (2008) For mixtures of α-solanine and α-chaconine, see **Table 12**
α-Chaconine [(b)]	**Swiss-Webster mice** (F) No/sex/group: 8 Diet: 0 or 2.4 mmol/kg diet (equivalent to 0 or 409 mg/kg bw per day [(d)]) Duration: 7 days	↓ bw gain (day 7) ↓ absolute and relative liver weight (day 7)		409 409	Friedman et al. (1996)

α-Chaconine [(a)]	**Golden Syrian hamster** (F) No/sex/group: 4–8 Gavage: 0 or 100 mg/kg bw per day Duration: 5 days	Lethality ↓ bw gain ↓ relative liver weight ↑ RBC, HGB, HCT ↑ serum glucose, creatine	0	100 100 100 100 100	Langkilde et al. (2008) For mixtures of α-solanine and α-chaconine, see **Table 12**
Solanidine [(a)]	**Swiss-Webster mice** (F) No/sex/group: 8–16 Diet: 0, 1.2, 2.4 or 4.8 mmol/kg diet (equivalent to 0, 95.5, 190.9 or 381.8 mg/kg bw per day [(d)]) Duration: 7, 14 or 28 days	↓ food intake (day 14) ↓ bw gain (day 7) ↓ bw gain (day 14) ↑ absolute and relative liver weight (day 7) ↑ absolute and relative liver weight (day 14, 28)	190.9 (no 95.5 group) 190.9 (no 95.5 group) 95.5	381.8 190.9 381.8 190.9 190.9	Friedman et al. (1996)
Solanidine [(a)]	**Swiss-Webster mice** (F) No/sex/group: 10 Diet: 0 or 2.4 mmol/kg diet (equivalent to 0 or 190.9 mg/kg bw per day [(d)]) Duration: 14 days	↓ bw gain ↑ absolute and relative liver weight		190.9 190.9	Friedman et al. (2003)

F: female. M: male. bw: body weight. RBC: red blood cell. HGB: haemoglobin. HTC haematocrit.
(a): Chemical standard obtained from a commercial supplier, with high purity (95% pure or higher).
(b): Chemical standard obtained by in-house isolation from plant material, and checked for purity (95% pure or higher).
(c): Poorly characterised chemical standard (e. g. purity or composition not defined).
(d): Applying EFSA default values.

1630 **Table 11.** Repeated oral toxicity studies with defined mixtures of glycoalkaloids (GAs) (α-solanine and α-chaconine) from edible parts of *S. tuberosum*.

Test compound	Species Dose route Doses	Observed effects	Highest dose with no effect GA (α-solanine/α-chaconine) (mg/kg bw per day)	Lowest dose with effect GA (α-solanine/α-chaconine) (mg/kg bw per day)	Reference
Potatoes containing 6.4 mg TGA (20 mg α -solanine; 37 mg α-chaconine) per kg fw, admixed as powder to diet [(d)]	**Swiss-Webster mice** (sex n.r) No/sex/group: 4–5 Oral application (diet): 0, 1, 5, 10, 20 or 40% potato powder in the diet with approximately 0, 2.9, 14.6, 29.1, 58.2 or 116.4 mg TGA/kg diet [(g)] (equivalent to 0, 0.6, 5.8, 11.6 and 23.2 mg TGA/kg bw per day [(f)]) Duration: 7 and 14 days	↑ absolute and relative pancreas weight	~11.6 (3.63/6.71)	~23.2 (7.25/13.4)	Friedman (1992)
α-Solanine:α-chaconine (1:1) [(a)]	**Syrian hamster** (M) No/sex/group: 2 Gavage: 0, 5, 10, 25 or 50 mg GA/kg bw per day Duration: 3 days	No effect	50 (25/25)		Phillips et al. (1996)
α-Solanine:α-chaconine (1:2.5) [(a)]	**Golden Syrian hamster** (F) No/sex/group: 4–8 Gavage: 0, 75 or 100 mg/kg bw per day Duration: 5 days	Lethality (in 25%) ↓ food intake ↓ bw gain ↓ relative liver weight ↑ relative adrenal weight	 75 (21.4/53.6) 75 (21.4/53.6)	75 (21.4/53.6) 75 (21.4/53.6) 100 (28.6/71.4) 75 (21.4/53.6) 100 (28.6/71.4)	Langkilde et al. (2008) For single compound controls see **Table 11**

		↑ HGB, HCT		75 (21.4/53.6)	
		↑ RBC	75 (21.4/53.6)	100 (28.6/71.4)	
		↑ serum creatine	75 (21.4/53.6)	100 (28.6/71.4)	
α-Solanine:α-chaconine (1:3.7) [(a)]	**Golden Syrian hamster** (F) No/sex/group: 4 Gavage: 0, 0.1, 0.33, 1, 3.3, 10 or 33.3 mg/kg bw per day Duration: 28 days	↑ WBC and RBC	10 [(h)] (2.1/7.9)	33 (7/26)	Langkilde et al. (2009)
		↓ albumin, urea and plasma proteins	10 [(h)] (2.1/7.9)	33 (7/26)	
		↑ serum AST	10 [(h)] (2.1/7.9)	33 (7/26)	
		Distended stomach and intestine	10 [(h)] (2.1/7.9)	33 (7/26)	
α-Solanine:α-chaconine (1:70) [(a)]	**Golden Syrian hamster** (F) No/sex/group: 4 Gavage: 0, 0.1, 0.33, 1, 3.3, 10 or 33.3 mg/kg bw per day Duration: 28 days	Mortality (25%)	10 (0.1/9.9)	33.3 (0.5/32.8)	Langkilde et al. (2009)
		↑ WBC and RBC	10 [(h)] (0.1/9.9)	33.3 (0.5/32.8)	
		↓ albumin, urea and plasma proteins	10 [(h)] (0.1/9.9)	33.3 (0.5/32.8)	
		↑ serum AST	10 [(h)] (0.1/9.9)	33.3 (0.5/32.8)	
		Distended stomach and intestine	10 [(h)] (0.1/9.9)	33.3 (0.5/32.8)	
α-Solanine:α-chaconine (1:3.1) *S. tuberosum* Desiree (added to chow as freeze-dried powder) [(d)] [(e)]	**Golden Syrian hamster** (F) No/sex/group: 10 Diet: TGA at 0, 6.9, 13.8, 20.6 or 33 [(f)] mg/kg bw per day Duration: 90 days	No effects	33 (8/25)		Langkilde et al. (2012)
α-Solanine:α-chaconine (1:25.5-26) *S. tuberosum* SGT9-2 (added to chow as freeze-dried powder)	**Golden Syrian hamster** (F) No/sex/group: 10 Diet: TGA at 0, 6.9, 13.8 or 20.6 mg/kg bw per day	No effects	20.6 (1.0/19.6)		Langkilde et al. (2012)

and spiked with GA from commercial source (d)	Duration: 90 days				
α-Solanine:α-chaconine (1:25.5–26) from *S. tuberosum* SGT 9-2 (added to chow as freeze-dried powder) (d)	**Golden Syrian hamster** (F) No/sex/group: 10 Diet: TGA at 0 or 8.6 mg/kg bw per day Duration: 90 days	No effects	8.6 (0.3/8.5)		Langkilde et al. (2012)

F: females. M: males. n.r.: not reported. TGA: total GA content. AST: aspartate aminotransferase. WBC: white blood cells. RBC: red blood cells. HCT: haematocrit. HBG: haemoglobin.
(a): Chemical standard obtained from a commercial supplier, with high purity (95% pure or higher).
(b): Chemical standard obtained by in-house isolation from plant material, and checked for purity (95% pure or higher).
(c): Poorly characterised chemical standard (e. g. purity or composition not defined) or with a non or only partly purified extract or material for which the GA composition and concentration is given.
(d): A non or only partly purified extract or material for which the GA composition and concentration is given.
(e): Spiked with GAs from commercial source.
(f): Applying EFSA default values
(g): Anticipating 78% water content of potatoes, the GA content would be 291 mg TGA, 91 mg α-solanine and 168 mg α-chaconine per kg potato powder.
(h): Approximative values read from the figures of the study.

1640 **Table 12.** Repeated oral toxicity studies with edible plant parts of *S. tuberosum* and sprouts.

Test compound	Species Experimental design	Observed effects	Highest dose with no effect (mg/kg bw/day)	Lowest dose with effect (mg/kg bw/day)	Reference
Sprouts of *S. tuberosum* (dried) admixed to chow as powder; no trypsin inhibitor activity [(a)]	**Wistar rats** (M) No/sex/group: 6 Diets: GA at 0, 60, 120, 180, 240 or 300 mg/kg (equivalent to: 0, 7.2, 14.4, 21.6, 28.8 or 36 mg/kg bw per day [(b)]) Duration: 28 days	No effects	36		Tuśnio et al. (2013a)
Protein concentrate of *S. tuberosum*, admixed to chow as powder; with trypsin inhibitor activity [(a)]	**Wistar rats** (M) No/sex/group: 6 Diet: GA at 0, 116, 125, 164, 215, 331 or 439 mg/kg diet (equivalent to 0, 13.9, 15, 19.7, 25.8, 39.7 or 52.7 mg/kg bw per day [(b)]) Duration: 24 days	↓ relative weight of stomach ↑ relative weight of intestine ↑ serum ALT	 25.8 39.7	13.9 39.7 52.7	Tuśnio et al. (2013a)
S. tuberosum; ripe or green as chow [(a)] 287.6 mg TGA/kg fw of green potato; 82.5 mg TGA/kg fw of normal potato	**Rabbits** (sex n.r.) No/group: 10 Diet: Dose of TGA: 21.5 mg (ripe) or 64.2 mg (green)/kg diet; Equivalent to 21.6 and 82.3 mg/kg bw per day [(e)] Equivalent to 4.8 and 14 mg/kg bw per day [(c)] NO CONTROL Duration: 30 days	↓ bw and relative heart weight ↑ relative liver weight ↑ plasma glucose and Ca^{2+} concentration ↓ Na^{+}, K^{+} and protein concentration in plasma		82.3 [(d)] 82.3 [(d)] 82.3 [(d)] 82.3 [(d)]	Azim et al. (1982)

S. tuberosum; ripe or green as chow [(a)] 203.8 mg TGA/kg of green potato; 75 mg TGA/kg TGA in normal potato	**Rabbits** (sex n.r.) No/group: 4 Diet: Dose of GA: 14–25 or 48–59 mg/kg bw per day NO CONTROL Duration: 20 days	↓ bw ↑ fecal protein excretion ↓ protein digestibility		48–59 [(d)] 48–59 [(d)] 48–59 [(d)]	Azim et al. (1983)
S. tuberosum; ripe or green as chow [(a)]	**Rabbits** (sex n.r.) No/group: 5 Diet: Dose of TGA: 16.1–17.9 mg (ripe) or 75–75.4 mg (green)/kg bw per day NO CONTROL Duration: 45 days	↓ RBC counts and HGB concentration		75.2 [(d)]	Azim et al. (1984)
Strain B5141-6, diced and admixed to the diet [(a)]	**Rhesus monkey**, mated but not pregnant (F) No/sex/group: 4 Diet: 3.47 mg TGA /kg bw per day NO CONTROL Duration: 25 days	No effects	3.47 (0.87 α-solanine)		Swinyard et al. (1973)

F: female. M: male. bw: body weight. n.r.: not reported. RBC: red blood cells. HGB: haemoglobin.
(a): A non or only partly purified extract or material for which the GA composition and concentration is given
(b): Applying EFSA default values.
(c): For the exposure estimates, a live weight of 2 kg and a daily feed intake of 75 g/kg bw were assumed (derived from Carabano and Piquer, 1998).
(d): Significant when compared to lower dose group.
(e): Taking bw and food intake data from Azim et al. (1983, 1984).

1647 **Table 13.** Repeated oral toxicity studies with glycoalkaloids (GAs) (α-tomatine, α-solasonine) and aglycones (tomatidine, solasodine) from edible parts of *S.*
1648 *lycopersicum* or *S. melongena*.

Test compound	Species Experimental design Doses	Observed effects	Highest dose with no effect (mg/kg bw per day)	Lowest dose with effect (mg/kg bw per day)	Reference
α-Tomatine (given as hydrochloride) (d)	**Rats** (M) No/sex/group: 5 Gavage of 250 mg/kg bw per day Duration: 5 days		250		Wilson et al. (1961)
α-Tomatine (given as hydrochloride) (d)	**Albino rats** (F) No/sex/group: 5 Diet: 0.0025–0.04% (equivalent to 1.25–20 mg/kg bw per day (e)) Duration: 200 days		20		Wilson et al. (1961)
α-Tomatine (c)	**Albino rats** (M) No/sex/group: 6 Diet: 0, 0.5 or 2% (equivalent to 0, 600 or 2,400 mg/kg bw per day (e)) Diets were supplied with 1% cholesterol Duration: 7 days	↓ intrahepatic cholesterol		600	Cayen (1971)
α-Tomatine (c)	Albino rats (M) No/sex/group: 6–9 Diet: 0 or 1% (equivalent to 0 or 1,200 mg/kg bw per day (e)) Duration: 14 days	↓ uptake of dietary cholesterol by liver ↑ hepatic and intestinal cholesterol synthesis ↑ fecal sterol excretion		1,200 1,200 1,200	Cayen (1971)

		without affecting bile acid excretion. ↓ serum HDL, LDL and triglyceride levels		1,200	
α-Tomatine [(c)]	**Swiss-Webster mice** (F) No/sex/group: 8 Diet: 0 or 2.4 mmol/kg diet (equivalent to 0 or 496 mg/kg bw per day [(e)]) Duration: 7 days	No effect on bw or liver weight	496		Friedman et al. (1996)
α-Tomatine [(c)]	**Golden hamster** (M) No/sex/group: 6–8 Animals kept on a high-fat and high-cholesterol diet Diet: 0, 0.05, 0.1 or 0.2% in diet (equivalent to 0, 30.5, 61 or 122 mg/kg bw per day [(f)]) Duration: 21 days	↓ LDL cholesterol ↓ sum of VLDL-, LDL- and HDL-cholesterol ↓ triglyceride ↑ fecal sterol and coprostanol excretion	61 61 61	122 122 122 30.5	Friedman et al. (2000a)
α-Solasonine [(a)]	**Swiss-Webster mice** (F) No/sex/group: 8 Diet: 0 or 2.4 mmol/kg diet (equivalent to 0 or 424.4 mg/kg bw per day [(e)]) Duration: 7 days	↑ bw gain		424.4	Friedman et al. (1996)
Tomatidine [(b)]	**Rats** (M) No/sex/group: n.r.		20		Wilson et al. (1961)

	Diet: 0.0025–0.04% (equivalent to 1.25 to 20 mg/kg bw per day [e] Duration: 200 days				
Tomatidine [c]	**Swiss-Webster mice** (F) No/sex/group: 8 Diet: 0, 1.2, 2.4 or 4.8 mmol/kg diet (equivalent to 0, 99.8, 199.5 or 399 mg/kg bw per day [e]) Duration: 7, 14 or 28 days	↓ food intake (day 7) ↑ absolute and relative liver weight (day 7) ↑ absolute and relative liver weight (day 14, 28)	199.5 (no value for 99.8) 199.5	399 199.5 399	Friedman et al. (1996)
Tomatidine [a]	**Swiss-Webster mice** (F) No/sex/group: 14 Diet: 0 or 2.4 mmol/kg diet (equivalent to 199.5 mg/kg bw per day [d]) Duration: 14 days	↓ bw gain ↑ relative liver weight		199.5 199.5	Friedman et al. (2003)
Solasodine [c]	**Swiss-Webster mice** (n.r.) No/sex/group: 4–5 Diet: 0, 50, 100, 200, 400, 800 or 1,600 mg/kg diet (equivalent to 10, 20, 40, 80, 160 or 320 mg/kg bw per day [e]) Duration: 7 and 14 days	↓ bw gain (day 7 and 14) Gastric gland degeneration Cholangiohepatitis ↑ relative liver weight ↑ serum GOT, GPT, bilirubin	80 80 80 80 80	160 160 160 160 160	Friedman (1992)
Solasodine [a]	**Swiss-Webster mice** (F) No/sex/group: 8–16	↓ food intake (day 7) ↑ absolute and relative liver weight (day 7, 14, 28)	198.6 99.3	397.2 198.6	Friedman et al. (1996)

	Diet: 0, 1.2, 2.4 or 4.8 mmol/kg diet (equivalent to 0, 99.3, 198.6 or 397.2 mg/kg bw per day [e]) Duration: 7, 14 or 28 days				
Solasodine [a]	**Swiss-Webster mice** (F) No/sex/group: 24 Diet: 0 or 2.4 mmol/kg diet (equivalent to 0 or 198.6 mg/kg bw per day [e]) Duration: 14 days	↓ bw gain ↑ absolute and relative liver weight		198.6 198.6	Friedman et al. (2003)

F: female. M: male. n.r.: not reported. bw: body weight. HDL: high-density lipoprotein. LDL: low-density lipoprotein. VLDL: very low-density lipoprotein. GOT: glutamic oxaloacetic transaminase. GPT: glutamic pyruvic transaminase.
(a): Chemical standard obtained from a commercial supplier, with high purity (95% pure or higher).
(b): Chemical standard obtained by in-house isolation from plant material, and checked for purity (95% pure or higher).
(c): Chemical standard obtained from a commercial supplier, may contain α-dehydrotomatine as an impurity.
(d): Chemical standard obtained by in-house isolation from plant material, may contain α-dehydrotomatine as an impurity
(e): Applying EFSA default values
(f): A mean body weight of 130 g and daily consumption of 8 g per animal were assumed; taken from Gattermann et al. (2002).

3.1.2.3. Developmental and reproductive toxicity studies

3.1.2.3.1. Developmental effects

In 1972, Renwick published a hypothesis that there might be a causal relationship between anencephaly and spina bifida and maternal consumption of blighted potatoes. He further proposed that this may be preventable by avoiding specific, at the time unidentified, compounds from potato tubers (Renwick, 1972).

3.1.2.3.1.1. GAs from edible parts of *S. tuberosum*

The studies identified are described below and details given in **Table 14**.

Blighted potatoes

After Renwick (1972) had raised the hypothesis above, a number of studies were published using material from blighted potatoes for exposure (Poswillo et al., 1972, 1973a,b; Chaube et al., 1973; Allen et al., 1977; Sharma et al., 1978). The common limitation of these studies is that no reference to the GA content of the test material is given. The studies were therefore not used for further evaluation.

α-Solanine

Ruddick et al. (1974) performed a teratology study for α-solanine and α-chaconine in Wistar rats. They performed a comparative study with uncharacterised raw material and purified GAs. The three experimental arms investigating the teratogenic potential of α-solanine included an experiment in which first nine litters each were examined from primipari mothers, which were exposed by gavage during gestational day (GD) 6–15 at doses of 0.3, 1 and 3 mg/kg bw per day. In the second experiment, 9 litters from mothers, exposed with 6 mg/kg bw per day during GD7–10 were examined, and in the third experimental arm 3–4 primipari pregnant Wistar rats were treated by gavage with 2, 10 and 25 mg α-solanine/kg bw per day. Three–four litters were examined in this experiment. In each experiment, animals only treated with the vehicle of GAs served as control. At GD22, females were sacrificed and the following parameters were evaluated: corpora lutea, resorption sites, litter size and weight, as well as potential gross visceral and skeletal anomalies. One out of 117 offspring animals of the 3 mg/kg bw per day group showed craniorachischisis and exopthalmos, and one out of 108 of the 0.3 mg/kg bw per day group showed twisted pelvic limbs and absent tail. The authors did not judge these effects as treatment related. Besides the effects detectable in these two animals no evidence for teratogenicity was observed.

Renwick et al. (1984) purified α-solanine and α-chaconine from sprout material of the British cultivar 'Arran Pilot'. They performed a comparative oral gavage study in pregnant Syrian hamsters with poorly characterised extract material and with pure compounds. For the latter, pregnant dams were exposed by gavage on either GD7.5 or GD8 to α-solanine at a dose of 200 mg/kg bw per day on GD8. On day 15 the dams were sacrificed and necropsied. Maternal death was observed for 3 dams, whereas no dam died in the control group. Examination of fetuses revealed 59% of litters with malformed fetuses. A total of 35 cases of cranial blebs and 39 cases of exencephaly were identified, which amounted to 23% of fetuses with malformation associated with the development of brain structures. Only one fetus in more than 300 fetuses investigated in the control group showed this phenotype.

Gaffield and Keeler (1996a) performed a comparative study investigating the potential teratogenic effects of GAs and some of their aglycones. Pregnant female Syrian hamsters were gavaged on GD8. Syrian hamsters were exposed to α-solanine at a dose of 243 mg/kg bw per day on GD8. Dams were sacrificed and necropsied on GD15. The dose of 243 mg/kg bw per day of α-solanine produced 21% of

abnormal litters and 5.5% of abnormal fetuses. The rate of resorption was increased. Abnormalities were exencephaly, encephalocoele and anophthalmia.

α-Chaconine

Ruddick et al. (1974) performed a teratology study for α-solanine and α-chaconine in Wistar rats. They performed a comparative study with uncharacterised raw material and purified GAs. In the experimental arms investigating the teratogenic potential of α-chaconine (in-house prepared standard) four litters from primiparity mothers were investigated following treatment of mothers by gavage during GD6–15 with 1.5 mg α-chaconine/kg bw per day. Vehicle treated animals served as a control. At GD22, females were sacrificed and the following parameters were evaluated: corpora lutea, resorption sites, litter size and weight, as well as potential gross visceral and skeletal anomalies. No evidence for teratogenicity was observed in this experiment.

In the above-mentioned study by Renwick et al. (1984), pregnant Syrian hamsters dams were gavaged with α-chaconine at a dose of 165 and 180 mg/kg bw per day on GD8. On day 15 the dams were sacrificed and necropsied. Maternal death was observed for 4 and 6 dams respectively, whereas no dam died in the control group. Examination of fetuses revealed 27 cases of cranial blebs and 19 cases of exencephaly in the 165 mg/kg bw per day exposure group, and 10 cases of cranial blebs and 4 cases of exencephaly in the higher dose group, which amounts to 23% and 18% of fetuses, respectively, with malformation in the central nervous system. Overall, the percentage of malformed litters as of live litters was lower in the 165 mg/kg bw per day dose group (63 %) than in the 180 mg/kg bw per day dose group. Only one fetus in more than 300 fetuses investigated in the control group showed this phenotype.

In the Gaffield and Keeler (1996a) study, pregnant female Syrian hamsters were gavaged with α-chaconine at a dose of 179 or 247 mg/kg bw per day on GD8. Dams were sacrificed and necropsied on GD15. The dose of 247 mg/kg bw per day of α-chaconine (n = 1 litter) resulted in an abnormal litter. The dose of 179 mg/kg bw per day of α-chaconine produced 20% of abnormal litters and 2.7% of abnormal fetuses. The rate of resorption was increased. Abnormalities were encephalocoele and anophthalmia.

Solanidine N-oxide

In the Gaffield and Keeler (1996a) study, pregnant female Syrian hamsters were gavaged with solanidine N-oxide at doses of 58, 120 or 178 mg/kg bw per day on GD8. Solanidine N-oxide was reported by the authors as a putative metabolite of solanidine. Dams were sacrificed and necropsied on GD15. The dose of 178 mg/kg bw per day of solanidine N-oxide produced 50% of abnormal litters and 16% of abnormal fetuses. The rate of resorption was increased. The two other doses were without effects. Abnormalities were exencephaly and encephalocoele.

Solanidine

Gaffield and Keeler (1993) performed a comparative study based on the hypothesis of C_5, C_6 unsaturation being a key structural factor in steroidal alkaloid-induced mammalian teratogenesis. Pregnant female Syrian hamsters were gavaged with solanidine at a dose of 176 mg/kg bw per day on GD8. Dams were sacrificed and necropsied on GD15. The dose of 176 mg/kg bw per day produced 50% of abnormal litters and 24% of abnormal fetuses. Abnormalities were exencephaly, encephalocoele, anophthalmia and external viscera.

Gaffield and Keeler (1996a) studied solanidine again. Pregnant female Syrian hamsters were gavaged with solanidine at a dose of 115 mg/kg bw per day on GD8 and dams were sacrificed and necropsied on GD15. The dose of 115 mg/kg bw per day produced 19% of abnormal litters and 2.3% of abnormal fetuses. Abnormalities were encephalocoele and anophthalmia.

Potential teratogenic effects of solanidine were also investigated by Friedman et al. (2003a). For the experiment the authors purchased time pregnant Swiss Webster mice, many of which apparently were not pregnant. Following acclimatisation, exposure to experimental chow was started on GD4 and terminated either at GD17 or GD18. Exposure to 205 mg solanidine/kg bw per day resulted in reduced maternal body weight, probably caused by reduced maternal feed intake. In maternal animals, an increased maternal relative liver weight was observed in addition. For the offspring, the authors identified reduction of litter size, litter weight, litter weight/body weight and average fetus weight in response to treatment.

Demissidine

In an earlier study by Gaffield and Keeler (1993), pregnant female Syrian hamsters were gavaged with demissidine at a dose of 176 mg/kg bw per day on GD8, and this dose produced 14% of abnormal litters and 3% of abnormal fetuses. Both effects did not reach the level of statistical significance. Abnormalities were not specified.

In a later study by Gaffield and Keeler (1996a), pregnant female Syrian hamsters were gavaged with demissidine at a dose of 176 mg/kg bw per day on GD8. The dose of 176 mg/kg bw per day produced 14% of abnormal litters and 3% of abnormal fetuses. Abnormalities were exencephaly, encephalocoele, cebocephaly, cleft palate and anophthalmia.

3.1.2.3.1.2. GAs from edible parts of food plants other than *S. tuberosum*

The studies identified are described below and details given in **Table 15**.

α-Tomatine

In 1990 Keeler et al. (1990) published a review on existing data on congenital craniofacial malformation following exposure to spirosolane-containing solanum species. To this review some original data were added, among them, on α-tomatine. Pregnant Simonsen hamsters were gavaged with 530 mg/kg bw per day of α-tomatine on GD8. 19 dams died from overdose symptoms largely related gastrointestinal tract lesions. From surviving dams, a percentage of 6% deformed litters was recorded.

Tomatidine

Keeler et al. (1976) investigated potential teratogenic effects of tomatidine and exposed pregnant Syrian hamsters on GD8 with 1,184 or with 2,664 mg/kg bw per day of tomatidine by gavage. Neither abnormal litters nor abnormal offspring, which was used as readout, was reported.

In Swiss Webster mice potential teratogenic effects of tomatidine were investigated by Friedman et al. (2003a) following the same scheme as that described for solanidine above. Exposure to 167 mg/kg bw per day of tomatidine resulted in reduced maternal body weight and increased maternal relative liver weight in response to treatment. No effects were detectable which affected offspring animals.

Solasodine

Keeler et al. (1976) comparatively studied the consequences of solasodine exposure at GD7 (1,184, 1,480 and 1,628 mg/kg bw per day) and at GD8 (1,184 and 1,480 mg/kg bw per day) in pregnant female Syrian hamsters (n = 115). The authors reported that a total of 10 dams distributed over all dose groups died from overdose symptoms, except for the 1,480 mg/kg bw per day dose group if exposed on GD7. For the remaining dams the major effect was abnormal litters, which did not follow a clear dose dependency if exposed at GD7 (25–29%). Following exposure on GD8 the percentage of abnormal litters increased dose dependently in a statistically significant manner from 36% (1,184 mg/kg bw per day) to 44% (1,480 mg/kg bw per day). The most often detected phenotypes in abnormal offspring comprised spina bifida, exencephaly and cranial bleb. The percentage of abnormal offspring was 5–7% if exposure occurred on GD7, and 6–15 % following exposure on GD8.

In the study by Gaffield and Keeler (1993) pregnant female Syrian hamsters were gavaged with solasodine at a dose of 1,400 mg/kg bw per day on GD8, that dose produced 75% of abnormal litters and 29% of abnormal fetuses. Abnormalities were encephalocoele, exencephaly and anophthalmia.

Using Swiss Webster mice Friedman et al. (2003) reported that exposure to 195 mg solasodine/kg bw per day resulted in reduced maternal body weight, probably caused by reduced maternal feed intake. In maternal animals an increased maternal relative liver weight was observed in addition. For the offspring authors identified reduction of litter size, litter weight, litter weight/body weight and average fetus weight in response to treatment.

Dihydrosolasodine (soladulcidine)

Gaffield and Keeler (1993) also studied the effects of dihydrosolasodine gavaged to pregnant female Syrian hamsters on GD8 at a dose of 1,400 mg/kg bw per day. That dose produced 45% of abnormal litters and 6% of abnormal fetuses. Abnormalities were encephalocoele, exencephaly, cebocephaly and anophthalmia.

3.1.2.3.1.3. Comparative evaluation of studies

To make use of the studies in which single doses have been used, studies are evaluated comparatively regarding relative potency of compounds. This approach represents rather a qualitative than a quantitative assessment. Only hamsters, for which the widest sets of data are available, have been used. Further, in the comparative studies, authors used similar molar exposure levels. However, these exposure scenarios are mostly only identical within a study but not across studies. Further, in order to be able to compare potency, endpoints which have been investigated across studies have to be selected. Finally, a reference point for comparison would be helpful, but is not available.

Keeler et al. (1990) were the first to take the attempt to order GAs according to their teratogenic potency. They took the same data base and some additional data to establish a relative potency list of GAs (Gaffield and Keeler, 1996b). This list contained data on GAs from Brown and Keeler (1978), Keeler et al. (1976), Renwick et al. (1984), Keeler et al. (1990), Gaffield and Keeler (1993) as well as Gaffield and Keeler (1996a). In this assessment, the authors aimed to compare relative teratogenic potencies of jervine, solanidine and spirosolane alkaloids. Relative potencies can only be compared by their induction of terata if a molar equivalent basis of exposure is applied (see **Appendix D**) or if effects are allocated to an ascending exposure level in mg/kg bw. The data which were consistently reported across studies were percentage of abnormal litters and percentage of abnormal fetuses. As can be seen from Table D.1. **(Appendix D)** a clear relative potency pattern cannot be delineated. It can also be seen, that there are huge differences between values from different laboratories, e.g. for α-chaconine and α-solanine, if results from the studies of Renwick et al. (1984) and from Gaffield and Keeler (1996a) are compared (see **Appendix D**). Further, the relative order of potency reported by Gaffield and Keeler (1996b) may only be used for a qualitative comparison, according to the authors. However, α-chaconine may be misplaced in this order, because at dosage 0.29 mmol/kg maternal lethality became apparent, not allowing to follow up the fate of the offspring.

3.1.2.3.1.4. Summary on developmental effects

The experimental developmental studies indicate that effects occur at doses of 165 mg/kg bw per day and above for GAs, and of 115 mg/kg bw per day and above for the aglycones. Amongst others, these effects to a large majority were observed in the central nervous system and most frequently comprised exencephaly, encephalocele and anophthalmia.

In comparative single dose assessment studies, the results suggest differences in the potency of GAs and their aglycones. However, due to limitations in the studies available (e.g. lack of dose-response study design), an order of potency cannot be established.

3.1.2.3.2. Reproductive effects

Kline et al. (1961) investigated the effects of α-solanine on the survival of rat pups (see **Table 16**). They exposed pregnant female Holtzman rats (duration not given, starting exposure following detection of pregnancy by weighing until delivery) to either commercial α-solanine at 3.6 or 4.8 mg/kg bw per day or to α-solanine, isolated from the potato sprouts (purity unknown) at 3.6 mg/kg bw per day. Compared to controls, all treatment procedures produced a decrease in the percentage of weaned pups and a comparable increase in the number of zero litters (litters in which all pups died). No dose-dependency was seen for the commercial α-solanine. However, the effects were more pronounced when applying the isolated α-solanine (23.6% weaned pups compared to controls) than the commercial α-solanine (37.5% for both dosages, compared to controls). Starvation may have contributed to the early death of the pups as no milk was found in the stomach of these animals, pointing to a lactational deficit. Cross fostering studies were performed but results were inconclusive. Adult males were investigated as controls and no noticeable effects were detectable (Kline et al., 1961).

Effects of solasodine on fertility were also investigated in male Rhesus monkeys (Dixit et al., 1989). Ten Rhesus monkeys were divided into two groups of five animals each, which were either treated for 150 days by placebo or 100 mg/kg bw per day of solasodine encapsulated into a banana (see **Table 17**). On day 151 testes and epididymis were surgically removed. Sperm count and motility were assessed in the cauda epididymis. Pathological parameters were assessed from tissue section of paraffin embedded material. Biochemical parameters were measured from frozen tissue samples. Interstitial testicular cells, like Leydig cells and their precursors were counted. 100 mg/kg bw per day reduced testis weight, weight of the epididymis, the diameter of the seminiferous tubule and the Leydig cell nuclear diameter. In terms of fertility parameters, the authors detected a reduced number of spermatids, spermatozoa, immature and mature Leydig cells. The number of degenerating Leydig cells was found to be unaltered. Analysis of the cauda of the epididymis revealed a dramatic reduction of sperm density and sperm motility. On the biochemical level total testicular protein content, content of sialic acid and of glycogen were reduced, whereas testicular testosterone levels were elevated. The CONTAM Panel notes that treatment of Rhesus monkeys with 100 mg/kg bw per day of solasodine encapsulated into a banana affected testicular function and male reproduction.

The impact of solasodine following oral application on male fertility was investigated in dogs (Gupta and Dixit, 2002). Five dogs were treated with 80 mg/kg bw per day of solasodine with the compound embedded in a capsule of mutton meet (see **Table 18**). In this study, only effects on the epididymis were investigated. The treatment did not affect the body weight of the animals but decreased the epididymal weight and cauda epididymal epithelial height. The epididymal lumen was found to be empty of sperm. On a biochemical level the total protein content, content of sialic acid and of glycogen were found to be reduced in the epididymis in response to 80 mg/kg bw per day of solasodine. Epidydimal levels of cholesterol were found to be elevated. In conclusion, oral treatment of dogs with 80 mg/kg bw per day of solasodine impacts on the integrity and the function of the epididymal gland in male dogs. The epididymal findings in this study using oral dosage are in line with a previous study which investigated effects of solasodine on male reproductive organs following i.p. application of 20 mg/kg bw per day (Dixit and Gupta, 1982).

In summary, reduced postnatal survival of pups due to insufficient milk production was found following exposure of pregnant Holtzman rats to α-solanine. Doses of approximately 3.6 mg/kg bw per day from different sources of α-solanine resulted in a percentage of successfully weaned pups between 23.6% and 37.8% if compared to controls. The observed decrease most likely is due to lactational failure of dams and results in the death of the pups by starvation. The CONTAM Panel noted the limitations in the description of the study design.

Effects on fertility were observed following exposure of male dogs to 80 mg/kg bw per day of solasodine. Following oral exposure, a decreased epididymal weight and cauda epididymal epithelial height as well as an epididymal lumen depleted of sperm was found. These findings are supported by observations in dogs by the same authors following i.p. injection of 20 mg/kg bw per day of solasodine, in which in addition to the epididymal effects a decreased weight of testes, a reduction of seminiferous tubule diameter and reduced nuclear diameter of Leydig cells was observed.

1885 **Table 14.** Developmental toxicity studies on glycoalkaloids (GAs) (α-solanine, α-chaconine) and aglycones (solanidine, demissidine) from edible parts of *S.*
1886 *tuberosum*.

Test compound	Species, experimental design and doses	Observed effects	Highest dose with no effect (mg/kg bw per day)		Lowest dose with effect (mg/kg bw per day)		Reference
			Mother	Offspring	Mother	Offspring	
α-Solanine							
α-Solanine [a]	**Wistar rats** Gavage Exposure at different periods during gestation, sacrifice on day 22. (I) Exposure during GD6–15: 0, 0.3, 1, 3 mg/kg bw per day (9 or 10 litters) (II) Exposure during GD7–10: 0, 6 mg/kg bw per day (9 litters) (III) Exposure during GD8–11: 0, 2, 10, 25 mg/kg bw per day (2, 4, 3, 4 litters, respectively)	all exposure groups: No significant effects	No effects				Ruddick et al. (1974)

1887

α-Solanine [b] (purified from sprouts of the British cultivar Arran Pilot)	**Syrian hamsters** Number of dams not specified, 85 litters and 964 fetuses in control, 55 litters and 573 fetuses in α-solanine group. Gavage on GD7.5 or GD8 only 200 mg/kg bw per day Sacrifice on GD15	maternal death (3 dams) 59% malformed litters 23% CNS malformed fetuses (gavage GD8)		200		200 200	Renwick et al. (1984)
α-Solanine [b]	**Pregnant syrian hamsters** No/group: 24 Gavage on GD8 0.28 mmol/kg bw (equivalent to 243 mg/kg bw per day) Sacrifice on GD15	21% abnormal litters 5.5% abnormal fetuses (exencephaly, encephalocele, anophthalmia) ↑ maternal fatality				243 243 243	Gaffield and Keeler (1996a)
α-Chaconine							
α-Chaconine [d]	**Wistar rats** Gavage on GD6–15 Exposure GD6–15: 1.5 mg/kg bw per day (4 litters) Sacrifice on day 22	no statistically significant effect					Ruddick et al. (1974)

α-Chaconine [b] Purified from sprouts of the British cultivar Arran Pilot	**Syrian hamsters** Number of of dams not specified, 85 litters and 964 fetuses in control, 60 litters and 645 fetuses in the α-chaconine groups, no specified how many animals per dosage group Gavage on GD8 only 0, 165, 180 mg/kg bw per day	maternal death (4 of 6 dams) 63% malformed litters 23% CNS malformed fetuses		165		165 165	Renwick et al. (1984)
α-Chaconine [b]	**Pregnant Syrian hamsters** No/Group: 15 and 1 litter Gavage on GD8 0.21, 0.29 mmol/kg bw (equivalent to 179 and 247 mg/kg bw) Sacrifice on GD15	maternal lethality (3/4) 20% abnormal litters 2.7% abnormal fetuses (exencephaly and encephalocele)		247		179 179	Gaffield and Keeler (1996a)
Solanidine							
Solanidine N-oxide [c]	**Pregnant Syrian hamsters** No/group: 3–14 litters Gavage on GD8 0.14, 0.29, 0.43 mmol/kg bw (equivalent to 58, 120, 178 mg/kg) Sacrifice on GD15	50% abnormal litters 16% abnormal fetuses (exencephaly and encephalocele)		120 120		178 178	Gaffield and Keeler (1996a)

Solanidine [c]	**Pregnant Syrian hamsters** No/group: 12 litters Gavage on GD8 0.44 mmol/kg bw (or 176 mg/kg bw) [e] Sacrifice on GD15	50% abnormal litters 24% abnormal fetuses (exencephaly, encephalocele, anophthalmia, external viscera)				176 176	Gaffield and Keeler (1993)
Solanidine [b]	**Pregnant Syrian hamsters** No/group: 21 litters Gavage on GD8 0.29 mmol/kg bw (equivalent to 115 mg/kg bw) Sacrifice on GD15	19% abnormal litters 2.3% abnormal fetuses (encephalocele and anophthalmia)				115 115	Gaffield and Keeler (1996a)
Solanidine [a]	**Swiss Webster mice** (time pregnant/time mated (not clear)) No/group: 20 pregnant females Diet. Exposure: Start GD4, Necroscopy GD17–18 2.4 mmol/kg diet (equivalent to 159 mg/kg bw per day)	↓ maternal feed intake ↑ maternal relative liver weight ↓ litter size ↓ litter weight ↓ litter weight/body weight ↓ average fetus weight			159 159 159	159 159 159	Friedman et al. (2003a)
Demissidine							
Demissidine [c]	**Pregnant Syrian hamsters** No/group: 14 litters Gavage on GD8	14% abnormal litters 3% abnormal fetuses				176 176	Gaffield and Keeler (1993)

	0.44 mmol/kg bw (or 176 mg/kg bw) [e] Sacrifice on GD15	(both values not statistically significant)					
Demissidine [c]	**Pregnant Syrian hamsters** No/group: 14 litters Gavage on GD8 0.44 mmol/kg bw (equivalent to 176 mg/kg bw) Sacrifice on GD15	14% abnormal litters 3% abnormal fetuses (both values not statistically significant)				176 176	Gaffield and Keeler (1996a)

GD: gestational day. MW: molecular weight (in g/mol). CNS: central nervous system.
(a): Chemical standard obtained from a commercial supplier, with high purity (95% pure or higher).
(b): Chemical standard obtained by in-house isolation from plant material, and checked for purity (95% pure or higher).
(c): Chemical standard obtained by in-house synthesis, and checked for purity (95% pure or higher).
(d): Poorly characterised chemical standard (e. g. purity or composition not defined).
(e): Doses as reported by the authors in the original publication.

1894 **Table 15.** Developmental toxicity studies on glycoalkaloids (GAs) (α-tomatine) and aglycones (tomatidine, solasodine, dihydrosolasodine) from edible parts of
1895 food plants other than *S. tuberosum*.

Test compound	Species, experimental design and doses	Observed effects	Highest dose with no effect (mg/kg bw per day)		Lowest dose with effect (mg/kg bw per day)		Reference
			Mother	Offspring	Mother	Offspring	
α-Tomatine							
α-Tomatine [(d)]	**Pregnant Simonsen hamsters** No/groups: 34 dams Gavage on GD8 0, 530 mg/kg bw per day Sacrifice on GD15	Maternal death 6% deformed litters from surviving dams			530	530	Keeler et al. (1990)
Tomatidine							
Tomatidine [(c,e)]	**Pregnant Syrian hamsters** No/groups: 15–24 pregnant dams Gavage on GD8 0, 1,184, 2,664 mg/kg bw per day Sacrifice on GD15	No effects	2,664	2,664			Keeler et al. (1976)
Tomatidine [(e)]	**Swiss Webster** time pregnant/time mated (not clear) mice. No/group: 23 pregnant mice Diet. Exposure GD4–18 2.4 mmol/kg diet (equivalent to 167 mg/kg bw per day)	↓ maternal bw ↑ maternal relative liver weight			167		Friedman et al. (2003a)
Solasodine							

Solasodine [c]	**Pregnant Syrian hamsters** No/group: 11–50 dams per group 0, 1,184, 1,480 and 1,628 mg/kg bw per day gavage on GD7, sacrifice on GD15 0, 1,184 and 1,480 mg/kg bw on GD8, sacrifice on GD15	<u>GD7 exposure:</u> Maternal death 25% abnormal litters 5% abnormal offspring <u>GD8 exposure</u>: Maternal death 36% abnormal litters 6% abnormal offspring			1,184 1,184	 1,184 1,184 1,184 1,184	Keeler et al. (1976)
Solasodine [c]	**Pregnant Syrian hamsters** No/group: 8 litters Gavage on GD8 3.4 mmol/kg bw (or 1,400 mg/kg bw) [f] Sacrifice on GD15	75% abnormal litters 29% abnormal fetuses (encephalocele, exencephaly, anophthalmia)				1,400 1,400	Gaffield and Keeler (1993)
Solasodine [a]	**Swiss Webster** time pregnant/time mated (not clear) mice. No/group: 45 pregnant females Diet. Exposure GD4–18 2.4 mmol/kg diet (equivalent to 156 mg/kg bw per day)	↓ maternal bw ↓ maternal feed intake ↑ maternal relative liver weight ↓ litter weight ↓ litter weight/bw ↓ average fetus weight			156 156 156	 156 156 156	Friedman et al. (2003a)
Dihydrosolasodine							

Dihydrosolasodine [(b)] (Soladulcidine)	**Pregnant Syrian hamsters** No/group: 11 litters Gavage on GD8 3.4 mmol/kg bw (1,400 mg/kg bw [(f)]) Sacrifice on GD15	45% abnormal litters 6% abnormal fetuses (encephalocele, cebocephaly, exencephaly, anophthalmia)				1,400 1,400	Gaffield and Keeler (1993)

GD: gestational day. MW: molecular weight (in g/mol).
(a): Chemical standard obtained from a commercial supplier of high purity (95% pure or higher).
(b): Chemical standard obtained by in-house synthesis, and checked for purity (95% pure or higher).
(c): Chemical standard obtained from a commercial supplier, purity not stated.
(d): Chemical standard obtained from a commercial supplier, may contain dehydrotomatine as an impurity.
(e): Chemical standard obtained from a commercial supplier, may contain dehydrotomatidine as an impurity.
(f): Doses as reported by the authors in the original publication.

1903 **Table 16.** Reproductive studies on glycoalkaloids (GAs) (α-solanine) from edible parts of *S. tuberosum*.

Test compound	Species Experimental design Doses	Observed effects	Highest dose with no effect (mg/kg bw per day)		Lowest dose with effect (mg/kg bw per day)		Reference
			Mother	Offspring	Mother	Offspring	
α-Solanine							
α-Solanine (commercial) (a) α-Solanine (isolated from sprouts) (a)	Female pregnant **Holtzman rats** N = 10/4 litters per group for testing of commercial/ isolated α-solanine Duration of exposure not clear (assumption that the study duration was 14 days) Diet: 0, 30, 40 mg α-solanine (commercial) per kg chow (0, ≈3.6, ≈4.8 mg/kg bw per day) Diet: 0, 30 mg α-solanine (isolated) per kg chow (≈3.6 mg/kg bw per day)	↑ number of litters in which pups died within the first three days ↓ percentage of weaned pups (37.5% weaned pups, compared to controls) ↓ percentage of weaned pups (23.6% weaned pups, compared to controls)				3.6 mg/kg bw per day (for commercial and isolated α-solanine) 3.6 mg/kg bw per day (for commercial α-solanine) 3.6 mg/kg bw per day (for isolated α-solanine)	Kline et al. (1961)

1904 (a): Poorly characterised chemical standard (purity and composition not defined, may have contained α-chaconine).

1905

1906 **Table 17.** Reproductive studies on aglycones (solasodine) from edible parts of *S. melongena*.

Test compound	Species Experimental design Doses	Observed effects	Lowest dose with effect (mg/kg bw per day)	Reference
Solasodine				
Solasodine [(a)]	Captured **Rhesus monkey** (no further specifications) 0, 100 mg/kg bw per day for 150 days Oral administration by encapsulation into a banana On day 151 surgical removal of testes and epididymis	↓ testes weight ↓ epididymal weight ↓ seminiferous tubule diameter ↓ Leydig cell nuclei diameter ↓ spermatid count ↓ spermatozoa ↓ immature and mature Leydig cells ↓ sperm density and motility in cauda epididymis ↓ total protein, sialic acid, glycogen in testes ↑ cholesterol in testes	100 100 100 100 100 100 100 100 100 100	Dixit et al. (1989)
Solasodine [(a)] Isolated from berries of *S. xanthocarpum*	**Dogs** (M, 12–17 kg, 5 dogs per group) 0, 80 mg/kg bw per day Oral in a mutton meet pellet Duration: 30 days	↓ epididymal weight ↓ cauda epididymal epithelial height ↓ total protein, sialic acid, glycogen in the cauda epididymis ↑ cholesterol in the cauda epididymis	80 80 80 80	Gupta and Dixit (2002)

1907 M: males.
1908 (a): Chemical standard obtained by in-house isolation from plant material, highly purified.

3.1.2.4. Immunotoxicity studies

No standard assays for immunotoxicity of GA and aglycones could be identified.

Plant extracts of *S. tuberosum* as well as GA and aglycones exerted pronounced anti-inflammatory effects in *in vitro* and *in vivo* experiments, as outlined below (see **Section 3.1.5**). Choi and Koo (2005) applied an ethanolic extract of the peeled tuber of *S. tuberosum* to mice at an oral dose of 100 mg or 200 mg/kg bw. One hour later paw oedema was induced by carrageenan- or formalin treatment. Pre-treatment with the extract reduced significantly the oedema. The same group (Choi et al., 2005) induced arthritis by injecting Freund's adjuvant into an angle of a murine paw. Symptoms of the arthritis (oedema) were reduced when animals were pretreated orally with the ethanolic extract of *S. tuberosum* at 0, 100 or 200 mg/kg bw per day for 21 days. Similar effects of the ethanolic extract of *S. tuberosum* were reported in an autoimmune disease model in mice (Choi, 2007).

Single GAs and aglycones, e.g. α-solanine, α-chaconine and solanidine, were also effective anti-inflammatory compounds *in vivo*. For example, Shin et al. (2016) applied orally α-solanine (0, 2.5, 5 or 10 mg/kg) to mice prior to an i.p. lipopolysaccharide (LPS) injection. The pretreatment with α-solanine suppressed the expression of pro-inflammatory genes, such as tumor necrosis factor α (TNF-α) and interleukin 1 β (IL-1β), in the liver. In another study, mice received orally α-chaconine at 0.5 or 2 mg/kg bw before a single i.p. LPS injection (Lee et al., 2015). This pretreatment reduced the LPS-induced lethality and the intrahepatic production levels of several pro-inflammatory cytokines. Emmanuel et al. (2006, abstract only) applied solasodine, extracted from plant material as single gavage of 50 mg/kg bw to rats before carageenan was injected to induce inflammation. Pretreatment with solasodine reduced the swelling of the paw.

To conclude, pronounced anti-inflammatory effects were seen in rodents after single oral applications of α-solanine, α-chaconine, solasodine or extracts of *S. tuberosum*. No indication of immunotoxicity was seen in these studies.

3.1.2.5. Studies on cardiovascular effects

No studies were identified. Mechanistic studies have been identified as described in **Section 3.1.5.1.4**.

3.1.2.6. Neurotoxicity studies

No studies were identified. Mechanistic studies have been identified as described in **Section 3.1.5.1.4.**

3.1.2.7. Genotoxicity

3.1.2.7.1. GAs from edible parts of S. tuberosum

Several assays were performed to test the genotoxic potential of potato GA, the aglycone solanidine, as well as extracts of *S. tuberosum* (see **Table 18**). The studies, applying the Ames test, reported on results obtained in only two strains of *Salmonella typhimurium*, which is a major limitation regarding the quality of data.

α-Solanine was subjected to the Ames test by two independent groups, both providing negative results. Ness et al. (1984) applied 10–50 µg per plate which did not increase the number of revertants in the strains TA98 and TA100 with or without metabolic activation. Friedman and Henika (1992) performed two independent Ames assays with α-solanine. One of the assays was weakly positive in TA100 with S-9 activation but only at the highest (cytotoxic) dose. The CONTAM Panel considered this study as

negative. The same group applied several doses of α-solanine via single i.p. administration to pregnant female mice at GD15-16, which did not increase the frequency of micronuclei in the fetal blood cells (Friedman and Henika, 1992). Similarly, there was no increase in the frequency of micronuclei in peripheral blood cells of adult male mice after single i.p application of a large dose range of α-solanine (Friedman and Henika, 1992).

α-Chaconine was tested in the same assays as α-solanine (Friedman and Henika, 1992). There was a small increase in the number of revertants in TA98 without S9 activation only at a cytotoxic dose. The CONTAM Panel considered this study as negative. The micronucleus tests performed on peripheral blood cells of male mice and fetal blood cells from exposed pregnant female mice were both negative.

The aglycone solanidine was tested by Friedman and Henika (1992) at concentrations/doses being equimolar to those of α-solanine and α-chaconine and was found to be uniformly negative in all the assays, i.e. in the Ames test and in the micronucleus assays on peripheral blood cells in both, the fetuses as well as in the male adult mice.

No increase in the number of revertants was observed when 7.5 µg extract/plate of *S. tuberosum* (plant part not specified) were assayed in strains TA98 and TA100 with or without metabolic activation (Ness et al., 1984). No conclusions can be drawn from this study since no information is provided on the method of extraction and the extent of its purification.

To conclude, from the limited number of studies available there is no evidence of genotoxicity for the potato GAs α-solanine and α-chaconine, and for the aglycone solanidine.

3.1.2.7.2. GAs from edible parts of food plants other than S. tuberosum

Several assays were performed to test the genotoxic potential of aubergine GAs, α-solamargine and α-solasonine (see **Table 19**). Almeida et al. (2010) used highly purified α-solamargine, extracted from fruits of *S. palinacanthum* Dunal, in an Ames test with *S. typhimurium* strains TA98, TA97a, TA100 and TA102 with and without metabolic activation. There was some weak mutagenicity in TA98 strain in absence of metabolic activation.

Munari et al. (2012) generated an extract of *S. lycocarpum* St. Hill, which was 89% pure and contained α-solasonine and α-solamargine in a ratio of 1:1. The clastogenic potential of the extract was tested *in vitro* in V97 Chinese hamster lung fibroblasts in the absence of metabolic activation with negative results both in the Comet and chromosomal aberrations assays. Using the same type of extract, Munari et al. (2014) performed *in-vivo* assays in mice. In a first test, male Swiss mice were exposed by gavage to 0, 15, 30 or 60 mg/kg bw per day for 14 days and peripheral blood samples were analysed 48 h, 7 days and 14 days after treatment. No increase in the frequency of micronucleated polychromatic erythrocytes (PCE) was observed. In subsequent tests, male Swiss mice received the extract at 60 mg/kg bw per day by gavage for a duration of 14 days. Negative results were obtained both in the bone marrow micronucleus test and in the Comet assay.

In conclusion, from the limited number of studies available, there was no evidence for genotoxicity of the aubergine GA α-solamargine.

1985 **Table 18.** Tests for genotoxicity/mutagenicity of potato glycoalkaloids (GAs) (α-solanine, α-chaconine) and the aglycone (solanidine).

Test compound (Source)	**Test system**	**Cells/animals**	**Concentration/Treatment**	**Result**	**Comment**	**Reference**
α-Solanine [a]	Reverse mutation assay	*S. typhimurium* TA98 and 100 +/- S9	10–50 µg/plate	Negative	No positive control	Ness et al. (1984)
α-Solanine [a]	Reverse mutation assay	*S. typhimurium* TA98, TA100 +/-S9	0.07–2.3 µmol/plate [c]	Negative Positive controls: expected results		Friedman and Henika (1992)
α-Solanine [a]	Micronucleus assay (fetal blood cells)	Swiss-Webster mice (F)	Single i.p. application at GD15–16: 0.01, 0.02, 0.045 mmol/kg bw [d] (equivalent to 8.7, 17.4 or 39.1 mg/kg bw)	Negative Positive controls: expected results	Number of fetuses per dose group: 54–55	Friedman and Henika (1992)
α-Solanine [a]	Micronucleus assay (peripheral blood cells)	Swiss-Webster mice (M) No/sex/group: 8	Single i.p. application: 0.02, 0.045, 0.09 mmol/kg bw [d] (equivalent to 17.4, 39.1 or 78.1 mg/kg bw)	Negative Positive controls: expected results		Friedman and Henika (1992)
α-Chaconine [a]	Reverse mutation assay	*S. typhimurium* TA98, TA100 +/-S9	0.07–2.3 µmol/plate [c]	Negative Positive controls: expected results		Friedman and Henika (1992)
α-Chaconine [a]	Micronucleus assay (peripheral blood cells)	Swiss-Webster mice (M) No/sex/group: 8	Single i.p. application: 0.01, 0.02, 0.045 mmol/kg bw [d] (equivalent to 8.5, 17.0 or 38.3 mg/kg bw)	Negative Positive controls: expected results		Friedman and Henika (1992)
α-Chaconine [a]	Micronucleus assay (fetal blood	Swiss-Webster mice (F)	Single i.p. application at GD15–16: 0.005, 0.01, 0.02,	Negative Positive controls:	Number of fetuses per dose group: 63–65 (0.005 and 0.01	Friedman and Henika (1992)

	cells)	No of dams/sex/group: n.r.	0.045 mmol/kg bw [(d)] (equivalent to 4.3, 8.5, 17.0 or 38.3 mg/kg bw)	expected results	mol/kg)	
Solanidine [(a)]	Reverse mutation assay	*S. typhimurium* TA98, TA100 +/-S9	0.04–0.58 µmol/plate [(c)]	Negative Positive controls: expected results		Friedman and Henika (1992)
Solanidine [(a)]	Micronucleus assay (peripheral blood cells)	Swiss-Webster mice (M) No/sex/group: 8	Single i.p. application: 0.01, 0.02, 0.045 mmol/kg bw [(d)] (equivalent to 4.0, 8.0, 17.9 mg/kg bw)	Negative Positive controls: expected results		Friedman and Henika (1992)
Solanidine [(a)]	Micronucleus assay (fetal blood cells)	Swiss-Webster mice (F) No of dams/sex/group: n.r.	Single i.p. application at GD15–16: 0.01, 0.02, 0.045 mmol/kg bw [(d)] (equivalent to 4.0, 8.0, 17.9 mg/kg bw)	Negative Positive controls: expected results	Number of fetuses per does group: 46–50	Friedman and Henika (1992)
Extract of *S. tuberosum* (plant part not specified) [(b)]	Reverse mutation assay	*S. typhimurium* TA98 and 100 +/-S9	7.5 µg/plate	Negative	No positive control	Ness et al. (1984)

F: female. M: male. N.r.: not reported. GD: gestational day.
(a): Chemical standard obtained from a commercial supplier, with high purity (95% pure or higher).
(b): A non- or only partly purified extract or material for which the GA composition and concentration is given.
(c): The authors reported on the application of 0.07–2.3 mM/plate of α-solanine (equivalent to 60.7–1996.6 mg per plate) of 0.07–2.3mM/p of α-chaconine (equivalent to 59.6 to 1,959.8 mg/plate) and of 0.04–0.58 mM/plate of solanidine (equivalent to 15.9–230.7 mg per plate). The CONTAM Panel assumed a typo and the application of the compounds in µmol/plate as realistic doses.
(d): The authors reported on the application of 0.01, 0.02, 0.045, or 0.09 mol/kg bw of α-solanine (equivalent to 8.7, 17.4, 39.1, or 78.1 g/kg bw), of 0.01, 0.02 or 0.045 mol/kg bw of α-chaconine (equivalent to 8.5, 17.0 or 38.3 g/kg bw) and of 0.01, 0.02 or 0.045 mol/kg bw of solanidine (equivalent to 4.8, 8.0 or 17.9 g/kg bw). The CONTAM Panel assumed a typo and the application of the compounds in mmol/kg bw as realistic doses.

1994 **Table 19.** Tests for genotoxicity/mutagenicity of aubergine glycoalkaloids (GAs) (α-solasonine and α-solamargine).

Test compound	Test system	Cells/animals	Concentration/Treatment	Result	Comment	Reference
α-Solamargine [(a)]	Reverse mutation assay	*S. typhimurium* TA100, TA98, TA97a and TA102, +/- S9 (rat liver)	Preincubation test 0, 1.25, 2.5, 3.5, 4.5, and 5.0 mg/plate Solvent: DMSO Appropriate positive controls	Positive in TA98 without S9 from 4.5 mg/plate onwards (mutagenic index >2) Statistical increase in the number of revertants in TA100 +S9 from 2.5 mg/plate onwards, but not biologically significant (mutagenic index <2) Positive controls: expected results	The CONTAM Panel noted the limited significance of this finding.	Almeida et al. (2010)
α-Solasonine and α-solamargine (1:1) [(b)]	*In vitro* Comet assay	Chinese hamster lung fibroblasts (V79) -S9	Preliminary cytotoxicity: 1-256 µg/mL Main test: 0, 4, 8, 16, and 32 µg/mL Positive control: MMS	Cytotoxicity observed from 64 µg/mL onwards Negative MMS as positive control: expected result		Munari et al. (2012)
α-Solasonine and α-solamargine (1:1) [(b)]	*In vitro* chromosomal aberration assay	Chinese hamster lung fibroblasts (V79) -S9	Preliminary cytotoxicity: 1-256 µg/mL Main test: 0, 4, 8, 16, and 32 µg/mL Positive control: MMS Exposure: 18 h and harvesting at the end of treatment	Cytotoxicity observed from 64 µg/mL onwards Negative MMS as positive control: expected result		Munari et al. (2012)

α-Solasonine and α-solamargine (1:1) [(b)]	*In vivo* micronucleus assay (peripheral blood cells)	Swiss mice (M)	Preliminary toxicity test: 15-1,000 mg/kg bw Gavage: 0 (water),15, 30 or 60 mg/kg bw per day for 14 days Blood samples collected 48h, 7 days and 14 days after start of treatment	Doses above 60 mg/kg bw were toxic No cytotoxicity observed Negative MMS as positive control: expected results		Munari et al. (2014)
α-Solasonine and α-solamargine (1:1) [(b)]	*In vivo* micronucleus assay (bone marrow cells)	Swiss mice (M)	Gavage: 0 (water) or 60 mg/kg bw per day for 14 days Blood samples collected at end of treatment	No cytotoxicity observed Negative MMS as positive control: expected result		Munari et al. (2014)
α-Solasonine and α-solamargine (1:1) [(b)]	*In vivo* Comet assay (liver cells)	Swiss mice (M)	Gavage: 0 (water), 60 mg/kg bw per day for 14 days	No statistically significant decrease of cell viability Negative MMS as positive control: expected result		Munari et al. (2014)

MMS: Methyl methanesulfonate. DMSO: Dimethyl sulfoxide.
(a): Extracted from *S. palinacanthum* Dunal, highly purified.
(b): Extracted from *S. lycocarpum* St. Hill., 89% purity.

3.1.2.8. Carcinogenicity studies

No long-term carcinogenicity studies on any of the GAs and aglycones addressed in this opinion could be identified.

3.1.2.9. Studies on metabolic effects

3.1.2.9.1. GAs from edible parts of S. tuberosum

No studies on potato GA and aglycones were identified.

3.1.2.9.2. GAs from edible parts of food plants other than S. tuberosum

There are several studies focusing on the impact of α-tomatine on the cholesterol metabolism. Cayen et al. (1971) tested whether α-tomatine forms complexes with cholesterol in the intestinal tract (see also **Section 3.1.2.2**). Rats received 600 mg of α-tomatine/kg bw per day via chow, which lowered significantly the uptake of dietary cholesterol by the liver. As compensation, 14 days treatment with 1,200 mg/kg bw per day elevated the rate of acetate incorporation into neutral lipids and de-novo cholesterol biosynthesis being more pronounced in hepatic than in the intestinal cells. Nevertheless, the cholesterol concentrations in the low-density lipoproteins (LDL) and high-density lipoproteins (HDL) fraction were reduced. Furthermore, α-tomatine increased fecal excretion of sterol but not of bile acids, suggesting that that dietary α-tomatine forms a nonabsorbable complex with cholesterol in the intestinal tract and leaves the bile acids unaffected.

Friedman et al. (2000a) also tested whether α-tomatine may lower dietary cholesterol absorption and plasma levels of cholesterol and triglycerides (see also **Section 3.1.2.2**). Hamsters were treated for 21 days with a high-fat/high-cholesterol diet containing from 500 to 2,000 mg/kg α-tomatine. The highest dose (122 mg/kg bw per day) decreased the cholesterol in the serum LDL without changing the cholesterol concentration in HDL. When compared to the controls, the fecal excretion of cholesterol and coprostanol was elevated dose-dependently up to 4–5-fold. This again confirmed the hypothesis of the generation of an insoluble complex between α-tomatine and cholesterol complex in the intestinal lumen.

Friedman et al. (2000b) treated hamsters (8 per group, sex not specified) for 21 days with semipurified diets containing (i) lyophilised ripe *S. lycopersicum*, admixed as powder (with 0.7 mg α-tomatine per kg dw) at a concentration of 610 g/kg chow: estimated uptake of α-tomatine at roughly 0.05 mg/kg bw per day, or (ii) lyophilised unripe (green) *S. lycopsersicum*, admixed as powder (with 743 mg α-tomatine per kg dw) at a concentration of 593.8 g/kg chow: estimated uptake of roughly 46.7 mg/kg bw per day. Compared to the control diet without tomatoes, there was a reduction in body weight and absolute liver weight, most probably due to dietary imbalances. The green and red tomatoes caused a reduction of LDL-cholesterol by 59% and 44%, very low density lipoprotein (VLDL)-cholesterol by 45% and 35% and of plasma triglyceride concentrations by 47% and 31%, respectively. However, the treatments did not impact on plasma levels of HDL but elevated fecal deoxycholic in hamsters fed green fruits, and fecal lithocholic acid concentrations in animals on both green and red tomato diets. However, feeding of both green and red tomato powder resulted in considerably reduced feed intake and weight gain, which impairs the interpretation of data.

To conclude, there is experimental evidence for the formation of undigestible complexes between α-tomatine and cholesterol in the gastrointestinal tract of rodents, which may enhance fecal elimination of sterols and lower the cholesterol concentration in the plasma lipoproteins.

Khaserao and Somani (2017) described an anti-obesity effect of the aubergine aglycone solasodine in obese rats kept on a high-fat diet. Rising concentrations of solasodine (25, 50 and 100 mg/kg bw per day), given for 6 weeks, reduced dose-dependently body weight, abdomen circumference,

retroperitoneal adipose tissue weight, and serum total cholesterol, triglyceride and glucose levels. The induction of a fatty liver was suppressed as well. It is presently unclear whether the blood lipid lowering effect of solasodine is based on a mode of action being similar to the formation of unresorbable complexes of α-tomatine and cholesterol in the gastrointestinal tract.

3.1.3. Observations in humans

3.1.3.1. GAs from *S. tuberosum*

The CONTAM Panel noted that some of the authors of the reports described in this section used a terminology (e.g. 'solanine poisoning') not distinguishing between solanine, chaconine, α-solanine, α-chaconine, solanine:chaconine mixtures and total potato GAs (potato TGAs). This might also be the case for the chemical analysis reported by the authors. The authors' terminology has been kept in this chapter.

Poisonings of humans by GAs, particularly from potato consumption, have been repeatedly reviewed, e.g. by Morris and Lee (1984), van Gelder (1989), JECFA (1993), Tice (1998), Lee (2006), Barceloux (2009), Koleva et al. (2012) and Milner et al. (2011), who report on more than 10 separate outbreaks of potato GA poisonings affecting more than 2,000 subjects including fatalities in more than 1% of the cases. Besides showing membrane disruption activity, potato GAs inhibit the enzyme acetylcholinesterase (AChE), thereby affecting the nervous system directly (see **Section 3.1.5**). At doses > 1 mg/kg bw, potato GAs are considered to be toxic to humans. The studies report that signs of intoxication may occur immediately after intake or with a latency period up to two days. Potato GAs induce symptoms such as sweating, vomiting, diarrhoea, severe abdominal pain, drowsiness, apathy, confusion, weakness, vision disturbances, fever, rapid and weak pulse, low blood pressure and bronchospasm. In severe cases, paralysis, respiratory insufficiency, cardiac failure, coma and death have been reported. Doses in the range of 3–6 mg potato TGAs/kg bw are described to be potentially lethal for humans (JECFA, 1993). However, estimated doses associated with toxicity are regarded as uncertain (McMillan and Thompson, 1979; Morris and Lee, 1984; Hellenäs et al., 1992; JECFA, 1993; Smith et al., 1996; Koleva et al., 2012; Milner et al., 2011).

Toxic GA concentrations are reported to be associated with taste alterations. Bitterness, e.g. when chewing a small piece of the raw peel, is indicative of a high GA concentration in potatoes (Beier and Nigg, 1992; Wood and Young, 1974). Normal levels of GAs in potatoes of less than 100 mg/kg are usually not detectable by taste. GA levels exceeding 100 mg/kg cause a slowly within 15 to 30 sec developing, burning sensation of the tongue and back of the mouth with a persistent irritation. Consumption of potatoes that contain more than 200 mg GAs/kg results in an immediate burning sensation (Wood and Young, 1974). Allergic reactions to raw and cooked potatoes, including cases of anaphylaxis, are only rarely reported and not associated with their content of GAs (e.g. Majamaa et al., 2001; Beausoleil et al., 2001).

In the following sections, only reports that are relevant for the risk assessment of the intake of potato GAs, e.g. giving information on dose-effect relationships, are presented in detail.

3.1.3.1.1 Reports on intoxications

Outbreaks

Several large outbreaks of potato poisonings in humans are known and include those in France (Cortical, 1888, as cited by Rothe, 1919), Germany (Schmiedeberg, 1892, as cited by Bömer and Matthis, 1924; Pfuhl, 1899), East Glasgow (Harris and Cockburn, 1918), Cyprus (Willimott, 1933), North Korea (Reelakh and Keem, 1958), South London (McMillan and Thompson, 1979) and Canada (Canada Diseases Weekly

Report, 1984). Most of the victims of potato poisonings were only moderately affected. However, in some cases serious poisonings including deaths occurred (Damon, 1928; McMillan and Thompson, 1979; Lee, 2006).

In an outbreak in 1898 in Germany, consumption of a large portion of potatoes resulted in 56 soldiers with typical symptoms of 'solanine poisoning', including irritation of the throat, rise of temperature, headache, abdominal pain, diarrhoea, and partly vomiting or nausea, loss of consciousness and jaundice. In one case convulsions were observed. Analysis revealed that the cooked peeled potatoes contained 240 mg/kg and the unpeeled, uncooked potatoes 380 mg 'solanine'/kg (Pfuhl, 1899). The author estimated that for those who consumed the whole meal, the intake of 'solanine' had been 300 mg. In the JECFA (1993) assessment the intake of 'solanine' was estimated to have been 3.4 to 5.1 mg/kg bw.

In 1917, 61 persons from 18 households in Glasgow in Scotland showed the typical picture of 'solanine poisoning' immediately or within 3 h after eating potatoes. Observed symptoms were vomiting, diarrhoea, headaches and debility. A 5-year-old boy died, and the post-mortem examination showing that death was due to bowel strangulation, was interpreted as a consequence of extreme retching and vomiting. The analysis of the potatoes consumed in the boy's family contained 410 mg 'solanine'/kg (Harris and Cockburn, 1918). JECFA (1993) concluded that, assuming a body weight of 18 kg and a consumption of 200 g, the lethal dose for the boy may be estimated at 4.5 mg/kg bw. Furthermore, for adults an intake of 3.4 mg 'solanine'/kg bw was estimated under the assumption that they ate 500 g potatoes and had a bw of 60 kg.

Rothe (1919) reported on an outbreak in Leipzig (Germany) in 1919, which affected 14 persons after consumption of potato dishes with immediate onset of gastrointestinal pain, followed by vomiting and diarrhoea. The author conducted a self-experiment from which he concluded that intake of 150 mg to 200 mg 'solanine' with a meal of cooked peeled potatoes by a male person could lead to gastric discomfort and nausea after 90 minutes, followed by accelerated pulse, symptoms fading within a day (Rothe, 1919).

McMillan and Thompson (1979) reported on an intoxication involving 78 British junior schoolboys (11 to 15 years of age), who became ill with diarrhoea and vomiting after eating two small boiled peeled potatoes each (weight of potatoes not indicated) as part of their midday meal. Symptoms developed mainly between 20.00 and 22.00 o'clock the same evening. Seventeen out of the 78 boys were hospitalised with symptoms of vomiting, severe diarrhoea, along with generalised abdominal pain, commencing together 7 to 19 h after the meal. Their feces were watery and in 13 cases showed markedly green colour by the second or third day as was the case for some of the vomitus. Fifteen of the 17 boys developed fever being maximal just before admission and subsiding within 1 to 3 days. Restlessness, sometimes extreme, was a common symptom often combined with other central nervous system (CNS) effects, such as stupor, delirium, hallucinations, confusion, drowsiness or disorientation. The three most seriously ill boys were comatose or stuporose on admission and had a peripheral circulatory collapse. All of the 17 boys were discharged home within 6–11 days following admission, and when seen 4–5 weeks later were without sequelae. From the 61 boys not admitted to hospital, only for 24 of them Public Health Department records were available. According to these records, all these boys suffered from abdominal pain, diarrhoea and/or vomiting. Twelve boys experienced pyrexia and 13 headaches. Other symptoms reported were delirium, dryness of the throat, trembling, dizziness and weakness, sometimes amounting to a feeling of paralysis. Bacteriological examinations of blood, serum, vomit, urine and feces as well as of samples of food recovered from the meal have all been proved to be negative. Testing of 22 samples of the school meal, 10 samples of the boys' urine and 23 of feces for excess lead, copper, arsenic and zinc revealed negative results as well. Assays for the presence of organic compounds, such as nicotine, organophosphorus or organochlorine pesticides, metaldehyde or

defoliant were also negative. Six days after eating the meal, pseudocholinesterase plasma levels in 10 out of 17 schoolboys were subnormal and returned to the normal range four or five weeks later, for all but one value. The source of poisoning was seen in a bag of potatoes left from the previous term that had been considered 'unfit' for consumption upon inspection, but that had inadvertently been cooked. Peeled boiled potatoes left-over, uneaten after the meal, were extracted and analysed for their anticholinesterase activity. Based on the enzyme activity, the concentration was estimated to amount to 250–300 mg 'solanine'/kg peeled, boiled potato. The authors claimed that this concentration considerably exceeded that of normal fresh peeled tuber (normal value for whole unpeeled tuber: less than 100 mg/kg, with most GAs in the peel). JECFA (1993) estimated the intake of 'solanine' by the schoolboys to have been approximately 1.4–1.6 mg/kg bw.

In 1983, a poisoning incident occurred in Alberta (Canada) when 61 out of 109 school children and staff members suffered from a sudden onset of symptoms after eating baked potatoes containing 494 mg solanine/kg (Canada Diseases Weekly Report, 1984). Predominant symptoms with frequency of occurrence indicated were the following: nausea (69%), abdominal cramps (43%), headache (33%), vomiting (25%), fever (11%) and diarrhoea (8%). The affected persons had recovered in about 3 h. From the affected individuals, 44% had noted a bitter or unusual taste and 18% had felt a burning sensation in the throat when eating the baked potatoes. According to the observations of several staff members the potatoes had a slight tinge of green. Testing of the potatoes for the presence of bacteria, viruses, moulds, pesticides or other chemicals yielded negative results (Canada Diseases Weekly Report, 1984). JECFA (1993) estimated the exposure in this poisoning outbreak at about 2.5 mg 'solanine'/kg bw assuming that the children consumed 200 g potatoes and had a body weight of 40 kg (JECFA, 1993).

Case Reports

While early reports of potato poisoning (Muncke, 1845, as cited by Bömer and Matthis, 1924; Morris, 1859, as cited by Van Gelder, 1989) refer to severe or fatal cases associated with the consumption of green, unripe or 'new' tubers, some studies in the years 1923 and 1924 addressed health impairments related to the intake of potato tubers of normal appearance (Griebel, 1923; Alfa and Heyl, 1923; Bömer and Mattis, 1923, 1924). Bömer and Mattis (1923, 1924) reported that, although measured 'solanine' contents in samples of the consumed potatoes were relatively high and ranged from 257 to 583 mg/kg (raw, unpeeled tubers), observed symptoms were not severe and included burning sensation in the mouth, irritation of mucous membranes, gastrointestinal disturbance and vomiting. Similar symptoms were reported by Griebel (1923) and Alfa and Heyl (1923) after intake of potatoes with 'solanine' levels ranging from 196 to 790 mg/kg (raw, unpeeled tubers) or 109 to 468 mg/kg (raw, peeled tubers), respectively.

In 1924, seven members of a family living in Illinois (USA) were poisoned by consumption of greened potatoes. Two of them died, while the others recovered. Symptoms were epigastric pain, nausea and vomiting. Patients were apathetic and extremely exhausted. Respirations were accelerated. In the two fatal cases, consciousness was lost 3 to 4 h before death (Hansen, 1925).

A food control report from Switzerland reported on two samples of potatoes, which caused burning in the throat and nausea after consumption, 'solanine' levels of 230 and 350 mg/kg (Viollier, 1941). No further information was available.

In the UK, a poisoning incident occurred involving four members of a family on three consecutive Sunday evenings about 9 h after they had consumed baked potatoes eating the flesh and the skin of the tubers. Symptoms consisted of sickness, abdominal pain, diarrhoea and general malaise. Two family members who had only eaten one potato suffered only slightly, a third one, who had two potatoes, suffered more severely and the fourth one who ate three potatoes suffered very severely. A fifth person, who only ate the flesh of the potatoes, did not develop any symptoms. The patients recovered within 24 h. The

potatoes contained 500 mg 'solanine'/kg (Wilson, 1959). JECFA (1993) estimated the intake of 'solanine' at 1.25 to 3.2 mg/kg bw assuming a weight of 150 g per potato.

A 70-years-old woman developed symptoms of poisoning, including vomiting, diarrhoea and bloody feces, after consumption of a juice prepared from partly peeled potatoes without sprouts (Gonmori et al., 1993). She was admitted to hospital and discharged after 27 days (no information on recovery available, only abstract in English). Her blood was analysed by GC and GC-MS on the 4th and 20th days without 'solanine' being detected. The potatoes from which the juice was prepared contained a 15-fold higher 'solanine' concentration compared to normal potatoes (levels not given by the authors) (Gonmori et al., 1993).

In November 2015, members of a family in Baden-Württemberg (Germany) experienced vomiting and stomach-ache after the consumption of potato dishes (jacket potatoes and potatoes boiled in the skin). Due to the bitter taste a salad prepared from the same potato charge was not consumed. Chemical analysis showed that the potatoes, which were without damage, green sites, sprouts or microbiological contamination, contained 236 mg potato TGAs/kg (141 mg α-solanine and 95 mg α-chaconine per kg). In the potato salad a lower concentration of 124 mg potato TGA/kg was found. No information was available on the quantity of potatoes consumed, if all members of the family were affected by the intoxication and on recovery (BfR, 2018a,b).

3.1.3.1.2. Studies in human volunteers

In 1919, Rothe conducted a self-experiment from which he concluded that intake of 150 mg to 200 mg 'solanine' with a meal of cooked peeled potatoes by a male person could lead to gastric discomfort and nausea after 90 minutes, followed by accelerated pulse, symptoms fading within a day (Rothe, 1919).

In three Swedish adult volunteers (two males and one female), who consumed daily for 1 week 200–300 g of cooked unpeeled potatoes of varieties high in GAs (180–300 mg/kg), no signs of acute toxicity were noted. However, the three subjects reported on a bitter taste (Harvey et al., 1985). JECFA (1993) estimated the intake of TGA at 1 mg/kg bw, assuming a weight of 150 g per potato.

In a kinetic study, 7 healthy male adults have been given a potato meal, which consisted of mashed potatoes prepared from two parts peeled potatoes and one part low-fat milk (Hellenäs et al., 1992, see **Section 3.1.1**). The dish contained 200 mg GAs (82 mg α-solanine and 118 mg α-chaconine) per kg. The amount consumed was adjusted in relation to each subject's body weight to standardise the TGA intake for 1 mg/kg bw. All except one of the volunteers experienced signs of toxicity (mainly light to severe nausea, in one case combined with diarrhoea) and an unpleasant off-taste after ingestion of the meal. In most subjects, symptoms started within about 30 min after exposure and lasted for 3 to 4 h. The peak serum concentrations for α-solanine and α-chaconine were on average reached after 5.1 and 6.0 h, respectively and amounted on average to 7.7 and 14.4 ng/mL (8.8 and 16.9 nmol/L), respectively. During the 25-h period the highest solanidine values varied among the subjects between 1.0 and 4.8 ng/ml (2.5 and 12 nmol/L). No correlation between the severity or duration of adverse effects and GA serum levels was seen and the highest maximum blood serum concentrations for α-solanine and α-chaconine of 11.3 and 21.4 ng/mL, respectively, were exhibited in the one subject not experiencing signs of toxicity (solanidine serum level not given). According to the authors, the rapid onset and short duration of the symptoms compared to the GA serum profiles suggest that the symptoms might be the consequence of local direct effects on the gastrointestinal tract rather than of systemic interactions. The authors noted that the study was not focusing on the toxic effects of the GAs and was not conducted double-blinded. Therefore, a placebo effect could not be excluded.

Bushway (1987, personal communication, as cited by Van Gelder, 1989) reported on a study in six male adult volunteers who consumed potato tubers which contained 320 mg potato TGA/kg. The potatoes

were baked and consumed unpeeled. The amounts of GAs ingested ranged from 1.75–2.58 mg/kg bw. Symptoms of intoxications occurred in all of the subjects, started about 2 to 4 h after ingestion and included nausea, vomiting and diarrhoea. Symptoms faded away after 8–10 h.

In a study in 14 volunteers (6 males, 8 females), aged 18–45 years, the toxicity and toxicokinetics of different oral single doses of the potato GAs, α-chaconine and α-solanine, were investigated (Mensinga et al., 2005, see **Section 3.1.1**). The subjects (2–3 per treatment) received solutions with potato TGA doses of 0.30, 0.50 or 0.70 mg/kg bw, or mashed potatoes adjusting TGA doses to 0.95, 1.10 or 1.25 mg/kg bw under standardised conditions. The solutions contained 50% α-solanine and 50% α-chaconine. The mashed potatoes had a TGA concentration of 199 mg/kg (51% α-solanine, 49% α-chaconine), which was recognised as upper limit of safety according to the authors. No adverse events did occur in any of the treatments, except for one of the two volunteers in the high-dose group receiving 1.25 mg potato TGA/kg bw in mashed potatoes. About 4 h after administration, this subject developed nausea and started vomiting, which was associated by the authors with GA toxicity, even though the doses was lower than those generally reported in the literature to cause gastro-intestinal disturbances. According to the authors, the serum GA concentrations of this subject were comparable to those of the subject receiving a similar dose and did not show any symptoms. For both subjects, maximum blood serum concentrations were ranging for α-solanine between 8.0 and 11.8 ng/mL and for α-chaconine between 6.3 and 9.7 ng/mL, no individual levels being indicated for each subject. The authors concluded from the results that some subjects may be more susceptible for adverse effects of potato GAs than others.

3.1.3.1.3. Epidemiological studies

Congenital anomalies

Intake of potato GAs, particularly with blighted or injured potatoes have been proposed as an aetiological factor in the pathogenesis of neural tube defects (NTD) (anencephaly and spina bifida) (Renwick, 1972; Harvey et al., 1986). This hypothesis has been addressed in two epidemiological studies (Nevin and Merrett, 1975; Harvey et al., 1986). Maternal consumption of sprouted potatoes as risk factor of NTD has also been investigated in two Chinese studies (Wang et al., 2008; Ni et al., 2018).

In the prospective study of Nevin and Merrett (1975) in women with a previous infant with either anencephaly and/or spina bifida and advised to avoid potato consumption, the incidence of NTD was 8.7% in the group of women adhering to the advice (n = 27), and 3.6% in the group of non-compliers (n = 61) (p = 0.58). In the prospective study by Harvey et al. (1986) conducted in NTD screening centres, outcome of pregnancy in 380 patients resulted in 210 NTD cases and 170 normal offspring, whereby in most of the 9 centres studied, serum total potato alkaloids levels and serum 'solanidine' levels were higher in the women with a normal fetus than in those with a fetus affected by NTD. For both groups serum total potato alkaloid concentrations varied widely between individuals as well as between each of the participating centres, whereby levels for individual subjects ranged from < 10 nmol/L to < 150 nmol/L.

Two Chinese case-control studies investigated the risk factors of NTD in the Shanxi province in Northern China (Wang et al., 2008; Ni et al., 2018). The results indicate that, among other factors, maternal consumption of sprouted potatoes during the periconceptional period may increase the risk of NTDs or orofacial clefts (OFCs). Subjects recruited between 2002 and 2007 by Ni et al. (2018) included 622 cases with NTD, 135 cases with orofacial clefts (OFCs) and 858 controls. Data collection comprised information on maternal sprouted potato consumption, lifestyle behaviours and folic acid supplementation. Intake of sprouted potatoes was not associated with spina bifida or encephalocele, but with increased odds of total NTDs (OR = 2.20; 95% CI, 1.12–4.32) and anencephaly (OR = 2.48;

95% CI, 1.10–5.58) and with elevated risk of total OFCs (OR = 3.49; 95% CI, 1.29–9.49) and cleft lip with or without cleft palate (CL ± P) (OR = 4.03; 95% CI, 1.44–11.28). The authors also found that maternal consumption of sprouted potatoes was significantly associated with NTDs and OFCs among mothers with lower consumption of B vitamins, but not among mothers with a higher consumption of B vitamins. Another case-control study was conducted by Wang et al. (2008) in subjects genotyped for methylenetetrahydrofolate reductase (MTHFR) C667T polymorphism. Multivariate conditional logistic regression analysis indicated that four maternal factors were linked to NTDs: frequency of pregnancy (OR = 2.87, 95%CI: 1.28 - 6.44), contacting chemicals in early pregnancy (OR = 16.18, 95%CI: 1.18 - 221.59), frequent taking of germinated potatoes in early pregnancy (OR = 4.66, 95%CI: 1.78 - 12.17) and MTHFR C677T mutation (OR = 2.13, 95%CI: 1.08 - 4.21). The CONTAM Panel noted that no information on GA levels in the sprouted potatoes or GA intake was given in the two studies.

The CONTAM Panel noted that the hypothesis of Renwick (1972) has not been supported by the outcome of the epidemiological studies of Nevin and Merrett (1975) and Harvey et al. (1986). The design and results of the two Chinese studies (Wang et al., 2008; Ni et al., 2018) do not allow conclusions on causal relationships of GA exposure and developmental effects.

Cancer

The relationship between the intake of potatoes and risks of cancer in the gastrointestinal tract has been investigated. As reviewed by Hopkins (1995) and Tice (1998) results of epidemiological studies were contradictory and remained altogether inconclusive. A causal relationship between diets with the consumption of large quantities of potatoes and increased risks of cancers of the brain, breast, endometrium, lung and thyroid could not be proven (Hopkins, 1995; Tice, 1998).

The CONTAM Panel noted that the design and outcome of existing studies do not allow concluding on the causal relationship between intake of potato GAs and cancer risks.

3.1.3.1.4. Conclusions

Overall, the CONTAM Panel noted that the information from the available human data is limited. However, results from kinetic studies in volunteers (Harvey et al., 1985; Hellenäs et al., 1992; Mensinga et al., 2005) and from reports on intoxications (e.g. Wilson, 1959; Bushway, 1987, personal communication, as cited by Van Gelder, 1989) indicate that acute toxic effects, such as nausea, vomiting and diarrhoea, may occur in adults from a potato TGA intake of 1 mg/kg bw or more (**Table 20**). Doses in the range of 3–6 mg potato TGAs/kg bw are considered to be potentially lethal for humans.

Results of studies conducted under standardised conditions (Hellenäs et al., 1992; Mensinga et al., 2005) suggest that differences in the individual susceptibility towards adverse effects associated with the intake of potato GAs may be expected. The available human data do not allow to draw conclusions if children or other subgroups of the population, such as individuals with inflammatory bowel diseases, may be more susceptible to adverse effects associated with potato GAs than others.

No data showing convincing evidence of GA-related health problems in humans associated with the long-term intake of potatoes have been identified.

2305 **Table 20.** Summary of reports on outbreaks and cases of intoxications in humans and on volunteer studies relevant for the identification of the reference point
2306 for acute toxicity.

Affected subjects	Potato type /preparation	Concentration of solanine or TGAs in the potatoes or potato preparation, respectively [a]	Intake per person	Estimated single dose of solanine or potato TGA/kg bw [b]	Taste, symptoms and outcome	Reference
3 adult volunteers (two males and one female),	potato varieties high in GAs	180–300 mg 'TGA' mg/kg potatoes	for 1 week daily 200–300 g of cooked unpeeled potatoes	1 mg 'TGA'/kg bw [c]	bitter taste, no signs of acute toxicity	Harvey et al. (1985)
6 out of 7 male adult volunteers	preparation of mashed peeled potatoes with milk	200 mg TGAs (82 mg α-solanine and 118 mg α-chaconine)/kg potato preparation	adjusted to bw resulting in a dose of 1 mg TGAs/kg bw [d]	1 mg TGAs/kg bw [d]	unpleasant off-taste; nausea, which in one case was combined with diarrhoea; all recovered within 4 h	Hellenäs et al. (1992)
6 male adult volunteers	baked potatoes, consumed unpeeled	320 mg TGAs/kg	n.r.	1.75–2.58 mg TGAs/kg bw [d]	nausea, vomiting, diarrhoea, all recovered within 8–10 h.	Bushway (1987, personal communication, as cited in Van Gelder, 1989)
1 out of 2 adult volunteers	mashed potatoes	199 mg TGA/kg fw (51% α-solanine, 49% α-chaconine)	adjusted to bw resulting in a dose of 1.25 mg TGAs/kg bw [d]	1.25 mg TGAs/kg bw [d]	nausea, vomiting	Mensinga et al. (2005)
4 adults	baked potatoes (flesh and skin eaten)	500 mg 'solanine'/kg potatoes	1–3 potatoes	1.25–3.25 mg 'solanine'/kg bw [c]	sickness, abdominal pain, diarrhoea, general malaise; all recovered within 24 h	Wilson (1959)

78 school-boys (11–15 years old)	old potatoes, 'unfit' for consumption	250–300 mg 'solanine'/kg peeled boiled potatoes	two small potatoes	1.4–1.6 mg 'solanine'/kg bw (c)	diarrhoea, vomiting, abdominal pain, 17 boys hospitalised due to severe symptoms including 15 subjects with fever and 3 being comatose or stuporose; all recovered	McMillan and Thompson (1979)
61 out of 109 school children and staff members	baked potatoes with a 'slight green tinge'	494 mg 'solanine'/kg potatoes	n.r.	2.5 mg 'solanine'/kg bw (c)	bitter or unusual taste (44%), burning sensation in the throat (18%); nausea (69%), abdominal cramps (43%), headache (33%), vomiting (25%), fever (11%), diarrhoea (8%); all recovered in about 3 h	Canada Diseases Weekly Report (1984)

(a): Concentrations are given as 'potato TGAs', 'solanine', 'α-solanine' or 'α-chaconine' depending on the subject of analysis as described in the referenced publication.
(b): Doses are given as 'TGAs', or 'solanine', depending on the subject of analysis as described in the referenced publication.
(c): As estimated in JECFA (1993).
(d): As estimated by the authors.

3.1.3.2. GAs from food plants other than *S. tuberosum*

The occurrence of GAs in foods other than potatoes has been reviewed by Tice (1998) and Dolan et al. (2010) (see **Section 1.3.1** and **1.3.3**). For example, tomato fruits, particularly the green unripe fruits, are known to contain, i.a. α-tomatine. However, according to Dolan et al. (2010) there is no evidence to consider α-tomatine as a substance of concern and there are no reports of acute toxicity in humans due to ingestion of green tomatoes.

3.1.3.2.1. Case Reports

In Okinawa (Japan) intoxication of a couple occurred three hours after consumption of a dish with a sauce containing aubergine. Observed symptoms were staggering, slurred speech, mydriasis and drowsiness. The vegetable originated from an aubergine grafted on Devil's Trumpet (*Datura metal*). Scopolamine and atropine were detected in the aubergine sauce and sera of the subjects (Oshiro et al., 2008, see **Section 1.3.3**).

Smith et al. (2008) reported on intoxications occurring on two different occasions with susumber berries originating from the plant *S. torvum*. Poisonings with susumber berries personally imported from Jamaica occurred in three persons in 2003 in Toronto and in three persons in 2006 in York City. Symptoms included diarrhoea, weakness, facial paralysis, slurred speech and ataxia. The most seriously affected patients developed hypertension, confusion, and respiratory failure requiring mechanical ventilation. The authors noted that the imported poisonous berries ingested appeared indistinguishable from non-toxic varieties commercially available on the market. α-Solasonine and α-solamargine, which share the same aglycone, solasodine, were isolated from the toxic berries but not from their non-toxic counterparts (Smith et al., 2008).

The French Agency for Food, Environmental and Occupational Health and Safety (Anses) reported in 2014 on 26 cases of poisoning in France associated with consumption of cherry tomatoes originating from Morocco (Anses, 2014). Gastrointestinal symptoms, including vomiting, abdominal cramps and irritation of the pharynx, were accompanied by an unpleasant bitter taste. The symptoms were observed in most cases 5 to 30 minutes after ingestion of the cherry tomatoes. Based on chemical analysis using LC-HRMS, rubijervine was postulated as a metabolite present in the cherry tomatoes. However, it was not possible to confirm the identity or to perform a quantification. Neither α-tomatine nor α-solanine could be detected in the tomatoes. Rubijervine is the 12α-hydroxy metabolite of solanidine which is known to occur also in other plants such as those of the genus *Veratrum*. Metabolome profiling studies conducted on wild and cultivated tomato species using similar techniques have not reported the presence of rubijervine in any of the investigated samples (Moco et al., 2007; Iijima et al., 2013; Schwahn et al., 2014), a fact also mentioned by the authors of the Anses report. The authors concluded that the intoxications could have been caused by GAs, but that further investigations would be necessary to identify the causative substances.

3.1.4. Adverse effects in farm animals, horses and companion animals

The selection criteria for studies relevant to inform the adverse effects in farm and companion animals are described in **Section 2.1**.

3.1.4.1. Ruminants

Three studies on ruminants could be identified (see **Table 21**), which lack significance due to the low number of animals investigated and due to lack of information on the exact composition and the degree

of purity of the test compounds. These limited studies indicate that acute toxicity may be expected to occur at oral doses above 40 mg/kg bw in sheep and cows.

3.1.4.2. Pigs

Kerr et al. (1998) treated weanling pigs with control diet containing 3% spray-dried animal plasma or diets with additional 2.6% or 5.1% potato proteins for 28 days (see **Table 22**). GA content of potato proteins was determined by HPLC-UV. No information is provided on the concentration of α-solanine and/or α-chaconine. Increasing concentrations of potato proteins had no effect on the average daily body weight gain or food intake of the animals.

3.1.4.3. Poultry

The studies identified are reported in **Table 23**.

Vogt and Stute (1969) used an extract of *S. tuberosum* as a source of α-solanine. However, information is lacking whether the extract contained also α-chaconine or further GA or aglycones. The extract was admixed to a diet containing dried ground potatoes with a α-solanine content of 100–150 mg/kg. This treatment increased the body weight gain in young male Acres chicken when treated daily with 4 to 12 mg α-solanine per kg bw for 46 days.

Tuśnio et al. (2013b) used two different potato protein concentrates containing 677–3,185 mg/kg TGA. No information is given on the concentrations of α-solanine and α-chaconine, and the ratio between these two GA is likely to be different in the potato concentrates. A GA dose of 101.5 mg/kg bw per day reduced feed intake and body weight gain in female chicken when treated over a period of 35 days. Furthermore, an increased relative weight of liver and pancreas could be observed in this study at a dose of 67.7 mg/kg bw per day. When applying 154.2 mg/kg bw per day, the authors observed morphological changes in the intestinal tract of female chicken. The interpretation of these observations is hampered by the fact that the diet contained also trypsin inhibitors at various concentrations, which might have affected the digestibility of the feed components and the activity of the gastrointestinal tract.

3.1.4.4. Rabbits

No studies on the adverse effects in rabbits have been identified other than the ones described already in **Section 3.1.2.1** and **3.1.2.2**. In the repeated-dose toxicity studies no NOAEL or LOAEL could be derived, since untreated controls were missing.

3.1.4.5. Fish

The studies identified in fish are reported in **Table 24**. One 84-day study in rainbow trout indicated that the most sensitive endpoints are decreased body weight, lowered serum proteins and an altered hepatic histology occurring at the lowest doses of α-solanine/α-chaconine of 0.008/0.012 mg/kg bw per day (Tusche et al., 2011). No NOAEL or LOAEL could be identified.

3.1.4.6. Horses

No studies were identified.

3.1.4.7. Companion animals (cats and dogs)

No studies were identified.

3.1.4.8. Fur animals

No studies were identified.

3.1.4.9. Reports on intoxications

The risks of feeding green potatoes or sprouts to livestock have been well understood for many years, and therefore there have been relatively few recent reports of intoxications following ingestion of these feeds. Outbreaks of poisoning of cattle in the UK fed green or sprouting potatoes have been reported (Milligan, 1941), while in Poland, outbreaks of 'solanine' poisoning in pigs were reported between 1949 and 1961. These involved 180 pigs, and resulted in 64% mortality, but levels of GAs were not given (Cooper and Johnson, 1984). Sheep are also susceptible to poisoning following consumption of green potatoes. In the USA, 14 (of 35) became weak, uncoordinated and subsequently died after consuming green potatoes which had been spread on the pasture (Bolin, 1962)

Poisoning of livestock following consumption of other GA-containing plants, such as *S. dulcamara* and *S. nigrum*, are uncommon. Because of their strong odour and bitter taste, animals do not eat the growing plants as long as other feed is available. However, these plants may grow as weeds in hay and silage crops, where they can be harvested with the crop and then fed to livestock. Drying or ensiling does not destroy the toxin (USDA, 2011), and as a result cases of accidental poisoning (or suspected poisoning) have been reported in horses, cattle, sheep, goats and pigs where livestock have consumed hay or silage, which have been contaminated with these plants (Coper and Johnson, 1984).

Norman et al. (2012) reported a case in which six horses were fed poor quality hay containing *S. eleagnifolium* (Silverleaf nightshade) and *S. dimidiatum* (Western horse nettle). *S. eleagnifolium* is known to contain solasodine-type GAs (Álvarez et al., 1994). Symptoms included gastrointestinal upset, respiratory depression, and central nervous system depression, obtundation, ataxia, pyrexia, muscle fasciculations, and ileus. Therapy for these horses was poorly described, but all the affected horses recovered. Gunning (1950) also reported a case of poisoning and death in a horse fed greened potato sprouts.

Poultry appear to be less susceptible to 'solanine' poisoning; in feeding trials laying hens tolerated green potato sprouts and ground tubers when included at 10% of their diet for long periods (Temperton, 1943).

Kees et al. (2015) reported a case of GA toxicity in a 10-week-old male Labrador retriever dog as a result of ingesting dried stems and unripe berries of *S. dulcamara. S. dulcamara* may contain a variety of GAs such as α-solasonine, α-solamargine, α-solamarine, β-solamarine and soladulcine A (Calf et al., 2018). The level of exposure to GAs was not determined.

2425 **Table 21.** Acute toxicity of glycoalkaloids (GAs) (α-solanine) and aglycones (solasodine) in ruminants.

Test compound	Species Dose route Doses	Observed effects	Highest dose with no effect (mg/kg bw)	Lowest dose with effects (mg/kg bw)	LD_{50} (mg/kg bw)	Reference
α-Solanine [(b)]	**Sheep** (F) No/sex/group: 1 Single oral dose: 100 or 225 mg/kg bw NO CONTROL	Anisocytosis, poikilocytosis, leukocytosis		225		König and Staffe (1953)
α-Solanine [(a)]	**Adolescent bull** (M) No/sex/group: 1 Single gavage: 77 mg/kg bw NO CONTROL		77			Liebenow (1970)
Solasodine [(a)]	**Adolescent bull** (M) No/sex/group: 1 Singe gavage: 43 mg/kg bw NO CONTROL		43			Liebenow (1970)

2426 F: female. M: male.
2427 (a): Chemical standard obtained by in-house isolation from plant material, and checked for purity (95% pure or higher).
2428 (b): Poorly characterised chemical standard (purity or composition not defined, may contain α-chaconine).

2429

2430 **Table 22.** Repeated toxicity of glycoalkaloids (GA) containing potato protein in pigs.

Test compound	Species Dose route Doses	Observed effects	Highest dose with no effect (mg/kg bw)	Lowest dose with effects (mg/kg bw)	LD_{50} (mg/kg bw)	Reference
Potato protein [(a)]	**Pigs** (M,F) No/sex/group: ~18 Diet: 0, 4.6 or 8.5 mg TGA/kg bw/day Duration: 28 days		8.5			Kerr et al. (1953)

2431 F: female. M: male.
2432 (a): A non-purified material for which the TGA concentration is given.

2433

2434 **Table 23.** Repeated toxicity of glycoalkaloids (GAs) (α-solanine) and potato protein concentrates in poultry.

Test compound	Species Dose route Doses	Observed effects	Highest dose with no effect (mg/kg bw/day)	Lowest dose with effect (mg/kg bw/day)	Reference
α-Solanine [(a)]	**Acres chicken** (M) No/sex/group: 3 Diet: 0, 12.5, 25 or 50 mg/kg (equivalent to 0, 1.1–3, 2.2–6 or 4.4–12 mg/kg bw per day) Duration: 46 days	↑ bw gain	2.2–6	4.4–12	Vogt and Stute (1969)
Potato protein concentrate containing GAs and also trypsin inhibitors at various concentrations [(b)]	**Chicken** (F) No/sex/group: 14 Diet: 0, 86.2, 87.7, 154.2, 274.5 or 318.5 mg/kg bw per day Duration: 23 days	↓ decreased height of villi in jejunum + ileum ↓ depth of crypts in jejunum	87.7 87.7	154.2 154.2	Tuśnio et al. (2013b)
Potato protein concentrate containing GAs and also trypsin inhibitors at various concentrations [(b)]	**Chicken** (F) No/sex/group: 24 Diet: 0, 33.8, 67.7 or 101.5 mg/kg bw per day Duration: 35 days	↓ feed intake ↓ bw gain ↑ relative weight of pancreas and liver ↑ relative weight of gastrointestinal tract	67.7 67.7 33.8 67.7	101.5 101.5 67.7 101.5	Tuśnio et al. (2013b)

2435 F: female. M: male. Bw: body weight,
2436 (a): Poorly characterised chemical standard (e. g. purity or composition not defined).
2437 (b): A non- or only partly purified extract or material for which the GA composition and concentration is given.

2438

2439 **Table 24.** Repeated toxicity of potato protein concentrates in fish.

Test compound	Species Dose route Doses	Observed effects	Highest dose with no effect (mg/kg bw/day)	Lowest dose with effect (mg/kg bw/day)	Reference
Potato protein concentrate containing GA at 7.41 mg/kg dw [(a)]	**Rainbow trout** Application oral Diet containing α-solanine/α-chaconine at 0/0, 0.4/0.6, 0.8/1.2, 1.2/1.7 or 1.6/2.3 mg/kg dw (equivalent to: 0/0, 0.008/0.012, 0.016/0.024, 0.024/0.034 or 0.032/0.048 mg/kg bw per day [(b)]) Duration: 84 days	↓ bw ↓ Serum glucose ↓ Serum protein Altered liver histology	 0.016/0.024	0.008/0.012 0.024/0.034 0.008/0.012 0.008/0.012	Tusche et al. (2011)
Potato protein concentrate containing GA at 2,150 mg/kg dw [(a)]	**Rainbow trout** Application oral Diet containing α-solanine/α-chaconine at 0/0, 166/149, 306/298, 459/446 or 612/595 mg/kg dw (equivalent to: 0/0, 3.32/2.98, 6.12/5.96, 9.18/8.92, 12.24/11.9 mg/kg bw per day [(b)]) Duration: 84 days	↓ bw ↓ Relative liver weight ↓ Serum glucose and protein ↓ Serum triglyceride Altered liver, intestinal and stomach histology	 	3.32/2.98 3.32/2.98 3.32/2.98 3.32/2.98 3.32/2.98	Tusche et al. (2011)

2440 dw: dry weight. bw: body weight.
2441 (a): A non- or only partly purified extract or material for which the GA composition and concentration is given.
2442 (b): Body weight of 2 kg and average feed consumption of 40 g per day were assumed (EFSA, 2012).

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3.1.5. Mode of action

Information from the available human data on poisoning with potato GAs, indicate that acute toxic effects of GAs comprise nausea, vomiting, abdominal cramps and diarrhoea (see **Section 3.1.3**). These symptoms may be caused by local irritating effects in the mucosa of the gastrointestinal tract referring to the ability of potato and tomato GAs to form complexes with membrane 3β-hydroxy sterols, thereby disrupting the integrity of cell membranes. The acute toxic effects in humans might be related also to the cholinergic toxidrome or toxic syndrome, comprising a set of symptoms caused by a group of toxic chemicals leading to over-stimulation of cholinergic receptors [21]. The various effects of GAs are outlined in detail below.

3.1.5.1. Membrane effects with implications for the gastrointestinal tract

GAs, like α-solanine, α-chaconine and α-tomatine, may exert saponine-like effects which may injure membranes. This membrane toxicity has been documented in several tissues or cell lines, such as rat liver mitochondria (Keukens et al., 1996), mammalian erythrocytes (Keukens et al., 1996), rodent cell lines (Gee et al., 1996) as well as in human colorectal carcinoma (Glynn et al., 2017; Arena et al., 2018), embryonic intestinal (Gee et al., 1996; Keukens et al., 1996; Patel et al., 2002) and MCF7 breast cancer cell lines (Sucha et al., 2013).

The molecular mechanism of toxicity may be based on membrane destabilisation (Friedman, 2006). The presence of sterols in the membrane appears to be an absolute requirement for the membrane disruptive action of the GAs (Keukens et al., 1992). It has been suggested that the initial insertion of the aglycone part into the membranes is followed by the generation of a 1:1 complex with membrane sterols, which disrupts the structure of the lipid bilayers. GAs interact with the sterols in the following potency order: α-tomatine > α-chaconine > α-solanine (Friedman, 2006). Deletion of one or more mono-saccharides from the GAs resulted in an almost complete loss of activity. Accordingly, solanidine was weakly active (Keukens et al., 1992, 1995). Keukens et al. (1995) proposed that on the one side the length of the sugar chain of the GA and on the other side a planar ring structure, a 3β-OH-group as well as a side-chain at position 24 of the sterol, are critical. As a consequence of this interaction, the permeability, stability, flexibility and fluidity of cell membranes may be considerably altered (Friedman, 2006).

Manrique-Moreno et al. (2014) showed that the aubergine-derived aglycone solasodine may cause structural perturbations in the membranes of human erythrocytes. In more detail, X-ray diffraction studies demonstrated that solasodine interacted mainly with dimyristoylphosphatidylcholine, one of the major phospholipids in the outer and inner erythryocyte membrane affecting the polar head and acyl chain regions of the molecule. Interestingly, the strongest effect was seen with the aglycone solasodine and the lowest with the GA α-solanine. It has not been investigated whether the potato aglycone soladinine exerts similar effects in phospholipids as solasodine.

The disintegrating effect of GA on membranes may be of significance in the gastrointestinal tract after enteral uptake of the compounds. For example, Gee et al. (1996) reported that GAs reduced the integrity of isolated rat mucosa, which impaired active nutrient transport and enabled high molecular weight compounds (such as allergens or GAs) to cross the epithelial barrier. The communication between intestinal cells by gap-junctions may also be a target of GAs, as observed in Caco-2 human epithelial colon carcinoma cells after treatment with α-chaconine and α-solanine (Keukens et al., 1996; Glynn et al., 2017). GAs also disturb the transport of Ca^{2+} and Na^{+} across cell membranes. For example, Blankemeyer et al. (1995, 1997) found that the transepithelial active transport of sodium decreased

[21] Report on the Toxic Chemical Syndrome. Definitions and Nomenclature Workshop. May 8-9 2012. Available at: https://chemm.nlm.nih.gov/Report_from_Toxic_Syndrome_Workshop_final_with_ACMT_edits_cover.pdf

considerably when incubating frog skin with α-tomatine, α-chaconine or α-solanine. In the rat duodenum, 5 mM α-solanine applied via drinking water, inhibited the active calcium transport in a non-competitive way (Michalska, 1985). The extent of these effects may depend on the pH within the intestinal lumen, e.g. GAs show a low solubility at neutral pH and a higher solubility around a pH of 6, as occurring in the proximal jejunum (Gee et al., 1996). However, it has to be considered that the intestinal mucosa in the intact organism of healthy individuals is protected by a layer of mucus, which may antagonize the effects of GAs.

Disruption of epithelial barrier integrity by GAs may be important in triggering inflammatory bowel disease in susceptible individuals. To test this hypothesis, Patel et al. (2002) treated homozygous IL-10 gene-deficient mice with overt enterocolitis with a 1:1 mixture of α-chaconine:α-solanine at 3 mg/kg bw per day via drinking water for 21 days. This treatment aggravated intestinal injury, as determined by pathohistological diagnosis, while in control animals no effect could be seen. The same group used Il-10-gene deficient male mice as well as male mice with dextran sodium sulphate-induced colitis exposed to fried skin from potatoes with low/medium (154 mg GA/kg skin, i.e. 30.8 mg/kg bw per day) or high GA content (460 mg GA/kg skin, i.e. 92 mg/kg bw per day) for 20 days (no further feed was provided to the mice) (Iablokov et al., 2010). Exposure to the low/medium GA skins elevated interferon-gamma (IFN-γ) in the Il-10-gene-deficient mice, and IFN-γ, TNF-α, and IL-17 in mice with the dextran sodium sulphate-induced colitis. In the latter model also enhanced colonic permeability could be observed.

To conclude, the cytotoxicity in the gastrointestinal mucosa may contribute to symptoms observed after GA intoxication, such as necrosis in experimental animals or diarrhoea and vomiting in humans (see **Sections 3.1.2** and **3.1.3**). It remains to be elucidated in more detail whether susceptible individuals are at an elevated risk for the development of inflammatory bowel disease after ingestion of GAs.

3.1.5.2. Inhibition of cholinesterases (ChEs)

3.1.5.1.1. Comparative determination of inhibition of ChEs in vitro

ChEs have to be subdivided into two classes of enzymes: acetylcholinesterase (AChE, EC 3.1.1.7 [22]) and butyrylcholinesterase (BuChE, EC 3.1.1.8 [22]). This distinction is important because in older studies the term ChE has been used in a general sense, not discriminating between the two classes of enzymes. The BuChE class of enzymes was also called 'pseudocholinesterase' or in older studies, 'unspecific ChE', representing the serum ChE.

The primary function of ChEs is to inactivate acetylcholine (ACh) by cleavage, thereby avoiding overstimulation of cholinergic processes. These cholinolytic functions of ChEs require the catalytic (enzymatic) function of the protein. This function is required in the case of the classical synaptic function (Vogel-Höpker et al., 2012). Within the enzymatic mode of action AChE most likely functions in cooperation with other components of the cholinergic system including the substrate ACh, both the nicotinic and muscarinic acetylcholine receptors, choline acetyltransferase, and potential ACh transporters.

Besides cleavage of ACh, BuChE is primarily known for its activity in metabolism of bioactive esters from food and medications, particularly for its role to inactivate the muscle relaxant succinylcholine, used to facilitate abdominal surgery (Brimijoin et al., 2016). Otherwise it was not known to play a direct role in mammalian physiology and was regarded as an orphan enzyme. This changed when it was recognized that BuChE cleaves ghrelin, a hormone involved in appetite control.

[22] International Union of Biochemistry and Molecular Biology (IUBMB) enzyme nomenclature.

Each class of enzymes can be inhibited by two typical modes of action, the competitive reversible and the non-competitive irreversible mode of inhibition. Reversible competitive inhibition of ChEs is used for clinical purposes using substances with a positively charged nitrogen, competing for the ligand binding domain of the enzyme. Chemically, these inhibitors include compounds with different functional groups (carbamate, quaternary or tertiary ammonium group). They are/were applied for diagnostic and/or treatment purposes for diseases such as myasthenia gravis, Alzheimer disease or glaucoma.

Non-competitive inhibitors represent largely organophosphates which are esters or thiols derived from phosphoric, phosphonic, phosphinic or phosphoramidic acid. They exert their main toxicological effects through non-reversible phosphorylation of esterases in the central nervous system thereby irreversibly inactivate the catalytic centre of the enzyme (Čolović et al., 2013). GAs are inhibitors of ChEs and follow the reversible, competitive mode of action.

Regarding inhibition of ChEs by GAs, Pokrovskii (1956) was the first to describe inhibition of enzymatic activity by α-solanine. The first description of inhibition of serum ChE in the English literature was by Orgell et al. (1958), who showed that a peel extract is up to 40-fold more potent than in the inner part of the tuber. In the meantime, there is a broad range of evidence supporting the observation that GAs, particularly α-chaconine and α-solanine, are potent inhibitors of ChEs, and that the GAs are more potent than the aglycones.

Inhibition of ChEs by GAs was reported in two different ways, representing two different experimental scientific approaches. Some studies described percentage of inhibition obtained at tested concentration(s) of the GAs or their aglycones against control, while others determined inhibitory constants (K_i). Consequently, results are summarised in two different tables representing percentage of inhibition values (**Appendix E**, Table E.1) and determined K_i values (**Appendix E**, Table E.2). A direct comparison of the potency of inhibition of the two ChE enzyme classes is difficult, because enzymes were measured in different organs or body fluids and concentrations used to study percentage of inhibition differed considerably. However, from direct comparative studies some general findings can be delineated. The most potent GAs in term of inhibition of ChEs were α-solanine and α-chaconine, with α-chaconine in tendency being slightly more potent. BuChE appears to be a slightly more sensitive target for inhibition by GAs than AChE. Human ChEs reacted equally if not more sensitive to inhibition by GAs than their counterparts from animals (see **Appendix E**).

Harris and Whittaker (1959) extracted 85 g of potato peel with 300 mL water and tested these extracts on human serum ChE. By electrophoretic procedures they could discriminate between three subtypes: the unusual phenotype, the intermediate phenotype and an atypical phenotype. The percentage of inhibition recorded was: 77–83% for the unusual phenotype, 53–67% for the intermediate phenotype and 16–24% for the atypical phenotype. Using the same discrimination of enzyme subtypes for human serum ChEs, the same authors made a comparative investigation of the inhibitory activity of α-solanine at 2.88 µM and its aglycone solanidine at 3.14 µM (Harris and Whittaker, 1962). For α-solanine, the authors found 86.2% inhibition for the unusual phenotype, 65.8% inhibition for the intermediate phenotype and 21% for the atypical phenotype. Inhibition by 3.14 µM solanidine was slightly lower, with 80% inhibition for the unusual phenotype, 53.2% for the intermediate phenotype and 0.8% for the atypical phenotype. Overall, the inhibitory activity of the aglycone seems to be lower than that of the glycoside. Nigg et al. (1996) did a comparative assessment of α-chaconine and α-solanine. They also used human serum ChE, specifically targeted BuChE using α-naphthylacetate as substrate at a concentration of 2.88 µM and had three timepoints of measurement (0, 10, 30 min). Whereas the inhibitory activity for α-solanine was 50% at all time points measured, α-chaconine treatment lead to an inhibition of BuChE of 68.3% at time point 0, 70.2% at 10 min, and 67.3% at 30 min. These results are indicative for a higher inhibitory of α-chaconine if compared to α-solanine.

The comparative approach of estimation of an inhibitory activity at a defined substrate concentration for both enzymes was used only once, meaning direct comparison of the inhibitory activity of α-chaconine and α-solanine (McGehee et al., 2000). Using human purified enzymes and substrate concentration of 100 µM, the authors determined the percentage of inhibition of AChE and BuChE. α-Chaconine inhibited 67.3% of AChE-activity and 92.8% of BuChE activity. α-Solanine showed a similar activity, inhibiting AChE by 76.5% and BuChE by 91.5%, pointing to the fact that BuChE may be more sensitive to inhibition by GAs than AChE.

Alozie et al. (1978) performed an inhibitory assay with 100 µM of α-chaconine on ChEs in serum, erythrocytes and brains of rats. By incubating electrophorese gels they were able to discriminate six isoenzymes. The inhibitory activities for isoenzymes 1–6 from plasma were 0%, 0%, 78.5%, 81.7%, 90% and 100%, respectively, from erythrocytes were 0%, 42%, 42%, 100%, 100% and 100%, respectively, and from brain were 23%, 27.1%, 15.6%, 19.5%, 67.3% and 100%, respectively.

Using ChE from erythrocytes, Roddick (1989) investigated three questions: (a) to compare the relative potency of α-chaconine and α-solanine alone and the individual activities to those of mixtures of both compounds, (b) to compare the activity of the glycosides to those of aglycones and, (c) to compare the relative potencies of the aglycones against bovine and human ChE from erythrocytes. α-Chaconine and α-solanine were tested at concentrations of 5 µM and 10 µM, as well as in a mixture containing 5 µM of each compound. α-Chaconine inhibited bovine erythrocyte ChE by 25.9% (5 µM) and by 54.4% (10 µM), whereas α-solanine inhibited by 27.7% (5 µM) and by 44.3% (10 µM). The 1:1 mixture of both compounds at 5 µM each lead to a 37.3% inhibition, indicative of an additivity of the two compounds. α-Solasonine and solamargine were tested at 50 and 100 µM each. While 50 µM concentrations of both compounds slightly increased enzyme activity, an inhibition was detectable at 100 µM, with 16.7% inhibition for α-solasonine and 13% inhibition for solamargine. In addition, Roddick (1989) comparatively assessed the inhibitory activity of the aglycones solanidine, tomatidine and solasodine. At 100 µM no consistent inhibitory activity using bovine and human erythrocyte AChE was detected.

3.1.5.1.2. Determination of inhibitory constants (K_i) for GAs on inhibition of ChEs in vitro

Most data for the assessment of the K_i are available for α-solanine. For human AChE, the range of determined K_i values stretches from a K_i of 5 µM (Orgell, 1963) to a K_i of 14 µM (McGehee et al., 2000). For bovine and equine AChE, the K_i values were determined to be 8.3 µM and 4 µM, respectively.

A lot more data is available for the K_i determination of α-solanine for inhibition of human BuChE. The range of the determined K_i values is between a higher nM to a lower µM range. McGehee et al. (2000) described a K_i of 0.17 µM for the inhibition of human BuChE by α-solanine. Benilova et al. (2006) determined an almost identical K_i of 0.22 µM for this enzyme using a biosensor-based method. A similarly low K_i was also reported by Loewenstein-Lichtenstein et al. (1996), although for an artificial mutant which was expressed recombinantly. The same authors determined a K_i of 3.3 µM for the wildtype human BuChE. This observation is in line with the findings of Sternfeld et al. (1997) who determined a K_i of 2.9 µM for the human placental BuChE and a K_i of 2.0 µM for the recombinantly expressed human BuChE. K_i values for the inhibition of equine BuChE were found to be higher (Benilova et al., 2006).

The K_i values determined for the inhibition of BuChE by α-chaconine were even lower than those for α-solanine. McGhee et al. (2000) reported a K_i of 0.066 µM for the purified human enzyme, whereas Benilova et al. (2006) reported a K_i of 0.17 µM. Like for α-solanine, human BuChE appears to react more sensitive than equine BuChE.

The data for α-tomatine are inconclusive. Orgell (1963) reported a K_i of 100 µM for human plasma ChE, whereas Benilova et al. (2006) determined a K_i of 1.3 µM for human BuChE and 1.66 µM for equine BuChE using a biosensor method.

In summary, biochemical data on inhibition of ChEs by GAs can be summarized as follows below and as illustrated in **Tables 25** and **26** (for details refer to **Appendix E**):

(i) the sugar moiety is a crucial determinant of ChE inhibition, as GAs are more potent inhibitors than their aglycones (**Table 25**),

(ii) α-chaconine seems to be a slightly more potent inhibitor of ChEs than α-solanine, particularly of BuChE, and

(iii) GA exposure leads to a slightly higher inhibition of BuChE activity than of AChE activity.

Table 25. Percent inhibition of bovine erythrocyte cholinesterase (ChE) by glycoalkaloids (GAs) and their aglycones (Roddick, 1989).

Compound	Concentration (µM)	% Inhibition [(a)]
α-Solanine	10	44.3
α-Chaconine	10	54.4
α-Solanine + α-chaconine	5 + 5	37.3
Solanidine, first sub-experiment	100	2–11
Solanidine, second sub-experiment	100	-14.1 – -6.2

(a): % inhibition of the activity measured for the solvent control DMSO.

Table 26. Inhibitory constants for glycoalkaloids (GAs) on human acetylcholinesterase (AChE) and human butyrylcholinesterase (BuChE) [(a)].

Compound	human AChE; K_i (µM)	human BuChE; K_i (µM)
α-Solanine	5–14 [(b)]	0.22–3.3 [(c)]
α-Chaconine	17 [(d)]	0.06–0.17 [(e)]

(a): In old papers (Orgell, 1963) BuChE was not known yet, data were reported as AChE activity.
(b): Orgell (1963) and McGehee et al. (2000).
(c): Löwenstein-Lichtenstein et al. (1996), Sternfeld et al. (1997), McGehee et al. (2000), Benilova et al. (2006).
(d): McGehee et al. (2000).
(e): McGehee et al. (2000), Benilova et al. (2006).

3.1.5.1.3. Inhibition of ChEs in vivo

Inhibition of ChEs in serum and erythrocytes

Inhibition of AChE by α-solanine is effective not only *in vitro* but also *in vivo*, as shown by several experimental studies. When α-solanine was applied i.p. to two rabbits of each sex, AChE activity declined over time both in serum and in erythrocytes and was lowest at time of death (range of lowest remaining activity: 55.2–73.2% in serum and 66.6–81.3% in erythrocytes). One male animal recovered. In this animal study AChE activity remained constant between time points 140 min after treatment and 210 min after treatment in the case of serum AChE, and even recovered from 71.4% to 82.9% in the same time range in erythrocytes (Patil et al., 1972).

Inhibition of ChEs in the central nervous system

As mentioned on **Section 3.1.3**, in humans acute effects of intoxication comprise sweating, vomiting, diarrhoea, severe abdominal pain, drowsiness, apathy, confusion, weakness, vision disturbances, fever, rapid and weak pulse, low blood pressure and bronchospasm in cases of milder intoxications, as well as

paralysis, respiratory insufficiency, cardiac failure, coma and death in severe cases (see **Section 3.1.3.1**). As only cholinergic neurons mediate both pre- and postganglionic activities of the parasympathic nervous system, and in addition, part of the preganglionic activities in the sympathic nervous system, it appears likely that anticholinolitic activities of GAs in this part of the nervous system result in the effects observed following human intoxication. Similar symptoms have been described for non-competitive organophosphorus inhibitors (Pereira et al., 2014). These effects may mechanistically explain the severe symptoms described for GA intoxication.

Since the 70ies experimental evidence was gained that GAs may inhibit ChE also in the central nervous system. In rats, treated i.p. with 0, 10, 30, 60, 90 mg/kg bw per day of α-chaconine, brain AChE activity dose dependently decreased at 10, 30 and 60 mg/kg bw to 79%, 55% and 18%, respectively (Alozie et al., 1978). These effects could not be related to the availability of ACh, as in Sprague Dawley rats receiving i.p. 3, 8 or 20 mg/kg bw of α-chaconine either three or 12 h prior to sacrifice, brain levels of ACh remained unchanged (Aldous et al., 1980). In addition to neurotransmitter levels in the brain, the authors assessed neuro- and electrophysiological parameters related to both brain function. After i.p. injection of 10, 20 and 30 mg α-chaconine/kg bw, both in the parietal and occipital EEGs the slow wave activity was affected at 20 mg/kg bw per day. The disappearance of the 6 Hertz peak in parietal records and the dose dependent disappearance of 6 Hertz peak in occipital records occurred at/started at 10 mg/kg bw per day (Aldous et al., 1980).

Baker et al. (1988) reported on effects on brain AChE activity in Syrian hamsters gavaged with increasing amounts of lyophilisized sprout material. At 300 mg/kg bw of the material, a statistically significant increase in brain AChE activity was observed, followed by significantly decreased activity at 400 and 500 mg/kg bw. This decrease was not as pronounced as the decrease seen for the organophosphate diethyl-tri-chloro-phosphorothioate (Dursban 2E), which was investigated against control in a parallel experiment and acts through a non-competitive mechanism.

Inhibition of cholinesterases in cardiac and neuromuscular transmission

The effect of α-solanine on the electrocardiogram (ECG) and respiration was also investigated in rabbits (Nishie et al., 1971). Below a dose of 15 mg/kg i.p., no effect was detectable. Tachycardia was observed both at low (10 mg/kg bw per day) and at very high doses (40 mg/kg bw per day). Intermediate doses (20 and 30 mg/kg bw per day) lead to bradycardia. Symptoms which became apparent at low doses were sedation, respiratory impairment and constriction of the abdominal muscles which became apparent at both 8 and 10 mg/kg bw per day. In conclusion, GAs may influence heart function at high systemic levels.

McGhee et al. (2000) investigated whether GAs would impact on mivacurium induced neuromuscular blockage. Female and male New Zealand white rabbits received an i.v. administration of mivacurium (27 nmol/kg in 30 sec), followed by an i.v. administration of 12 µmol/kg of α-solanine or α-chaconine for 20–60 min. A decrease in plasma BuChE, responsible for the detoxification, e.g. of cocaine, in response to infusion of the GAs resulted in elevated mivacurium levels and consequently for a prolonged time for recovery from mivacurium induced neuromuscular blockage. The effect of α-chaconine was thereby more pronounced than the effect of α-solanine.

3.1.5.1.4. Developmental and reproductive effects of GAs and their aglycones

Teratology occurred at relatively high doses of both GAs and their aglycones, and the teratogenic phenotypes observed included exencephaly, encephalocele, cebocephaly, cleft palate and anophthalmia (see **Section 3.1.2.3**). Experimental evidence suggests that these phenotypes in the central nervous system are presumably not, or not exclusively, caused by inhibition of the catalytic function of AChE. First, the AChE knock-out mouse does not show a phenotype in the central nervous system. High sensitivity against BuChE inhibitors indicates that this enzyme may be able to compensate for the

missing AChE (Xie et al., 2000). Comparative immunohistochemical examination between brain tissues of knock-out and wildtype animals not even revealed differences at the microscopic level in neuronal, dendritic, astrocytic, synaptic, microglial, and endothelial tissue compartments (Rice et al., 2007). AChE knock-out mice died around PND14 from their inability to obtain sufficient nutrients. This phenotype could be rescued by a special diet and enabled examination of older knock-out animals. The phenotypes detected comprised body tremors, abnormal gait and posture, absent grip strength, inability to eat solid food, pinpoint pupils, decreased pain response, vocalization, and early death caused by seizures or gastrointestinal tract ileus (Duysen et al., 2002), with body tremors resembling the human situation. Second, the teratologic phenotypes were caused by both the GAs and their aglycones, the latter being much weaker inhibitors of ChE catalytic activity than the GAs or do not inhibit at all (see **Appendix C**). However, the papers dealing with the phenotype of the AChE -/- mice point to the hypothesis that BuChE may substitute for AChE protect from excess acetylcholine (Mesulam et al., 2002). As GAs inhibit both enzymes, still the possibility remains that inhibition of ChEs is involved in causing the teratology, neurodevelopmental phenotypes.

There is increasing notion that ChE shows also non-cholinolytic enzymatic side activities like acylamidase activity (Vogel-Höpker et al., 2012; Layer et al., 2013). Furthermore, the number of descriptions of non-enzymatic roles of ChEs have increased over the years, including non-catalytic activities of the enzymes in processes of neuronal and non-neuronal developmental (Vogel-Höpker et al., 2012). Developmental neuronal functions comprise amongst others, involvement of proliferative control of neuronal cells, their migration, neurite outgrowth and synaptogenesis, as well as cell adhesion. Non-neuronal developmental functions concern association with the development of the immune system, hemopoietic stress responses, as well as of limb and bone development (Vogel-Höpker et al., 2012; Lionetto et al., 2013). It remains to be studied whether GA and their aglycones interfere with any of these non-catalytic functions of ChE and whether such effect could have impact on the neuronal development of the unborns.

Reproductive effects occurred at the relative low doses of 3.6 and 4.8 mg/kg bw per day, producing a decrease in the percentage of weaned pups, due to early death of the pups. This was presumably due to starvation as no milk was found in the stomach of these animals. Most clinically used AChE inhibitors represent competitive AChE inhibitors. They are believed to be compatible with human breast feeding, except for the use of neostigmine or combinations of neostigmine and atropine (Tsiaoussis et al., 2018). However, the use of AChE inhibitors is intended to increase the levels of Ach, and this increase in ACh may have an impact on lactation at three different levels. First, in animals, the systemic or central administration of ACh or AChE inhibitors leads a decrease in the prolactin release from the anterior pituitary gland (Grandison et al., 1974), which may be mediated through an increased oxytocin release from the neurohypophysis. This lowering of prolactin may affect the growth and lactation within the mammary gland. Second, the sustained reflex-release of oxytocin during suckling leads to an increase in intramammary pressure through muscarinic and nicotinergic AChE receptor mechanisms (Clarke et al., 1978). And third, by the decreased prolactin release the transcription of casein mRNA is decreased or eliminated, leading to a decreased synthesis of α-lactalbumin.

Negative (or dysfunctional) effects of solasodine on fertility were accompanied by morphological alterations in the testes, the epididymis and by reduced sperm quantity and particularly quality in dogs and monkeys. The mechanisms causing these effects are largely unknown. Inhibition of ChEs as cause of this male reproductive phenotype can presumably be ruled out, because solasodine is a poor inhibitor of ChEs. Similar findings to those reported in the studies with Rhesus monkey and with orally treated dogs as described above (Dixit et al., 1989) were observed in dogs following i.p. injection of 20 mg/kg bw per day solasodine (Dixit and Gupta, 1982). In both dog studies the authors interpreted their findings as anitiandrogenic based on the reduced sialic acid content, an androgenic response marker. This interpretation is plausible as in both studies the number of androgen producing Leydig cells is decreased.

On the other hand, i.p. or oral treatment of animals with solasodine and testosterone propionate did not restore the testicular and epididymal phenotypes, pointing to the fact that hormonal effects may be involved in the disruption of male reproductive organ function by solasodine, but hormonal effects alone may not be sufficient to explain the observed effects entirely. Further, an effect of GAs on male reproductive organs may be compound specific. An i.p. treatment of mice with 2.5 mg/kg bw of α-solanine every third day for 10 times did not result in a reduced testis weight, but decreased mRNA levels of cyclin D1 and of Inhibin A and B in the testis, which is in line with the hypothesis that hormonal effects contribute to the observed effects (Park et al., 2009). Finally, i.p. injection of 50 mg/kg bw per day of α-solamargine showed no effect on the weight of the testes and epididymis or the number of spermatozoa in male Wistar rats (Al Chami et al., 2003).

In summary, α-solanine in rats impacts on the availability of milk to the pups. Regarding the mechanisms underlying the effects of solasodine with the structure and function of male reproductive organs, little is known.

3.1.5.1.5. Inhibition of cholinesterases and effects in the immune system

Plant extracts of the peeled tuber of *S. tuberosum* as well as GA and aglycones exerted pronounced anti-inflammatory effects in *in vitro* and *in vivo* experiments (as outlined in **Section 1.1.1.3**). With regard to the putative mode of action of these anti-inflammatory effects, it should be noted that a tight cross-talk between the immune system and the cholinergic neurotransmitter system has been demonstrated, the so-called 'anti-inflammatory non-neuronal cholinergic pathway'. In particular, stimulation of a specific acetylcholine-receptor (α−7 nAChR) suppresses the release of pro-inflammatory mediators from macrophages. The stimulation of α−7 nAChR may be due to non-neuronal ACh, which is secreted by many cell types, such as macrophages, lymphocytes or dendritic cells. These cells also express AChE and BuChE to regulate the levels of ACh. There are reports that an increased BuChE activity may suppress the cholinergic anti-inflammatory responses and amplify systemic inflammation, a pathogenetic mechanism discussed for multiple sclerosis and other diseases (Pohanska, 2014; Reale et al., 2018;). It may be speculated that the GA induced lowered BuChE activity may result in an anti-inflammatory outcome. The mode of action underlying the anti-inflammatory effect of the aglycones, with weak impact on BuChE activity, remains to be clarified.

3.1.5.3. Interference with metabolism

Elkahoui et al. (2018) tested whether dietary supplementation with potato peel powder would impact on body weight gain in mice on high fat diet containing 25% of lard. The powder was prepared from four different potato cultivars with well-defined amounts of α-solanine and α-chaconine. The isocaloric and isonitrogenous peel-containing diets dose dependently reduced both weight gain and epididymal adipose tissue in male C57B1/6J mice, although the food consumption remained unaltered. Similar observations were reported for α-solanine (Gull, 1960) or α-chaconine (Friedman, 1996; Langkilde et al., 2008). The reasons for the lowered gain in body weights in GA-treated rodents are presently unclear.

Recently, it was discovered that BuChE acts also as an esterase of ghrelin (Brimijoin et al., 2016, 2018). Ghrelin is an orexigenic hormone, produced by the pancreas and mucosa of the stomach and stimulating the appetite via inducing the secretion of neupeptide Y in the hypothalamus. Immediately after food intake, ghrelin is degraded into smaller peptide fragments, involving the activity of BuChE. Ghrelin plays a role also in the regulation of glucose homeostasis through the modulation of insulin secretion and insulin sensitivity (Heppner et al., 2011). It is presently unclear whether BuChE inhibition by GAs may affect food intake, body weight and the metabolism via altered ghrelin levels and/or impact on insulin sensitivity.

There is experimental evidence that dietary α-tomatine forms a non-absorbable complex with cholesterol and other sterols in the gut (see **Sections 3.1.2.2** and **3.1.2.9**). It appears likely that the underlying mechanisms are similar to those described in **Section 3.1.5.5**. The insoluble complexes in the intestinal lumen are excreted via feces. Accordingly, an α-tomatine-containing diet elevated the fecal elimination of cholesterol and sterols in rats and hamsters (Friedman et al., 2000a). While α-tomatine did not affect bile acid secretion in the rats, in hamsters there were significant increases in fecal deoxycholic and lithocholic acid content after treatment with green and/or red tomatoes (Friedman et al., 2000b). This supports that α-tomatine interferes with enterohepatic recirculation of sterols.

In rats, the increased fecal efflux and the concomitant decreased uptake of dietary cholesterol by the liver was counteracted by an increased de-novo synthesis of hepatic and intestinal cholesterol (Cayen et al., 1971). Nevertheless, plasma HDL-cholesterol and triglycerides were reduced when rats received α-tomatine via diet for a period of 14 days. α-Tomatine also lowered plasma levels of LDL-cholesterol and triglycerides in hamsters, while HDL cholesterol remained unchanged (Friedman et al., 2000a,b).

To conclude, α-tomatine reduces enteral absorption of cholesterol and serum cholesterol levels in rodents. However, it remains to be studied whether similar effects of α-tomatine or other GAs are to be expected in humans.

3.1.6. Considerations of critical effects and dose-response analysis for the human risk assessment

3.1.6.1. GAs from edible parts of *S. tuberosum*

3.1.6.1.1. Considerations of critical effects and dose-response analysis

Acute toxicity

In humans, the limited human data available from kinetic studies in volunteers (2–3 per treatment) showed that potato TGA (50% α-solanine; 50% α-chaconine) administered as a solution at 0.30, 0.50 or 0.70 mg/kg bw p.o. did not elicit symptoms. Bitter taste, gastric discomfort and nausea may occur within ~30 min after ingestion of ~1 mg potato TGA/kg bw contained in a potato meal. Subsequently the volunteers developed diarrhoea and started to vomit. A number of reports on outbreaks and cases of intoxications substantiate that acute toxic effects may occur in adults from a total potato GAs intake of 1 mg/kg bw or more. In severe cases of intoxications hallucinations, paralysis, respiratory insufficiency and cardiac failure have been reported. Doses in the range of 3–6 mg potato TGAs/kg bw are considered to be potentially lethal for humans. Results from limited volunteer studies suggest differences in the human population with respect to the individual susceptibility towards adverse effects associated with the intake of potato GAs.

In experimental animals, no lethality was observed in mice at a single dose of 1,000 mg α-solanine/kg bw p.o., and no effects at 500 mg of solanidine/kg bw p.o. In rats, no adverse effects were reported for a single oral application of 250 mg/kg bw of α-solanine. Diarrhoea occurred after application of α-solanine/α-chaconine mixtures at 300 mg/kg bw and above (see **Section 3.1.2.1**). This indicates acute toxicity of GAs at doses above 250 mg/kg bw in rats and mice with diarrhoea being the most frequent symptom.

The acute symptoms, described in humans and experimental animals, may be due to the ability of α-solanine and α-chaconine to complex with membrane sterols, thereby causing disruption and loss of integrity of cell membranes. Deletion of one or more mono-saccharides from the GAs resulted in reduced activity. After oral exposure of potato GAs, these effects may lead to local irritation of the mucosa of the gastrointestinal tract leading to gastric discomfort, vomiting and diarrhoea. The adverse effects may

also be due to the inhibitory effect of α-solanine and α-chaconine on the activity of AChE. The aglycone solanidine exerted almost no inhibitory effects, suggesting that the sugar side chain is obligatory for inhibiting the AChE activity. As a consequence of the impaired termination of the cholinergic transmission at the neuronal and neuromuscular junctions, intoxications with α-solanine and α-chaconine might lead to the development also of a cholinergic toxidrome (see **Section 3.1.5**), with diarrhoea and vomiting being predominant. Further symptoms described in case reports and outbreaks may have developed secondary to dehydration, such as low blood pressure, dizziness, cardio-respiratory insufficiency, or coma. However, some prototypical symptoms of a cholinergic toxidrome that may not develop secondary to dehydration, such as miosis and fasciculations of the skeletal musculature, have not been reported so far.

In mice, the aglycone solanidine exerted no effect at 500 mg (1.26 mM)/kg bw i.p., contrasting to the pronounced toxicity of GAs after the identical route of exposure, i.e. LD_{50} of α-solanine was 42 mg (48.3 µM)/kg bw, and of α-chaconine was 32.3 mg (37.9 µM)/kg bw. The relative potency of toxicity of α-solanine and α-chaconine appear to be similar also in rats and rabbits, e.g. in rats LD_{50} for α-solanine and α-chaconine were reported to be 67 mg (77.2 µM)/kg bw and 84 mg (98.6 µM)/kg bw, respectively. These findings are substantiated by (i) a similar potency of α-solanine and α-chaconine to interact with sterols and disintegrate cellular membranes and act as inhibitors of AChE activity and (ii) the weak effects of the aglycone with regard to membrane disruption and AchE inhibition. Based on these experimental findings, the CONTAM Panel considered that (i) the aglycone solanidine is of lower acute toxicity than the respective GAs and that (ii) there is the potential for additivity of α-solanine and α-chaconine, recognising that they share common modes of action and exert similar adverse effects. Therefore, the CONTAM Panel performed the risk assessment on the basis of the total content of GAs (TGA) in potatoes comprising α-solanine and α-chaconine, assuming equal potency.

When comparing experimental rat and human data on acute oral toxicity of potato TGAs it may be deduced that:

(i) in humans, gastric discomfort and nausea occurred rapidly after ingestion of ~1 mg/kg bw, followed by vomiting and diarrhoea. Studies in volunteers give some indication on differing individual susceptibilities towards adverse effects of GAs,

(ii) gastric discomfort and vomiting are symptoms not covered by bioassays in rats. In this species the most frequent symptom is diarrhoea occurring at a single oral potato TGA application of 300 mg/kg bw,

(iii) based on the likelihood to develop diarrhoea after ingestion of potato TGA, humans show an approximately 300-fold higher susceptibility towards this adverse effect of potato TGA than rats.

The reasons for these marked species differences are presently unclear. It may relate to anatomical, histological and/or functional features of the gastrointestinal tract differing considerably between humans and rodents, e.g. the unique murine non-glandular forestomach, an intragastric pH around 3–4 in rodents and around 1 in humans, and a relatively long small and large intestine and a large caecum in rats and mice. These factors may affect the local concentration of the GA in the gastrointestinal tract and the vulnerability of the mucosa (Hugenholtz and de Vos, 2018). The differences in susceptibility may also be due to a relatively high oral bioavailability and low clearance rates of potato TGA in humans when compared to rodents. Furthermore, it is not well documented whether there are differences in the sensitivity of human and rat AChE towards the inhibitory effects of GAs.

Due to these uncertainties, the CONTAM Panel considered that the use of rodent data on acute toxicity was not appropriate to establish a reference point for acute exposure in humans.

Based on human data on case reports, outbreaks and studies in volunteers, the CONTAM Panel selected the LOAEL of 1 mg potato TGA/kg bw per day as the reference point for acute exposure to potato TGAs via food.

Repeated dose toxicity

In humans, no evidence of health problems associated with repeated or long-term intake of GAs via potatoes has been identified. Thus, the CONTAM Panel was not able to identify critical chronic effects in humans.

In experimental animals, toxicological studies have been carried out in rodents and monkeys using different experimental designs with administration of different doses and with durations of up to 90 days. However, no long-term chronic toxicity or carcinogenicity study for GAs or their aglycones could be identified.

The purity of the GA preparations tested varied considerably. Furthermore, a number of studies were performed according to non-standard experimental protocols and do not provide results that are suitable for risk assessment purposes.

In repeated oral dose toxicity studies, the main effects for α-solanine and α-chaconine were reduced body weight gain (accompanied by unaltered or, as reported occasionally, reduced feed intake) and decreased absolute and relative liver weight. Solanidine lowered also food intake and body weight gain but elevated relative and absolute liver weights.

In hamsters, mortality was reported after oral administration of 100 mg/kg bw per day of α-solanine or α-chaconine given for 5 days. In rats, an oral dose of 90 mg α-solanine/kg bw per day applied over 5 weeks did not cause any effect. Mice exhibited reduced body and absolute liver weights at 416.6 mg α-solanine/kg bw per day, and reduced body and absolute and relative liver weights at 409 mg α-chaconine/kg bw per day, applied over 7 days. These findings indicate that hamsters may be more sensitive towards the adverse effects of α-solanine and α-chaconine than mice and rats. This is substantiated by a marked species difference in the toxicokinetics of the GAs, with hamsters showing higher oral bioavailability and lower clearance rates of GAs than rats (see **Section 3.1.1.1**).

Relative potency of oral toxicity of α-solanine and α-chaconine can be partly deduced from two independent studies with durations of up to 7 days. Identical doses of these two GAs elicited comparable effects (i.e. reducing the body weight) at a similar frequency in the two species tested (mouse and hamster). These findings are supported by a similar potency of α-solanine and α-chaconine with regard to acute toxicity. Oral administration of solanidine did not lower but increased the absolute and relative murine liver weight, which is some indication that the adverse effects in repeated dose studies may differ between the GA and the respective aglycones.

A NOEL for potato TGA of 33.3 mg/kg bw per day was identified by Langkilde et al. (2012). This study was performed in hamsters over a period of 90 days, the longest duration of all studies identified. The animals were treated via diet, thus accounting for dietary matrix effects and simulating GA uptake via food. However, the CONTAM Panel noted several limitations of the study: the animals were dosed with freeze-dried powder of two different potato cultivars, partly spiked with commercial standards of α-solanine and α-chaconine to obtain different ratios between α-solanine and α-chaconine from 1:3.1–1:70. To obtain these highly variable ratios, different proportions of potato powder of the two cultivars were admixed to the chow, creating considerable nutritional imbalances between the treatment and the control groups (for details see **Section 3.1.2.2.1**). In addition, the authors did not state whether the potato powder contained trypsin inhibitor activity or not. All other studies identified were either not on hamster, of short duration and/or applied GAs via bolus not mimicking intake of GAs via the diet. Thus,

the CONTAM Panel could not identify a study on GAs of sufficient quality to derive a reference point for repeated dose toxicity.

For the aglycone solanidine, there are only two toxicity studies in female mice over a period of 28 days. It is considered that the sugar moieties of the ingested α-solanine and α-chaconine are cleaved off during metabolism and that any toxicity testing of the two GAs will cover the adverse effects of solanidine. Therefore, the CONTAM Panel considered it was not appropriate to identify an additional reference point for a human risk assessment of solanidine.

To conclude, the data available for repeated dose toxicity are not sufficient to identify a reference point for chronic exposure to potato GAs.

Genotoxicity

Several assays were performed to test for mutagenicity and clastogenicity of potato GAs, the aglycone solanidine, as well as extracts of *S. tuberosum*. No genotoxicity was observed in the available tests. However, some tests provided limited information (see **Section 3.1.2.7**, **Table 18**). Overall, the CONTAM Panel concluded that there is no evidence of genotoxicity for the potato GAs α-solanine and α-chaconine, and for the aglycone solanidine.

Developmental toxicity

Results from limited developmental toxicity studies with potato GAs in experimental animals show effects at high oral doses. In hamsters, severe malformations, affecting mainly the central nervous system, occurred in offspring when dams received 200 mg α-solanine/kg bw or 165 mg α-chaconine/kg bw per day on GD7.5/8. The teratogenic phenotypes comprised exencephaly, encephalocele, cebocephaly, cleft palate and anophthalmia. Interestingly, the aglycones solanidine and demissidine caused also malformations in the central nervous system in hamsters. Exencephaly, encephalocele and/or anophthalmia occurred in offspring at maternal doses of 176 mg demissidine/kg bw or 115 mg solanidine/kg bw applied orally on GD8 (see **Section 3.1.2.3**).

There are no dose-response studies, no indications that species other than hamster may be affected, and no mechanistic explanations for the malformations described above.

Several epidemiological studies failed to provide evidence that the intake of potato GAs may be an aetiological factor in the pathogenesis of NTD. Two studies indicated a possible association between maternal consumption of sprouted potatoes during the periconceptional phase and an elevated risk for NTDs or orofacial clefts. However, incomplete reporting impairs any conclusion on causal relationship of GA exposure and developmental effects from these studies.

3.1.6.1.2. Derivation of a health-based guidance value (HBGV) or margin of exposure (MOE) approach

Based on the available information, the CONTAM Panel considered the LOAEL of 1 mg potato TGA/kg bw per day based on human data from case reports, outbreaks and studies in volunteers, as the reference point for acute exposure to potato TGAs via food.

The available data on acute toxicity were considered insufficient to establish an HBGV, and instead the Panel used the margin of exposure (MOE) approach to assess a possible health concern from acute exposure to potato TGAs via food.

An MOE higher than 10 indicates that there is no health concern. This MOE of 10 takes into account the interindividual variability in toxicodynamics (a factor of 3.2), and extrapolation from a LOAEL to a NOAEL (a factor of 3). As outlined in **Section 3.1.6.1.1**, the CONTAM Panel considered that the critical effects of acute exposure to potato TGAs may be mainly due to the local effects in the gastrointestinal tract,

following probably local irritation rather than inhibition of AChE activity. Thus, the possible interindividual variability in toxicodynamics is more relevant for the evaluation of the MOE than the interindividual variability in toxicokinetics.

No reference point could be identified to characterise the risk following chronic exposure.

3.1.6.2. GAs from edible parts of food plants other than *S. tuberosum*

3.1.6.2.1. Considerations of critical effects and dose-response analysis

With respect to GAs from food plants other than *S. tuberosum*, reports of intoxications or outbreaks, on the effects of acute or chronic ingestion of tomato- or aubergine-derived GAs or aglycones or relevant epidemiological studies are missing.

In experimental animals, acute and repeated dose toxicity studies on relatively high doses of tomato GAs showed mainly reduced absorption of cholesterol and bile acids from the gastrointestinal tract and lowered blood lipid levels.

With regard to the mode of action, tomato and aubergine GAs interfere with sterols in cell membranes and/or the lumen of the gastrointestinal tract, which may impair the resorption and enterohepatic recirculation of cholesterol and bile acids. The inhibitory effect on ChE activity is much weaker for tomato and aubergine GAs than for potato GAs.

Experimental developmental toxicity studies indicate that in hamster severe malformation (mainly of the central nervous system) occurred in offspring, when dams received doses of 1,400 mg of solasodine or dihydrosolasodine per kg bw per day on GD8. However, dose-response studies, studies in other experimental species, studies covering not only one or few days of the gestation period and mechanistic studies are missing.

Adverse effects on male fertility (decreased epididymal weight and cauda epididymal epithelial height, lack of sperm in epididymal lumen) occurred when dogs or rhesus monkeys received the aglycone solasodine orally at 80 or 100 mg/kg bw per day, respectively.

From the limited number of studies available, there was no evidence for genotoxicity of the aubergine GA, α-solamargine.

No data for determining a reference point for GAs or aglycones occurring in edible parts of food plants other than *S. tuberosum* are available.

3.1.7. Consideration of critical effects and dose-response analysis for the farm animal, horses and companion animals risk assessment

The CONTAM Panel reviewed the available studies in order to identify a NOAEL or LOAEL for farm animals, horses and companion animals.

No studies were identified in horses, companion animals (cats and dogs) or fur animals.

The few studies identified on the adverse effects of GAs in ruminants, pigs, poultry and fish could not be used for the identification of a reference point.

For rabbits, no studies have been located other than those described under acute toxicity (i.p or i.v.) and repeated dose toxicity (see **Sections 3.1.2.1** and **3.1.2.2**). No reference point could be identified.

3.2. Occurrence data

3.2.1. Occurrence data submitted to EFSA

Occurrence data on food

An initial number of 1,300 results for 651 food samples analysed on GA from three European countries were available for the assessment. The major contributor of data on GA in terms of number of results was Germany (73% of the results) while the Netherlands and Sweden contributed with 21% and 6%, respectively. Data were reported on samples collected between the years 2005 and 2017. Only data on GAs in potato or potato products were reported. No data on GAs present in tomato or aubergine or products were reported.

Free text information was analysed to better specify the food category to which the sample was referring. The number of samples and results available for each food category are shown in **Table 27**.

Table 27. Number of samples and results analysed for the glycoalkaloids (GAs) content, submitted to EFSA through the continuous call for annual collection of chemical contaminant occurrence data in food.

Foodex category	Results	Samples
Potatoes and potatoes products	14	7
New potatoes	171	86
Main-crop potatoes	1,071	536
Potato flakes	2	1
Mashed potato powder	16	8
Ready-to-eat meal for infants and young children	6	3
Ready-to-eat meal for children, vegetable-based	4	2
Ready-to-eat meal for children, cereal-based	2	1
Ready-to-eat meal for children, meat/fish-based	14	7
Total	**1,300**	**651**

All samples were analysed for both α-solanine and α-chaconine besides two samples that were analysed only for α-solanine and were excluded from the final cleaned dataset as the total GA content (sum of α-solanine and α-chaconine) was used for the exposure assessment.

A total of 86 samples were available for 'New potatoes'. Considering the uncertainty linked to the reporting of this category in the consumption database, the CONTAM Panel decided to combine the occurrence of 'New potatoes' with that of 'Main-crop potatoes'. Thus, in the following text samples from these two categories will be referred to as samples from main-crop potatoes.

A total of 29 samples referred to potato products including only few food categories plus a generic 'Potatoes and potatoes product', thus not covering the variety of composite food categories containing potatoes in the Consumption Database. Based on this, the CONTAM Panel decided that the best approach for the exposure assessment would be to use the occurrence values in the RPC, main-crop potatoes (combined with the new potatoes as described above) and the RPC Consumption Database (see **Section 2.3.1**).

Two samples were removed because they were reported as sampled based on 'Suspect sampling'.

In addition, 7 main-crop potato samples were excluded as they were reported as 'processed', thus not relevant for the calculation of the occurrence in the raw primary commodity.

Information about the potato cultivar was available for 210 samples and only few samples were available for each cultivar. Information on the cultivar is also not reported in the Consumption Database, thus the CONTAM Panel considered any analysis based on possible difference among cultivars not feasible. Information about the growing method (available for only 86 samples) was not taken into account in the analysis for the same reasons.

Specific information about the peeling was available for 40 samples out of 619 samples. Of these, only 7 were reported as 'peeled' potatoes. However, most of the samples were reported explicitly as 'unprocessed'. The CONTAM Panel assumed samples for which no information about the peeling nor the processing was available (156 samples), to be unpeeled and they were included in the final dataset. The 7 samples of peeled potatoes were excluded.

The occurrence data of total GAs (sum of α-chaconine and α-solanine) in unpeeled main-crop potatoes was used for the exposure assessment. The final dataset contained 605 samples referring to unpeeled potatoes and individual occurrence values are available in **Annex A4**. A summary of the available samples by country and year is provided in **Table 28**. Summary statistics of the occurrence UB values for main crop-potatoes (merged with new-potatoes) in the final dataset used in the assessment and for the other food categories for which data were available are provided in **Table 29**.

Table 28. Number of samples by country and year of sampling of the final GAs occurrence dataset.

Country/Year	2005	2007	2015	2016	2017	Total by country
Germany	222	8	8	9	185	**432**
Netherlands	-	-	81	52	-	**133**
Sweden	-	-	36	-	4	**40**
Total by year	**222**	**8**	**125**	**61**	**189**	**605**

Table 29. Summary statistics of the total potato glycoalkaloid (potato TGA) concentrations (mg/kg, upper bound) for 'Main crop-potatoes' (merged with 'New-potatoes') in the final dataset used in the assessment and for the other food categories for which data were available.

Food category	N	Mean	Min	Max	P95[(a)]
Main-crop potatoes (including new potatoes)	605	52.0	1.1	550.3	117.0
Potatoes and potato products	7	214.7	85.0	381.3	-
Potato flakes	1	95.6	-	-	-
Mashed potato powder	8	73.9	14.0	205.0	-
Ready-to-eat meal for infants and young children	3	0.8	0.1	1.4	-
Ready-to-eat meal for children, vegetable-based	2	3.3	2.0	4.6	-
Ready-to-eat meal for children, cereal-based	1	0.1	-	-	-
Ready-to-eat meal for children, meat/fish-based	7	1.1	0.1	2.7	-

(a): The 95th percentiles obtained on occurrence data with fewer than 60 analytical results may not be statistically robust (EFSA, 2011b) and are therefore not reported in the table.

The mean UB occurrence in the RPC main-crop potatoes was 52.0 mg/kg with a P95 of 117.0 mg/kg. The minimum and maximum reported concentrations were 1.1 mg/kg and 550.0 mg/kg, respectively.

As described in **Section 2.5**, reduction factors for peeling and heat processing were applied to these occurrence data.

Distribution of α-solanine and α-chaconine in the food samples

The samples in the final dataset were analysed for the distribution between α-solanine and α-chaconine, using the UB results. In **Figure 4** the results are expressed as ratio α-chaconine to α-solanine, while in **Figure 5** the results are expressed as the contribution of α-chaconine to the total GA content (sum of α-solanine and α-chaconine) in the potato samples. From **Figure 4** and **5** it is evident that there is a wide range reported for the ratio of α-chaconine to α-solanine. The ratio reported ranges from 0.015 to 7.4, equivalent to a relative contribution of α-chaconine to the TGA ranging from 1.5% to 88.1%. The average ratio of α-chaconine to α-solanine is 1.39 and the median is 1.28. The average contribution of α-chaconine to the total is 51.5% and the median contribution is 56.2%, showing a slight overall preferrence for α-chaconine in the samples. In 52.7% of the samples α-chaconine contributes between 50% and 70% to the TGA.

It was noted by the CONTAM Panel that in the database there are 22 samples with a very low α-chaconine content compared to the α-solanine content (**Figure 5**) while there are no samples with a similarly low content of α-solanine. This is related to the fact that in 21 samples α-chaconine was not detected above the LOQ of the method, while this was the case for α-solanine in only four samples. In these cases the UB value (LOQ) of the method is used. It should be noted that α-chaconine seems more susceptible to enzymatic degradation reactions (producing β-chaconine and solanidine) than α-solanine (Friedman and McDonald, 1995; Swain et al., 1978). Partial degradation of α-chaconine may occur during sample preparation and analysis unless specific conditions are applied (Friedman and McDonald, 1997; see also **Section 1.3.2.1**).

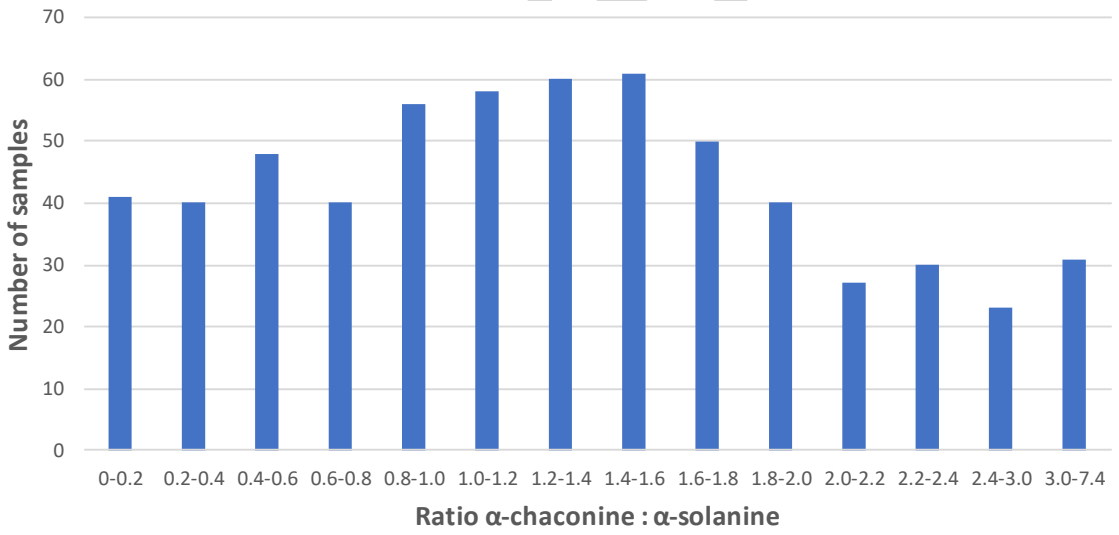


Figure 4. Ratio α-chaconine to α-solanine in potato samples present in the EFSA occurrence database (N = 605).

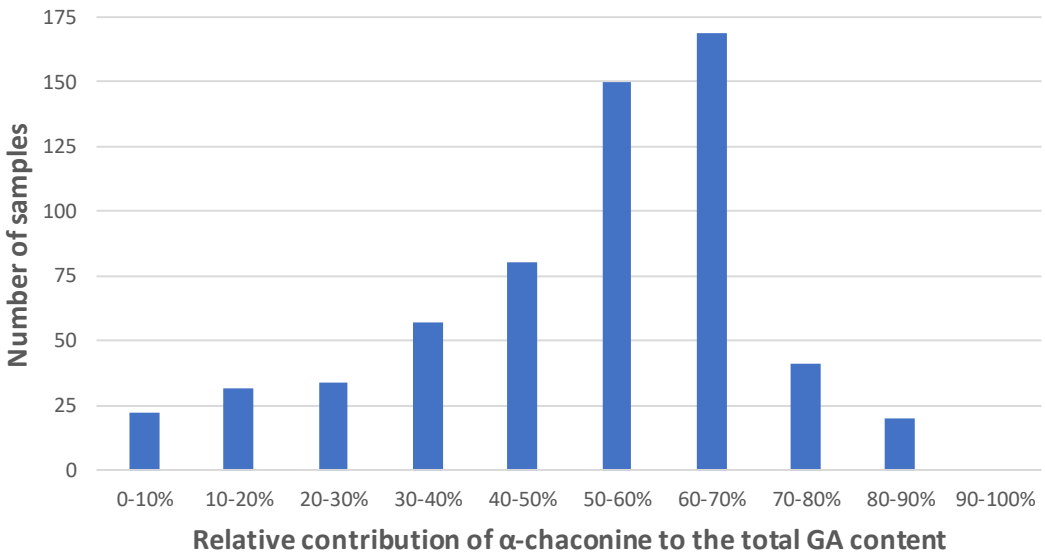


Figure 5. Relative contribution of α-chaconine to the total glycoalkaloid (GA) content in potato samples present in the EFSA occurrence database (N = 605).

Additional occurrence data received

The European Starch Industry Association submitted data to EFSA concerning 1,728 samples including samples on dietary fibre and potato proteins, pulp, juice and starch and on potatoes used for starch production (**Table 29**). Detailed results are available in **Annex A5**.

Due to uncertainty on the occurrence data reported (e.g. it was not known if the occurrence data referred to dry or wet weight and it was not always known if the samples referred to feed or food for human consumption) and the difficulties expressed by the data provider in retrieving this information, these data were not included in the exposure assessment. Mean occurrence values for each category are provided in **Annex A**. Values should be taken with caution as individual values might have referred to different concentration expressions (information on the water content of the samples was not provided, data may reflect results based on dry vs wet weight).

Table 29. Number of samples submitted by the European Starch Industry Association by product type and country of sampling for which total glycoalkaloid (GA) occurrence was available (14 samples reporting on only one GA were excluded from the table).

Product/Country	DK	FI	FR	DE	NL	SE	Total
Dietary fibre	6	-	-	40	-	7	53
Isolated proteins and other protein products	-	-	-	-	505	-	505
Potato juice, concentrated (feed)	-	-	-	-	27	-	27
Potato protein (feed)	25	1	45	55	415	28	569
Potato pulp (feed)	-	-	9	-	-	-	9
Potato starch	-	1	10	49	41	-	101
Potatoes (for starch production)	-	-	-	-	450	-	450
Total	**31**	**2**	**64**	**144**	**1,438**	**35**	**1,714**

3.2.2. Previously reported occurrence data in the open literature

3.2.2.1. Literature on occurrence data on food

3.2.2.1.1. Occurrence data on GAs in potatoes

Occurrence data on GAs in potato samples obtained from field studies

In this section studies are discussed that focussed on the effect of pre-harvest factors on the GA content of potatoes, such as the genotype, year-to-year variability, location, altitude, environmental conditions and farming system under controlled field conditions. In **Table 30** an overview of the studies that have investigated the TGA content of potatoes under field conditions is presented. The studies cover the effects on the TGA content of cultivars grown on different locations, in different years and with different production methods (e.g. organic, integrated or conventional). The studies which investigated the effects of location and environmental conditions are described, while those studies focussing mainly on the effect of the farming system are discussed under a separate heading. In addition, data from the registration and authorisation of seed potatoes bred in the Netherlands are presented, that includes the measurement of TGA levels under field conditions.

Effect of location, environmental conditions and year-to-year variability on the GA content of potatoes

Sinden and Webb (1972) conducted a large 2-year field trial including 5 commercial cultivars grown on 39 locations in the USA and Alaska. Overall, the average TGA content ranged from 43 mg/kg for cultivar 'Red Pontiac' in 1970 to 100 mg/kg for the cultivar 'Kennebec' in 1971. For 3 cultivars tested, relatively small variations between years for the average TGA content were found. A significant correlation between the location and the TGA content was noted, with potatoes from Alaska and North Dakota containing the highest average TGA content for the 5 cultivars (150 mg/kg and 143 mg/kg, respectively), while lowest TGA contents were found in Florida (31 mg/kg) (data for 1970). Important factors influencing the GA content were the (local) climatic conditions, length of daylight and soil type. A cold and wet, as well as a hot climate appears correlated with an elevated TGA content, as well as locations with extended day lengths of 18–20 h. Higher TGA contents can be associated with immaturity of the tubers, which is in turn related to the climate conditions and extended daytime length (Sinden and Webb, 1972).

The importance of location and the environmental conditions was also noted in a German study covering several locations over a span of 5 years (Ross et al., 1978). The variation in TGA content between locations amounted to 50% and the variation between years (strongly related to the weather conditions) to 78%. A 3-year study conducted in the UK also showed that although the TGA content is primarily determined by the genotype (cultivar), location and environment are important co-influencing factors (Bintclife et al., 1982; Parnell et al., 1984). A significant yearly variation was found in a 3-year study conducted in Hungary with 5 different cultivars (Tömösközi-Farkas et al., 2006). This was also concluded from a 2-year study on 6 early potato cultivars in Sweden, as the TGA content for the 6 cultivars in 1986 was 30–130% higher compared to 1987 (Hellenäs et al., 1995a). Papathanasiou et al. (1999) noted a significant difference in the amount of yearly variation in TGA content for two cultivars ('Pentland Dell' and 'Estima') studied. The cultivar 'Pentland Dell', which has intrinsically a higher TGA content, was much more sensitive to differences in environmental conditions than the cultivar 'Estima', which has a low TGA content. The effect of location on 14 cultivars of different flesh-colours (white/yellow, red or blue) was investigated during a 3-year period by Hamouz et al. (2014). With respect to TGA content, genotype was the most important factor, followed by the year-to-year variation, while location was the least important. In a follow-up study the effect of altitude, climatological conditions and soil type using

16 cultivars on two locations (298 m–460 m) were studied over a 3-year period (Urban et al., 2018). The authors concluded that differences found in TGA content were mainly correlated to the genotype, followed by differences in environmental conditions and the soil type. Ieri et al. (2010) studied the TGA content of 9 purple or red skinned cultivars, grown on 2 locations in Italy. The average TGA content was slightly higher in the tubers grown in central Italy (100 mg/kg) compared to the tubers grown at higher altitude (1,000 m) in the south (91 mg/kg). It was noted that the difference was mainly related to the TGA content in the flesh of the tubers, which averaged to 8.6 mg/kg in south Italy and 28.3 mg/kg in central Italy. Panovska et al. (1997) studied the effect of altitude (110 m, 460 m, 860 m) on the TGA content of 11 cultivars, but no significant effects were noted.

Several studies reported besides the TGA content, also the concentration of α-chaconine and α-solanine. From these data the ratio of α-chaconine to α-solanine was calculated. The reported ratio of α-chaconine to α-solanine ranges from 0.30 to 5.93. From the data it follows that, generally, α-chaconine is present in higher concentrations than α-solanine. There is however a substantial variation in the ratio between α-chaconine and α-solanine, not only for individual samples within a study, but also between studies.

In the Netherlands, the Nederlandse Algemene Keuringsdienst (NAK) (Dutch General Inspection Service) is responsible for the registration and authorisation of seed potatoes. New cultivars are grown for at least two years at two locations under controlled conditions together with the reference potato cultivar Innovator, which has an average TGA content of 152 mg/kg. To be allowed to the market the TGA content of the new cultivars must be lower than the TGA content of the Innovator cultivar. TGA content is determined according to the HPLC-UV method as described by Houben and Brunt (1994).

The NAK list of registered potato cultivars in 2018 [23] contained 552 entries, and for 464 cultivars the TGA content is listed. Before 2000 it was not mandatory to establish the TGA content and in particularly for older cultivars this information has not been recorded. The TGA distribution in the cultivars is shown in **Figure 6**, where it can be seen that the large majority of potatoes grown under the Dutch test conditions have a TGA content of <100 mg/kg. The average content of the registered cultivars is 48.4 mg/kg.

The NAK list [25] includes also 77 potato cultivars grown specifically for starch production. For 70 cultivars the TGA content is listed. Starch potatoes [24] contain higher amounts of GAs: the average TGA content is 107.7 mg/kg, while 34 cultivars exceed 100 mg/kg and 7 exceed 200 mg/kg. The cultivar 'Kuras' has the highest TGA content (338 mg/kg) and also the cultivars 'Aventra' and 'Elles' (both 260 mg/kg) are also high in TGA content.

Overall, it can be concluded from the field studies that genotype is the most important factor determining the TGA content. However, also environmental conditions, location and year-to-year variability contribute to the differences in TGA content observed during the pre-harvest stage.

[23] https://assets.plantum.nl/p/229377/CGO%20publicatie%202018.pdf
[24] Starch potatoes are specifically grown for the production of potato starch. They are not used for human consumption.

3178 **Table 30.** Summary results of field studies conducted on total glycoalkaloid (TGA) concentrations in potatoes.

Country	Year of survey	Cultivars/location/ year/farming system	N	TGA average (mg/kg fw)	Range (mg/kg fw)	Ratio α-chaconine: α-solanine	Analytical technique	Reference
CZ	1993–1994	11 cultivars, 3 locations, 2 years	66	74	31–166	n.r.	HPLC-UV	Panovska et al. (1997)
	1996–1999	8 cultivars, 2 locations, 4 years, 2 systems	82	70	15–245	n.r.	HPLC-UV	Hajšlová et al. (2005)
	2006–2008	5 cultivars, 2 locations, 3 years, 2 systems	120	87	49–134	1.26–2.52	LC-MS/MS	Bártová et al. (2013)
	2009–2011	14 cultivars, 2 locations, 3 years	82	54	19–102	n.r.	LC-MS/MS	Hamouz et al. (2014)
	2012–2014	16 cultivars, 2 locations, 3 years	96	81	34–168	1.18–3.78	LC-MS/MS	Urban et al. (2018)
DE	1970–1974	10 cultivars, 5 locations, 5 years	184	29	3–92	n.d.	Colorimetric	Ross et al. (1978)
HU	2002–2004	10 cultivars, 1–3 years	20	33	1–150	n.r.	HPLC-UV	Tömösközi-Farkas et al. (2006)
	2007–2009	3 cultivars, 3 years, 2 systems	18	43	<1–281	n.r.	HPLC-UV	Tömösközi-Farkas et al. (2014)
IT	<2006	9 cultivars, 1 year [(a)]	9	32	10–51	2.91–5.93	HPLC-UV	Finotti et al. (2006)
	<2010	9 cultivars, 2 locations, 1 year	18	95	50–209	n.r.	LC-MS	Ieri et al. (2010)
LV	2010–2011	20 cultivars, 1 location, 1 year, 2 systems	40	21	3–104	0.30–4.60	LC-MS	Skrabule et al. (2013)
NL	1985–1986	3 cultivars, 2 locations, 3 years	60	81	40–146	n.r.	HPLC-UV	Van Gelder et al. (1988)
	Ongoing	All cultivars [(b)]	464	48	3–152	n.r.	HPLC-UV	Website NAK
PT	2003	2 cultivars, 1 location, 1 year, 3 systems	6	51	37–80	0.81–1.82	HPLC-UV	Abreu et al. (2006)
SE	1986–1987	6 cultivars, 3 locations, 2 years	12	120	51–210	n.r.	HPLC-UV	Hellenäs et al. (1995a)
	2009	21 cultivars, 1 location, 1 year	39	127	46–293	1.09–2.67	HPLC-UV	Petterson et al. (2013)
UK	1980–1982	36 cultivars, 7 locations, 3 years [(c)]	141	71	36–175	n.d.	Colorimetric	Bintclife et al. (1982); Parnell et al. (1984)
	1994–1995	2 cultivars, 1 location, 2 years	4	87	40–163	2.32-3.08	HPLC-UV	Papathanasiou et al. (1999)
USA	1970–1971	5 cultivars, 39 locations, 2 years [(d)]	255	79	14–387	n.d.	Colorimetric	Sinden and Webb (1972, 1974)
	<2003	8 cultivars, 1 location, 1 year	8	52	7–187	1.4–2.2	HPLC-UV	Friedman et al. (2003b)

3179 TGA: total glycoalkaloid. UV: ultraviolet. LC: liquid chromatrography. MS: mass spectrometry. MS/MS: tandem MS. n.r. = not reported; n.d. = not determined.
3180 (a): From wholesale.
3181 (b): For details see **Figure 6**.
3182 (c): The publication by Bintclife et al. (1982) reports the results for 1980–1981.
3183 (d): The variety B5146-6 was excluded.

The NAK list of registered potato cultivars in 2018 [25] contained 552 entries, and for 464 cultivars the TGA content is listed. Before 2000 it was not mandatory to establish the TGA content and in particularly for older cultivars this information has not been recorded. The TGA distribution in the cultivars is shown in **Figure 6**, where it can be seen that the large majority of potatoes grown under the Dutch test conditions have a TGA content of <100 mg/kg. The average content of the registered cultivars is 48.4 mg/kg.

The NAK list [25] includes also 77 potato cultivars grown specifically for starch production. For 70 cultivars the TGA content is listed. Starch potatoes [26] contain higher amounts of GAs: the average TGA content is 107.7 mg/kg, while 34 cultivars exceed 100 mg/kg and 7 exceed 200 mg/kg. The cultivar 'Kuras' has the highest TGA content (338 mg/kg) and also the cultivars 'Aventra' and 'Elles' (both 260 mg/kg) are also high in TGA content.

Overall, it can be concluded from the field studies that genotype is the most important factor determining the TGA content. However, also environmental conditions, location and year-to-year variability contribute to the differences in TGA content observed during the pre-harvest stage.

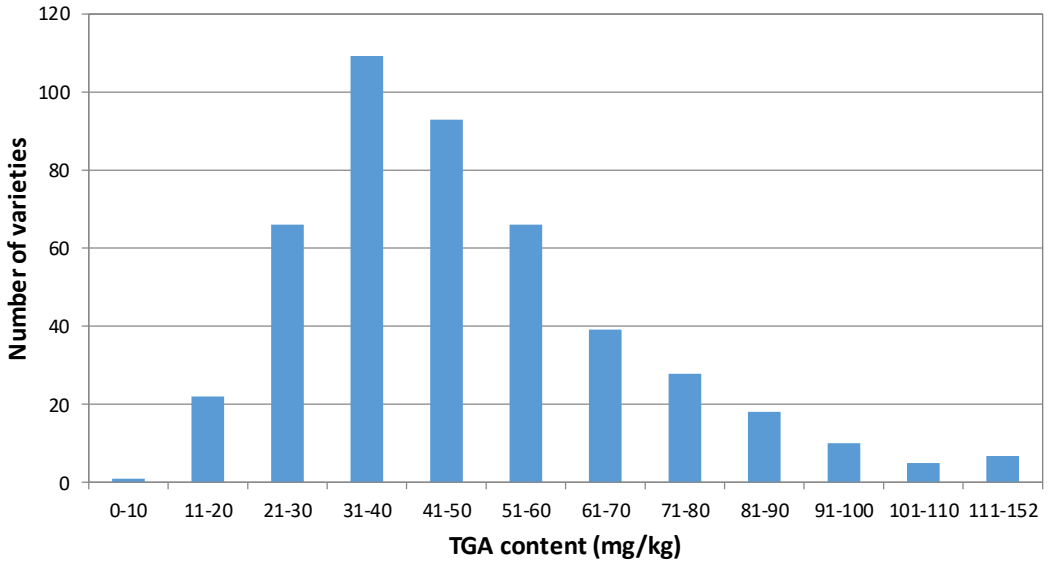


Figure 6. Distribution of total glycoalkaloid (TGA) content in 464 potato cultivars registered in the Netherlands [25].

Differences in GAs content between organic and conventionally grown potatoes

Several studies evaluated the effect of farming systems (organic versus conventional) on the GAs content in potato tubers as shown in **Table 30** and some further details are given in **Table 31**.

In the study of Abreu et al. (2006) higher TGA concentrations were found in the conventionally grown Raja tubers, compared to the organically grown ones, while for the Santé tubers no clear difference in TGA content was observed between the farming systems. Skrabule et al. (2013) evaluated GAs in 20 potato genotypes grown under organic and conventional farming systems in the years 2010 and 2011. The genotype did significantly influence the GAs content, while the effect of farming system was not

[25] https://assets.plantum.nl/p/229377/CGO%20publicatie%202018.pdf
[26] Starch potatoes are specifically grown for the production of potato starch. They are not used for human consumption.

significant, except for the content of α-solanine in the 2011 season. Bártová et al. (2013) showed the importance of the individual reaction of the genotypes to stress conditions existing in organic and conventional crop management. Crop management seemed to only influence the GA content in tubers in interaction with other evaluated agro-ecological factors (cultivar, year and locality). Although a significantly lower concentration of α-solanine was found in organically grown tubers, with respect to the TGA content the difference between the two farming systems was not significant. Hajšlová et al. (2005) studied the tuber quality of eight potato cultivars grown in organic and conventional farming systems. No statistically significant difference was found between the TGA contents in tubers produced by the two production systems. The observed difference in TGA content seems to be caused by a substantial variation in the content of GAs between years, geographical area of cultivation and cultivar. Additionally, Tömösközi-Farkas et al. (2014) investigated the GA content in three Hungarian potato species grown in organic and conventional farming systems. The respective farming system was found to have no effect on the level of GAs, and was mostly influenced by season and genotype. Diehl and Wedler (1977) studied the effect based on the rate of fertilizers applied. No difference was found between α-solanine content in potatoes in relation to organic and mineral fertilization.

Distribution of GAs between potato peel and flesh

In a number of field studies the distribution of GAs between the peel and the flesh was studied. The most comprehensive study is that of Valcárcel et al. (2014) in which the α-solanine and α-chaconine content of potato skin and flesh of 60 potato cultivars grown in two locations in Ireland over a 2-year period was investigated. TGA levels varied from 4 to 957 mg/kg dw in the potato flesh and from 150 to 8,133 mg/kg dw in the potato peels. The ratio α-chaconine to α-solanine was higher in the peels (average ratio: 1.44; range 0.7-4.5) than in the flesh (average ratio: 0.87; range 0.3-1.6). Deußer et al. (2012) conducted a similar study on 17 cultivars grown in Luxembourg. TGA contents were determined for the peel (1–2 mm thickness), the outer flesh (1 cm) and inner flesh. The peels contained between 545 and 5,342 mg TGA/kg dw, while the outer and inner flesh contained 19–466 and 7–187 mg TGA/kg dw, respectively. The α-chaconine:α-solanine ratio varied between 1.05 and 3.45 (average: 1.94) in the peel, between 0.83 and 2.36 (average: 1.46) in the outer flesh and between 1.00 and 2.38 (average: 1.69) in the inner flesh. Friedman et al. (2003) reported for 8 potato cultivars a range of 84 to 3,526 mg TGA/kg dw in potato peels, and of 6 to 592 mg TGA/kg dw in potato flesh. A study on 12 potato cultivars grown in Mexico reported for potato peels 176–5,497 mg TGA/kg dw and for flesh <5–642 mg TGA/kg dw (Sotelo and Serrano, 2000).

Table 31. Total glycoalkaloids (TGAs) content in potato tubers grown in conventional and organic farming systems.

Country	Potato cultivars	Year of harvest	TGA content conventional (mg/kg fw)			TGA content organic (mg/kg fw)			Analytical technique	Reference
			α-solanine	α-chaconine	TGA	α-solanine	α-chaconine	TGA		
CZ	Karin	1995–1997			53–113			59–157	HPLC-UV	Hamouz et al. (2005)
CZ	Christa, Karin, Koruna, Krasa, Krystala, Monalisa, Rosara, Rosella	1996–1999			15–245			16–157	HPLC-UV	Hajšlová et al. (2005)
CZ	Bionta, Karin, Marabel, Rosara, Satina	2006–2008	35.3	55.6	90.9	30.4	53.2	83.6	LC-MS/MS	Bártová et al. (2013)
HU	Hópehely, Rioja, White Lady	2007–2009			0.0–15			0.0–28	HLPC-UV	Tömösközi-Farkas et al. (2014)
LV	20 potato genotypes	2010–2011	1.0–54.5	1.5–42.5		1.5–59.0	1.5–48.5		LC-MS	Skrabule et al. (2013)
PT	Raja, Santé	2003	17.7–43.9	19.6–35.5	37.3–79.5	13.6–23.3	21.3–24.8	38.4–44.6	HPLC-UV	Abreu et al. (2006)

TGA: total glycoalkaloid. UV: ultraviolet. LC: liquid chromatrography. MS: mass spectrometry. MS/MS: tandem MS.

Occurrence data on GAs in potato samples obtained from retail surveys

In **Table 32** an overview is given of surveys conducted on GAs levels in potatoes collected from retail outlets. Information on such surveys is available only for three different countries in the EU: Denmark, Sweden and the UK.

Panovská et al. (1997) studied the TGA content in 14 Czech cultivars available on the Czech market. It was found that for all cultivars, larger sized tubers (100–160 g) consistently contained lower GA levels than smaller sized tubers (20–50 g). TGA levels in all large sized tubers were less than 100 mg/kg, while for the small sized tubers for two cultivars ('Javor' and 'Oreb') levels above 200 mg/kg were found.

The Danish survey spanned seven years, from 2001 to 2007, and a total of 386 potato tuber samples were analysed (Knuthsen et al., 2009). The number of samples and the potato cultivars collected varied from year to year. There was a significant yearly variation noted in the average TGA content of the potatoes sampled; a relatively high average TGA content was obtained for the seasons 2001–2002 and 2004–2005. In total, 56 samples exceeded 100 mg TGA/kg fw, and three exceeded 200 mg/kg fw. Cultivars that exceeded 200 mg/kg included 'Sava' (1 sample out of 38 analysed, origin Denmark), an unknown cultivar (origin: Cyprus) and 'Grenailles' (origin: Italy).

When excessive levels of GAs were detected in Swedish potatoes, particularly in the cultivar 'Magnum Bonum', a broad survey was conducted in Sweden. The survey was conducted in 1986–1987 and a total of 617 potato samples were analysed, including 348 'Magnum Bonum' samples (Hellenäs et al., 1995a,b). Besides the 'Magnum Bonum' cultivar, high TGA levels were also found in potatoes of other cultivars, but to a lower extent. The most prominent of the other cultivars that regularly exceeded 200 mg/kg were the early potato 'Ulster Chieftain' and the main crop potato 'Snöboll (British Queen)'. In a follow up study in 1997, 240 samples of main crop and early potato cultivars were sampled and analysed (Branzell and Hellenäs, 1999). TGA levels were only slightly lower than in 1986–1987. The 6 samples collected from the early potato 'Ulster Chieftain' all exceeded 200 mg/kg and also about 30% of the early potato 'Pentland Javelin' samples also exceeded this limit. A further survey was conducted by the Swedish Food Safety Authority in 2015, in which 38 samples of potato tubers were collected and analysed for GA content (Swedish NFA, 2016). The 38 samples represented 31 different cultivars. Twelve samples had a TGA content in excess of 100 mg/kg, and five exceeded 200 mg/kg: the cultivars 'Caro', 'Folva', 'Rosagold', 'Sava' (for each cultivar one sample analysed) and 'Snöboll' (one out of two samples analysed). Overall, the TGA levels were slightly lower than in 1986–1987 and similar to the 1997 surveys.

In the UK study of 1981, 514 potato samples were collected from supermarkets, greengrocers and farm shops (Davies and Blincow, 1984). Samples were pooled to produce 133 composite samples. Sampling of cultivars was made on basis of available consumption data. Samples were grouped in three categories: foreign early potatoes (42 samples), UK early potatoes (28 samples) and UK main crop potatoes (63 samples). The TGA content was determined using a less specific colourimetric method. The average TGA content of the samples was 113 mg/kg fw, with a range of 56–212 mg/kg. No statistically significant differences in TGA content between the different groups could be found. One sample (cultivar 'Pentland Javelin', six samples tested) exceeded 200 mg TGA/kg.

Processed potato food products

Several studies have determined the levels of GAs in laboratory processed and in commercial potato products such as potato crisps (or potato chips) and French fries (Sizer et al., 1980; Jones and Fenwick, 1981; Bushway and Ponnampalam, 1981; Friedman and Dao, 1992; Rytel, 2012a; Liu et al., 2014; Tajner-Czopec et al., 2014). The results are summarised in **Table 33**. In potato crisps TGA levels

ranging from 16 to 720 mg/kg have been reported. In French fries the reported levels are lower, ranging from 0.4 to 58 mg/kg. Dehydrated products contained TGA levels between 15 and 75 mg/kg.

Potato peel products

Studies that investigated the GA content of potato peel products are summarised in **Table 34**. Bushway et al. (1983) analysed a range of different potato products available on the US market and reported a range from 52–242 mg TGA /kg. They also studied the TGA content of baked, fried and baked-fried potato peels (see **Section 3.2.4.1.2**). An increase in TGA content was noted compared to unprocessed peels, what could be explained by a lower water content in the processed products.

Friedman et al. (2017) measured the GAs content of potato peel powder prepared from three conventional and three organic store-bought potatoes ('Gold', 'Red' and 'Russet') by HPLC and LC-MS and detected a wide variation in GAs content between the samples. Highest TGA levels were found in potato peel powder from the organically grown 'Gold' cultivar (3,580±390 mg/kg dw) and the lowest in conventionally grown 'Russet' (639±52 mg/kg dw). It was noted that the corresponding cultivars did not always have a higher TGA level when comparing the organic versus the conventional samples. For the 'Red' cultivar the TGA level in potato peel powder of conventionally grown potatoes was higher than in organically grown potatoes (1,709±61 vs 850±120 mg/kg dw). In a follow up study Elkahoui et al. (2018) reported the GAs content in potato peel powder prepared from three potato cultivars ('Gold', 'Red', 'Russet') grown under organic and/or conventional conditions. A higher TGA content was reported for the conventionally grown 'Russet' cultivar (1,128±1 mg/kg dw) vs the organically grown 'Russet' cultivar (861±10), but higher TGA levels were reported for the other two cultivars grown only under organic conditions (1,940–2,180 mg/kg dw).

3309 **Table 32.** Summary results of surveys conducted on total glycoalkaloids (TGAs) levels in consumption potatoes sampled in retail outlets.

Country	Year of survey	Potato cultivars	N	TGA average (mg/kg fw) [(b)]	Range (mg/kg fw)	Analytical technique	Reference
CZ	1993–1994	All cultivars (14) (tuber size 100–160 g) [(a)]	14	63	23–95	HPLC-UV	Panovská et al. (1999)
		All cultivars (14) (tuber size 20–50 g) [(a)]	14	110	40–237		
DK	2001–2002	All cultivars (6)	28	102	45–167	HPLC-UV	Knutsen et al. (2009)
	2002–2003	All cultivars (14)	50	64	26–159		
	2003–2004	All cultivars (10)	54	97	26–280		
	2004–2005	All cultivars (28)	99	67	15–223		
	2005–2006	All cultivars (20)	64	54	5–109		
	2006–2007	All cultivars (22)	91	49	<5–105		
		All DK samples	**386**	**67**	**<5–280**		
SE	1986	'Magnum Bonum'	332	254	61–665	HPLC-UV	Hellenäs et al. (1995a,b)
		Others (8 main crop cultivars)	105	108	35–315		
	1987	'Magnum Bonum'	16	151	60–293	HPLC-UV	Hellenäs et al. (1995a,b)
		Others (8 early cultivars)	126	120	31–344		
	1997	Main crop potatoes (8 cultivars)	113	105	36-244	HPLC-UV	Branzell and Hellenäs (1999)
		Early potatoes (7 cultivars)	127	98	35-267		
	2015	All cultivars (31)	38	95	24–277	HPLC-UV	Swedish NFA (2016)
		All SE samples but excluding 'Magnum Bonum'	**509**	**107**	**24–344**		
UK	1980-1981	Foreign early potatoes (8 cultivars)	42	123	56–208	Colorimetric	Davies and Blincow (1984)
		UK early potatoes (5 cultivars)	28	115	70–212		
		UK main crop potatoes (3 cultivars)	63	105	60–195		
		All UK samples	**133**	**113**	**56–212**		

3310 LC: liquid chromatography. UV: ultraviolet.
3311 (a): From wholesale.
3312 (b): Average as reported by the authors or calculated based on the provided individual data.

Table 33. Totalglycoalkaloids (TGAs) in laboratory processed and in commercial potato products.

Country	Year of study	Product [a]	N	TGA average (mg/kg)[f]	Range (mg/kg)	Analytical technique	Reference
USA	1980	Potato chips [b]	3	304	95–720	Colorimetric	Sizer et al. (1980)
		Potato chips [c]	4 (different production methods [d]) Potato TGA content: 42.3 mg/kg	166	123–236		
UK	1981	Potato crisps [b]	2	65	59–70	Colorimetric	Jones and Fenwick (1981)
		Pre-cooked frozen chips [b]	2	39	23–55		
USA	1981	Potato chips [b]	14	71	27–162	HPLC-UV	Bushway and Ponnampalam (1981)
		Frozen fries (steak, French, crinkle cut) [b]	18	22	3.3–58		
		Frozen fried potato balls [b]	4	19	14–23		
		Frozen mashed potatoes [b]	3	3.3	2.0–4.6		
		Frozen baked potatoes [b]	2	102	80–123		
		Frozen fried potatoes [b]	5	6.4	4.3–8.4		
		Dehydrated potato flour [b]	2	70	65–75		
		Dehydrated potato flakes [b]	2	19	15–23		
		Canned potatoes [b]	4	1.5	0.9–2.5		
		Canned home fries [b]	2	1.3	1.1–1.5		
USA	1992	Potato chips [b]	3	61	23.5–109	HPLC-UV	Friedman and Dao (1992)
		French fries [b]	3	18	0.4–44		
		Pancake Powder [b]	2	45	44–45		
PL	2012	Dehydrated cooked potatoes [c]	4, potato TGA content: 200–296 mg/kg dw; 5 processing stages	65	61–69	HPLC-UV	Rytel (2012a)
CN	2014	Potato crisps [b]	20	n.d.	16–61	LC-MS/MS	Liu et al. (2014)
PL	2014	French fries [c]	4 (colour-fleshed potatoes) Potato TGA content: 41–61 mg/kg; 5 processing steps [e]	8	6–10	HPLC-UV	Tajner-Czopec et al. (2014)

dw: dry weight. LC: liquid chromatography. UV: ultraviolet. MS: mass spectrometry. MS/MS: tandem MS.

(a): Potato chips (often just chips), or crisps, are thin slices of potato that have been deep fried or baked until crunchy. Regarding naming, British crisps are US chips, while British chips are US fries/French fries (Oxford dictionary online). In the above table each product is named according to the naming in the article.
(b): Commercial product.
(c): Experimental product.
(d): Different frying temperature, frying time and slice thickness.
(e): For details see **Section 3.2.4**.
(f): Average as reported by the authors or calculated based on the provided individual data.

Table 34. Total glycoalkaloids (TGAs) in laboratory processed and in commercial potato peel products.

Country	Year of study	Potato product	N	TGA average (mg/kg fw) [c]	TGA range (mg/kg fw)	Analytical technique	Reference
USA	1981	Fried peels [a]	2	142	139–145	HPLC-UV	Bushway and Ponnampalam (1981)
USA	1983	Unprocessed potato peels [b]	12 cultivars	276	18–1,068	HPLC-UV	Bushway et al. (1983)
		Baked potato peels [b]	12 cultivars	317	24–1,098		
		Fried potato peels [b]	12 cultivars	414	33–1,649		
		Baked-fried potato peels [b]	12 cultivars	444	42–1,556		
		Frozen peel wedges [a]	6	98	76–121		
		Frozen peel slices [a]	2	68	66–71		
		Restaurant fried peels [a]	3	57	52–63		
		Restaurant baked-fried peels [a]	3	181	120–242		
USA	2017	Potato peel powder [b]	6 (3 cultivars, organic and conventional)	1,541 (dw)	639–3,580 (dw)	HPLC-UV	Friedman et al. (2017)
USA	2018	Potato peel powder [a]	4 (3 cultivars, organic and conventional)	1,527 (dw)	861–2,180 (dw)	HPLC-UV	Elkahoui et al. (2018)

Dw: dry weight. LC: liquid chromatography. UV: ultraviolet.
(a): Commercial product.
(b): Experimental product.
(c): Average as reported by the authors or calculated based on the provided individual data.

3.2.2.1.2. Occurrence data on GAs in tomatoes

A number of publications were identified on the occurrence of tomato GAs in tomatoes. The results are summarised in **Table 35**. The levels of α-tomatine reported in ripe tomatoes are generally low, ranging from not detectable to 23 mg/kg fw. In green, but mature, tomatoes somewhat higher amounts of α-tomatine were found, ranging from not detectable to 90 mg/kg fw. Green, immature, tomatoes contain the highest TGA levels, ranging from 35 to 550 mg/kg fw. It should be noted that the results were obtained with methods that had a wide range of LOQs (ranging from 0.05 to 2.5 mg/kg).

Baldina et al. (2016) investigated 15 landraces and 3 hybrids with different morphological characteristics (flattened/ribbed, pear/oxheart, round/elongate) for the presence of 7 tomato glycosides (α-tomatine, α-dehydrotomatine, esculeoside A, lycoperosides A, B and H and tomatoside A). On average slightly higher concentrations were found for tomatoside A (mean 14.4 mg/kg fw, range 0.25–5.5 mg/kg fw) than for α-tomatine (mean 12.2 mg/kg fw, range 0.36–3.4 mg/kg fw), esculeoside A (0.91 mg/kg fw, range 0.36–2.7 mg/kg fw) and lycoperoside H (0.88 mg/kg fw, range 0.28–2.8 mg/kg fw). The other GAs were present at lower levels.

Differences in GAs content between organic and conventionally grown tomatoes

Koh et al. (2013) studied the α-tomatine content in dried tomato samples grown by organic and conventional crop management over a period of 10 years. Mean levels of α-tomatine in organically grown tomatoes (42.3 mg/kg dw) were significantly higher than in the conventionally (23.2 mg/kg dw) grown crops. The differences were attributed to environmental factors, external to nitrogen fertilization, affecting the α-tomatine content in tomatoes.

Processed tomato products

Most of the studies on GA content in processed tomato products have been made by the same research group (Friedman et al., 1994; Friedman and Levin, 1995; Friedman, 2004). **Table 36** shows the results of a survey in which different tomato products were obtained from a local market. Pickled green tomatoes and tomato red sauce showed the highest concentrations of α-tomatine (Kyzlink et al., 1981; Kibler et al., 1985; Friedman and Levin, 1995).

3.2.2.1.3. Occurrence data on GAs in aubergines

The literature on the occurrence of GAs in the aubergine (*S. melongena*) is very limited. The data available are summarised in **Table 38**. Bajaj et al. (1979), using a colorimetric method, reported on the TGA content in mature fruits of 21 different cultivars (including round type, long type and oblong type fruits). The fruits contained GAs, expressed as α-solanine equivalents, between 65 and 205 mg/kg fw (Bajaj et al., 1979). Using a similar approach, Jones and Fenwick (1981) analysed two aubergine samples imported to the UK. The samples contained 76 and 82 mg GAs (determined as α-solanine) per kg fw.

Using a HPLC-UV method, Eanes et al. (2008) reported for *S. linneaeanum* (a wild type aubergine) a solmargine content of 25,500 ± 11,200 mg/kg dw, while no GAs could be detected in a commercial sample of aubergine (reported LOQ was 100–200 mg/kg dw). Assuming a water content of 90%, the LOQ in fresh material was 10–20 mg/kg.

Using an LC-MS/MS method, Sánchez-Mata et al. (2010) reported for the fruits of four different varieties of the gboma aubergine a TGA content of 1,402–2,210 mg fw. The majority was made up by α-solamargine (1,241–1,975 mg/kg), while the concentration of α-solasonine (161–235 mg/kg) was lower.

In three varieties of the scarlet aubergine, α-solamargine (5.8–48.6 mg/kg) was also the major GA compared to α-solasonine (4.1–10.0 mg/kg). Also, for three varieties of aubergine, α-solamargine (8.5–16.1 mg/kg) was present in higher concentrations than α-solasonine (1.7–4.0 mg/kg) (Sánchez-Mata et al., 2010).

Mennella et al. (2012) studied the effect of ripening on the GA concentration in 10 experimental lines of aubergine. Interestingly, they noticed a substantial increase of GA concentration during ripening of the fruits. The TGA concentration increased from approximately 10 mg/kg fw in unripe aubergines to 20 mg/kg fw in commercially ripe aubergines to 160 mg/kg fw in physiologically ripe aubergines. In the immature and commercially ripe stage, the flesh appears soft and greenish, the peel glossy (unripe) or a bit dull (commercial stage). The physiologically ripe stage is typified by a brownish peel colour, spongy, fibrous and yellowish flesh and visible and hard mature seeds. The ratio between α-solamargine and α-solasonine fluctuated substantially between the different samples. In approximately 50% of the samples α-solasonine could not be detected (LOQ was 0.02 mg/kg fw). Overall, α-solamargine contributed to the total content, for the unripe, commercially ripe and physiologically ripe stage, 77%, 83% and 69%, respectively (Mennella et al., 2012).

3.2.2.1.3. Occurrence data on GAs in other food products

The Rapid Alert System for Food and Feed (RASFF) has received, up to the endorsement of this Opinion, no notifications on the presence of high levels of GAs in potato, aubergine, tomato or other food products. However, within the period 1982–2019, there have been seven notifications on the presence of *S. nigrum* (black nightshade) in food products (see **Appendix D**). Three recent notifications (2015–2018) concern the presence of *S. nigrum* in food supplements originating from India. The four other notifications (1982–2013) report the presence of *S. nigrum* berries in green beans (canned or frozen). None of the notifications report on concentrations of GAs in the products. The potential of *S. nigrum* and the related species *S. ptycanthum* Dun (eastern black nightshade) to contaminate food crops, such as beans, peas and soybeans, is a problem noted by several authors (Eldridge et al., 1983; Ogg et al., 1981; Ogg and Rogers, 1989). Cavlovic et al. (2013) reported unripe berries of *S. nigrum* to contain 1,100 mg/kg fw α-solasonine and 900 mg/kg fw α-solamargine. In a follow up of the RASFF notification 2013.0804, six samples of *S. nigrum* green berries were analysed by LC-MS/MS (P. Mulder, 2019, personal communication). The berries contained 1,540 ± 170 mg/kg fw α-solasonine and 1,580 ± 260 mg/kg fw α-solamargine. Other GAs (α-solanine and α-chaconine) as well as the aglycones solasodine and solanidine could not be detected (LOD = 1 mg/kg fw).

3.2.2.1. Literature occurrence data in feed

No surveys on the levels of GAs in potatoes and potato by-products used as feeds for livestock have been identified in the open literature.

3402 **Table 35.** Summary results of literature reports on total glycoalkaloids (TGA) concentrations in tomatoes.

Country	Year of survey	Tomato	N	TGA average (mg/kg fw) [e]	Range (mg/kg fw)	Analytical technique	Reference
UK	1984	Ripe (3 cultivars) [a]	3	3.7	3–6	Radioimmunoassay	Eltayeb and Roddick (1984)
NL	1987	Ripe [a]	1	5	-	GC-FID and GC-NPD	Van Gelder and De Ponti (1987)
NL	1994	Red [b]	14	0.6	<0.5–4.1	HPLC-UV	Keukens et al. (1994)
		Orange [b]	14	6.2	<0.5–18.2		
		Green [b]	14	26	4.5–90		
USA	1994	Red (5 cultivars) [b]	99	-	<2.5–23 (94 are ND)	HPLC-UV	Bushway et al. (1994)
		Green (4 cultivars) [b]	80	18.3	<2.5–88		
USA	1994–2004	Red, conventional (3 cultivars) [c]	54	1.16 [f]	<0.05–5.6 [f]	LC-MS/MS	Koh et al. (2013)
		Red, organic (3 cultivars) [c]	54	2.11 [f]	<0.05–4.9 [f]		
USA	1994	Red, ripe [d]	4	1.1	0.3–2.8	HPLC-UV/ HPLC-PAD	Friedman et al. (1994)
		Red [a]	2	2.7	1.5–3.9		
		Breaker [a]	1	16.5	-		
		Green, mature [a]	1	45	-		
		Green, immature [a]	1	69	-		
USA	1995	Red, yellow, ripe (8 cultivars) [d]	8	3.6	0.3–11	HPLC-PAD	Friedman and Levin (1995)
		Green, unripe [d]	1	16	-		
		Red [a]	2	1.8	1.2–2.5		
		Breaker [a]	2	10.5	6.7–14		
		Green, mature [a]	2	40	25–58		
		Green, immature [a]	2	125	40–210		
JP	1997	Green, mature [a]	7	165	140–180	Colourimetric	Furui et al. (1997)
		Turning [a]	4	44	30–60		
		Red, mature [a]	6	3	1-8		
USA	1998	Red [a]	15	1.4	0.4–4.3	HPLC-PAD	Friedman and Levin (1998)
		Green, immature [a]	15	205	35–550		

IT	2000	Green-orange, red [a]	5	15	ND–43	HPLC-UV	Leonardi et al. (2000)
IL	2006	Ripening [c]	3	2.5 [f]	0–6.7 [f]	HPLC-UV	Mulatu et al. (2006)
		Green, mature [c]	3	0.65 [f]	0–1.4 [f]		
		Green, immature [c]	3	1.4 [f]	0.13–3.5 [f]		
JP							
IT	2016	Ripe (15 landraces and 3 hybrids) [c]	18	1.5 [g] 5.1 [h]	0.6–4.0 [g] 1.8–14.2 [h]	LC-HRMS	Baldina et al. (2016)
IT	2019	Various ripening stages [d]	3	1.1 [g]	0.49–2.0 [g]	LC-MS/MS	Tamasi et al. (2019)

TGA: total glycoalkaloids. LC: liquid chromatography. UV: ultraviolet. MS: mass spectrometry. MS/MS: tandem MS. HR: high resolution. HP: high performance. PAD: Pulsed amperometric detector. FID: flame ionisation detector. NPD: nitrogen-phosphorous detector.
(a): Greenhouse.
(b): Wholesale.
(c): Field trial.
(d): Retail.
(e): Average as reported by the authors or calculated based on the provided individual data.
(f): Data were presented as dry weight. It has been assumed that the water content was 95%.
(g): Results for α-tomatine + α-dehydrotomatine.
(h): Results for all 7 GAs analysed (α-tomatine, dehydrotomatine, esculeoside A, lycoperosides A, B and H and tomatoside A).

3413 **Table 36**. Total glycoalkaloid (TGA) content of processed tomato products reported in the literature.

Country	Year of study	Product	N	TGA average (mg/kg fw) [f]	TGA range (mg/kg fw)	Analytical technique	Reference
CZ	1981	Tomato pulp [a] [b]	1	193		HLPC/colourimetric [i]	Kyzlink et al. (1981)
		Sweetened tomato juice [a] [b]	1	165			
		Tomato marmalade [a] [b]	1	182			
		Fruit butter [a] [b]	1	143			
DE	1985	Green tomato marmalade [a]	2	35 [g]	25-44 [g]	Colourimetric [i]	Kibler et al. (1985)
		Sweet sour pickled green tomatoes [a]	2	31 [g]	20-42 [g]		
CZ	1992	Green tomato salad with onion, sterilized [a] [c]	2	24	20–28	HPLC-UV	Voldrich et al. (1992)
		Green tomato salad with onion and paprika [a]	1	8			
		Fermented green tomatoes [a]	1	14 [h]			
		Tomato puree [a]	3	18	16–20		
USA	1995	Condensed tomato soup	3	2.2	1.4–3.7	HPLC-PAD	Friedman and Levin (1995)
		Stewed red tomatoes	1	1.1			
		Sun dried tomatoes	1	21			
		Fried green tomatoes [a] [d]	1	11			
		Microwaved green tomatoes [a] [e]	1	12			
		Tomato ketchup	1	25			
		Tomato juice	1	28			
		Tomato red sauce	1	57			
		Green salsa	2	9.5	0–19		
		Pickled green tomato product	2	50	28–72		
USA	2017	Red and tangerine tomato juice	2	0.65	0.5-0.8	LC-HRMS	Cichon et al., 2017

3414 TGA: total glycoalkaloids. LC: liquid chromatography. UV: ultraviolet. MS: mass spectrometry. HR: high resolution. HP: high performance. PAD: Pulsed amperometric detector.
3415 (a): Prepared in the laboratory.
3416 (b): Prepared from unripe tomatoes.
3417 (c): 1 sample sterilized at 90°C and 1 sample sterilized at 120°C.
3418 (d): Frying was done as follows: tomatoes were sliced and breaded with equal amounts of flour and corn meal after which they were fried in ¼ inch of oil.
3419 (e): Microwaving was for five minutes at 'high'.
3420 (f): Average as reported by the authors or calculated based on the provided individual data.
3421 (g): Reported as solanine, probably α-dehydrotomatine has been measured.
3422 (h): The brine was analysed as well, containing 7 mg/kg α-tomatine.
3423 (i): Colourimetric methods cannot discriminate between α-solanine, α-chaconine and α-dehydrotomatine.

3424 **Table 37**. Summary results of literature reports on total glycoalkloid (TGA) concentrations in aubergine.

Country	Year of survey	Aubergine species	N	TGA average (mg/kg fw) [c]	Range (mg/kg fw)	Analytical technique	Reference
IN	1979	*S. melongena* [a]	21	113	65–205	Colorimetry	Bajaj et al. (1979)
UK	1981	*S. melongena* [b]	2	79	76–82	Colorimetry	Jones and Fenwick (1981)
TR	2006	*S. melongena* [b]	1	<LOQ (10)	-	HPLC-UV	Eanes et al. (2006)
ES	2010	*S. melongena* [a]	3	14.3	11–20	LC-MS/MS	Sánchez-Mata et al. (2010)
		S. macrocarpon [a]	4	1639	1,402–2,210		
		S. aethiopicum [a]	3	19.5	10–54		
IT	2012	*S. melongena*, unripe [a]	10	10.2 [d]	0.6–19.6 [d]	HPLC-UV	Mennella et al. (2012)
		S. melongena, commercial state [a]	10	20.6 [d]	<0.02–85 [d]		
		S. melongena, physiologically ripe [a]	10	163 [d]	25.0–403 [d]		
		S. aethiopicum, unripe [a]	1	527	-		
		S. aethiopicum, commercial stage [a]	1	193	-		
		S. aethiopicum, physiologically ripe [a]	1	349	-		
IT	2016	*S. melongena*, commercial state [a]	3	311 [d]	231–430 [d]	HPLC-UV	Lo Scalzo et al. (2016)
IT	2019	*S. melongena*, commercial state [a]	3	435 [d]	-	LC-HRMS	Lelario et al. (2019)

3425 TGA: total glycoalkaloids. LC: liquid chromatography. UV: ultraviolet. MS: mass spectrometry. MS/MS: tandem MS. HR: high resolution. HP: high performance.
3426 (a): Field trial.
3427 (b): Retail.
3428 (c): Average as reported by the authors or calculated based on the provided individual data.
3429 (d): Concentrations were presented as dry weight; fresh weight concentrations have been calculated based on the water content of the samples as provided in the publication.

3.2.3. Influence of pre-harvest factors on the content of GAs

Factors that influence the content of GAs pre-harvest have been briefly described in **Section 1.3.3** and **3.2.2.1.1**.

3.2.4. Influence of storage and processing on the content of GAs

Post-harvest, both storage and processing may influence the content of different GAs in edible parts of plants such as in the potato tuber and in the tomato fruit.

3.2.4.1. GAs from *S. tuberosum*

3.2.4.1.1. Storage of potatoes

The main factors that may influence the GA content during storage of potato tubers include the duration, temperature, humidity and light exposure (wavelength and intensity), as well as tuber damage and the matrix on/in which the tubers are stored. Several studies on the influence of such factors on the content of GAs in potatoes have been performed. In these studies, the potato tubers were usually exposed to a combination of two or more of these factors, and different potato cultivars have been tested in the different experiments.

In general, studies have reported increases in the GA content with storage time (e.g. Abdel-Gawad et al., 1993; Amer et al., 2014; Abassi et al., 2016; Romanucci et al., 2016), although no variations in the levels have been observed for certain cultivars (e.g. Amer et al., 2014; Edwards and Cobb, 1999), while Wilson et al. (1983) reported a decrease. Storage at low temperatures (4°C) has been shown to increase the levels in some cultivars compared to storage at 10°C (Cieslik and Praznik, 1998), while others showed the opposite (Kumar et al., 2009) or no change (Edwards and Cobb, 1997).

Exposure to light has generally been reported to induce formation of GAs. Both the wavelength of the light source and the cultivar influence the degree to which the overall content of GAs is enhanced (Nema et al., 2008; Paradiso et al., 2019). In general, exposure to fluorescent light has been shown to induce higher levels of GAs (e.g. Dao and Friedman, 1994; Percival, 1999; Machado et al., 2006; Abbasi et al., 2016). While some cultivars have been found to be nearly insensitive to light exposure (e.g. 'Maris Piper'), others were reported to be very sensitive (e.g. 'Kestrel') (Percival, 1999).

Previous studies observed that UV light at a wavelength about 300 nm induced GA synthesis, while it did not induce the synthesis of chlorophyll (Conner, 1937). The formation of chlorophyll in the outer 3 mm of the tuber is stimulated by irradiation with red light at higher wavelengths (between 600 and 700 nm) (Liljemark and Widoff, 1960). The light-induced chlorophyll and GA formation have been shown to be metabolically independent events in the plant tissue (Edwards et al., 1998; Friedman, 2006).

The storage temperature affected the levels of GAs in response to light exposure. Storage at 10°C prior to 6 weeks storage at 4°C has been reported to reduce the sensitivity of the tubers to subsequent light exposure (Griffiths et al., 1998).

An increase in the levels of GAs after mechanical damage or wounding of the tuber has been observed (e.g. Ahmed and Mueller, 1978; Fitzpatrick et al., 1978; Mondy et al., 1987; Dale et al., 1998; Petersson et al., 2013; Dastmalchi et al., 2019). The combination of light and wounding was also reported to increase the levels of GAs, especially around the wound surface (Petersson et al., 2013; Chen et al., 2018; Nie et al., 2019).

The influence of the matrix in which the potatoes are stored has been studied under different storage conditions, such as temperature and light, and for different potato cultivars. Potatoes packaged in low light transmission packaging (e.g. craft paper coated with black polyethylene, high density polyethylene)

have been reported to contain lower levels than those packed in high light transmission packages (e.g. polyethylene blue, transparent plastic film) (Bitsch et al., 1974; Rosenfeld et al., 1995; Abbasi et al., 2016), although other studies have reported that potatoes stored in polyethylene bags showed higher GA levels that those packaged in mesh or paper (Gosselin and Mondy, 1989).

Different methods to reduce the content of GAs in stored potatoes for later distribution and consumption have also been investigated. Submersion in different solutions such as 0.1 M of NaOH (pH 12) has been reported to reduce the α-solanine and α-chaconine content by 43% and 27%, respectively (Romanucci et al., 2018). Methods that inhibit sprouting, such as surface coverage with hydrophobic nanosilica (Zhang et al., 2018) or γ-irradiation (Bergers, 1981; Dale et al., 1997; Afify et al., 2012) have also been proposed, as well as the addition of sulfur-containing compounds, such as methionine, thiamine and garlic, which have been reported to reduce the GA content up to 60% and more (Surjawan et al., 2001; El-Said, 2013; Karaway, 2014).

Several national bodies have provided a general advice to consumers on the home-storage of potato tubers in relation to GAs. For example, the BfR[27], the Netherlands Nutrition Centre Foundation (Voedingscentrum)[28] and the Swedish National Food Agency[29] advise to store potatoes in a dark and cool place.

Other bodies have provided advice to food business operators on the storage of potatoes but with regard to the potential formation of acrylamide after further processing (UK-FSA[30], US-FDA[31], Commission Regulation (EU) 2017/2158[32]). The latter stipulates that the temperature shall be appropriate to the potato cultivar stored and it shall be above 6°C, the level of humidity shall be such as to minimise senescent sweetening, and sprouting shall be suppressed in long term stored potatoes where permitted, using appropriate agents.

3.2.4.1.2. Processing of potatoes for food consumption

Potato tubers in general are only eaten after they have undergone some processing, which may take place in private households or in the food industry. Private household processing (domestic cooking) of potatoes may include the following methods:

(1) peeling
(2) boiling, blanching or steaming in water,
(3) frying and deep-frying in hot oil ('French fries' and crisps),
(4) oven baking,
(5) microwaving,
(6) grating or cutting (e.g. followed by frying (potato pancakes) or boiling (potato dumplings)

Industrial processing of potatoes intended for food consumption includes the above-mentioned methods. In addition, other processing methods may be applied for the production of other products such as dehydrated potato flakes and potato granules, flour and starch.

[27] https://mobil.bfr.bund.de/cm/343/speisekartoffeln-sollten-niedrige-gehalte-an-glykoalkaloiden-solanin-enthalten.pdf
[28] https://www.voedingscentrum.nl/encyclopedie/bewaarwijze.aspxv
[29] https://www.livsmedelsverket.se/en/food-habits-health-and-environment/food-and-environment/take-care-of-the-food--minimize-food-waste/forvara-maten-ratt/?_t_id=1B2M2Y8AsgTpgAmY7PhCfg%3d%3d&_t_q=solanine&_t_tags=language%3aen%2csiteid%3a67f9c486-281d-4765-ba72-ba3914739e3b&_t_ip=91.200.210.5&_t_hit.id=Livs_Common_Model_PageTypes_ArticlePage/_6d780797-482a-4e65-8746-96a6430a6299_en&_t_hit.pos=1#Potatoes
[30] https://www.food.gov.uk/print/pdf/node/281
[31] https://www.fda.gov/food/chemicals/acrylamide-and-diet-food-storage-and-food-preparation
[32] Commission Regulation (EU) 2017/2158 of 20 November 2017 establishing mitigation measures and benchmark levels for the reduction of the presence of acrylamide in food. OJ L304/24.

The scientific literature identified includes studies on the influence of single processing steps mentioned above on the resulting final content of GAs in the processed potato, and also studies on the effect of a typically industrial, multistep processing to produce, e.g. frozen French fries or potato chips. In this section, a summary of these studies is provided with the focus on the effectiveness of the most important processing steps with regard to reducing the GAs content in potato products.

Tuber peeling

A number of investigations provide values for the reduction of GAs in potatoes and potato products as a result of peeling. Since the major part of the total GA content in the tuber is situated just under the cork layer, peeling is expected to result in a reduction of the GA content. The reduction depends on the depth and peeling technique used (Rytel, 2012a).

In households, peeling of cooked potatoes is normally performed using a peeling knife. In the industry, the peeling of potatoes may take place using different types of mechanical peeling machines (abrasion or knife peelers), steam peeling machines or using caustic peeling (lye peeling) where the potato is surface treated with a caustic solution prior to the peeling (Muneta et al., 1977; Dornow Food Technology, 2019).

In **Table 38** the reduction in GA content found in studies published in the open literature is shown. Overall, the reduction is between 25% and 75% and the average reduction is 48%.

Boiling, blanching or steaming in water

The effects of boiling, blanching and steaming in water on the content of GAs have been studied alone or as part of studies that included peeling and/or other methods of processing. This thermal process can further reduce the contents of GAs, and the degree of reduction may differ depending on the applied temperatures and degree of disintegration of the raw materials (Rytel et al., 2018).

In **Table 39** the results of some of these studies are shown. It should be noted that the reduction is given as the percent reduction in the GA content after peeling.

A considerable part of the GAs has been shown to be released into the boiling water in their intact form, both when the potatoes are boiled as no-peeled and when boiled as peeled. Mondy and Gosselin (1988) reported higher TGA levels in potatoes boiled with peel than without peel (peel referred to in this study as the periderm and 1–2 mm of the cortex tissue). The authors also reported an increase in TGAs in the cortex tissue if the peel was not removed before boiling, concluding that GAs migrated from the peel into the cortex tissue during cooking.

Overall, boiling and certain blanching regimes result in reductions of between 5% and 65% and the average reduction is 33%.

Frying

A decrease in the content of TGA has been shown after frying in oil, e.g. for the production of French fries or potato crisps [33], and the results from several studies are summarised in **Table 40**.

Takagi et al. (1990) studied the effect of frying temperature on the TGA content in peeled potato cubes. At 150°C there was only a minor loss (5%) of GAs, at 170°C the loss increased to 21% and at 210°C TGA levels were reduced by 38%. It was remarked by the authors that 170°C is the standard temperature for frying, while frying at 150°C or at 210°C is outside the typical range.

[33] In this opinion, the terminology 'potato crisps' refers to crunchy thin slices of deep-fried/baked potato usually eaten as snacks, whereas 'French fries' refers to batons of deep-fried potato usually served as an accompaniment during a meal. When citing literature data, the original food name is used. If it is different from the standard terminology used in the opinion, then the standard terminology is also indicated in brackets, when possible.

Rytel et al. (2005) reported a reduction of 62–79% (from 48–55 mg/kg dw to 10–21 mg/kg dw) after first stage frying for 1 min of blanched potato strips, while a reduction of 75–87% (48–55 mg/kg dw to 6–14 mg/kg dw) was obtained after two stage frying for an additional 5 min. A reduction of approx. 92% was reported by Tajner-Czopek et al. (2014) when blanched potato strips were fried at 175°C for 5–6 min. Rytel et al. (2018) reported a reduction of 84% for blanched potato strips that were fried for 5–6 min at 175°C (from 43.9–60.4 mg/kg dw to 7.0–9.5 mg/kg dw).

Pęksa et al. (2006) reported that during frying of blanched potato slices at 170–180°C to produce chips, the TGA levels were reduced by 39% from 82.1–85.3 mg/kg dw to 22.6–23.5 mg/kg dw). Chips produced from peeled raw potato slices that were fried for 2 min at 190°C had a 45–54% reduced TGA level (from 62.7–97.4 mg/kg dw to 28.3–53.0 mg/kg dw) (Rytel et al., 2018).

In some of these studies, the ratio α-solanine:α-chaconine was studied and it remained constant regardless of the different conditions (e.g. temperature and time) employed in the studies (Takagi et al., 1990; Pęksa et al., 2006; Rytel et al., 2018).

In conclusion, a reduction between 20% and 90% in the final fried potato product compared to the peeled potato is reported.

Other processing methods

The effects of baking and microwaving on the content GAs have also been studied. In **Table 41** the results of some of these studies are shown. It should be noted that the reduction is given as the percent reduction compared to the original GA content in the unpeeled potato.

Treatment of non-peeled tubers, cut into cubes, at 750 W for 10 min in a microwave oven resulted in an average reduction of the TGA content of 45% (from 267 to 146 mg/kg dw) (Lachman et al., 2013). Lower reductions (15%) were reported by Takagi et al. (1990), while Mulinacci et al. (2008) did not observe significant variations in the content of GAs in microwaved unpeeled potatoes compared to fresh potatoes. In a comparative study by Bushway and Ponnapalam et al. (1981), baking and microwaving of potatoes resulted in lower levels (99–113 and 124–133 mg/kg fw, respectively) compared to levels in raw potatoes (103–161 mg/kg fw). The study by Lachman et al. (2013) reported that baking non-peeled tubers diced into small cubes resulted in nearly the same reduction, i.e. from 267 to 134 mg/kg dw as did microwaving (from 267 to 146 mg/kg).

When dried potato powder was heated to 150°C for 2 h, α-solanine and α-chaconine were found to be stable (Nie et al., 2018). Above 150°C both GAs started to degrade, α-chaconine being somewhat less stable than α-solanine.

Industrial multistep processing of potatoes

Studies presenting the complete process chain for the production of potato-based products are available. Some of these studies provide data on the reduction of the GA content in the intermediate as well as the initial and final products. The methods of processing leading to the different final potato products may differ according to the end-product as well as according to the producer. In general, the more recent methods will include the highest number of steps. Examples on production schemes for French fries and dehydrated potato flakes are shown in **Figure 7**.

In the production of French fries, Tajner-Czopek et al. (2014) reported reductions of up to 97.5% after exposing the potatoes to six steps of processing (**Figure 7A**). Similar results were obtained by Rytel et al. (2005) who reported that the levels in two potato cultivars processing (peeling, cutting, 2-stage blanching, drying and 2-stage frying) decreased from an initial content of 209 and 186 mg/kg dw (unpeeled potato tubers) to 14 and 6 mg/kg dw, i.e. reductions of around 93 and 98%, respectively for

the two cultivars studied. The same authors reported, in a later study, a reduction of 97% and of 84% in the production of French fries and potato crisps, respectively (Rytel et al., 2018).

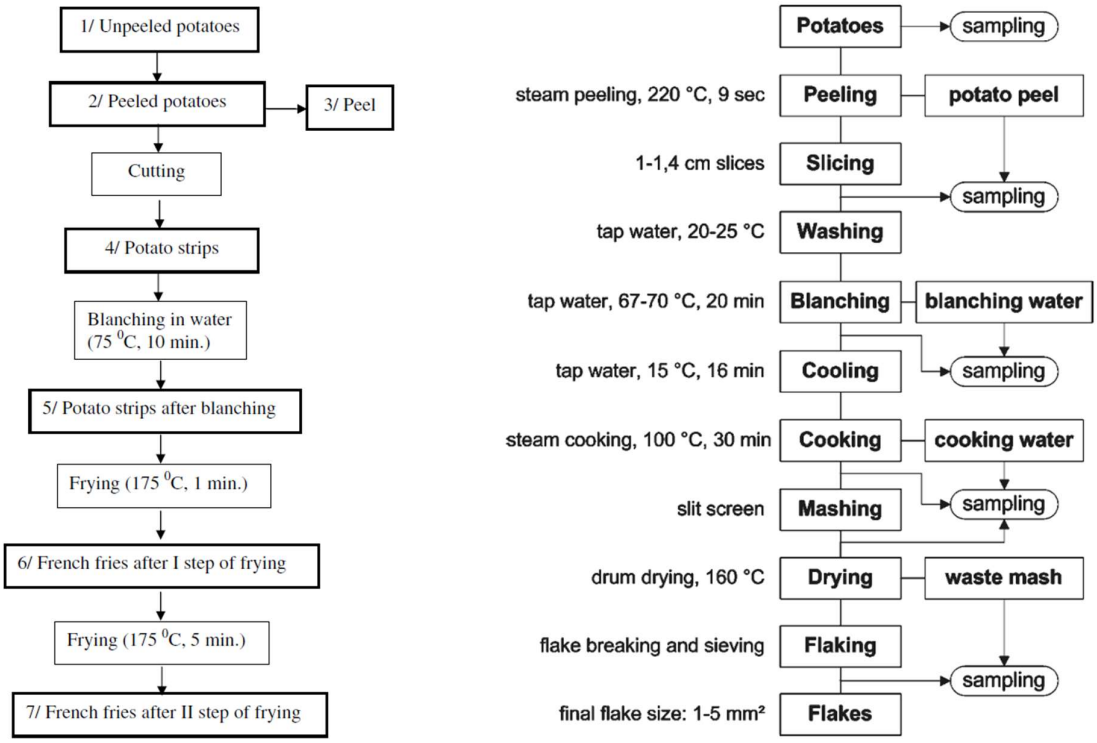


Figure 7. Examples of production schemes for **(A)** French fries (from Tajner-Czopek et al., 2014[34]) and **(B)** dehydrated potato flakes from the cultivar 'Karlena' (from Mäder et al., 2009).

The production of dehydrated potato flakes was studied using one cultivar with a starting content on TGA of 294 mg/kg dw, which after peeling, blanching, steam cooking, mashing and drying resulted in a TGA concentration of 29 mg/kg dw (corresponding to 5.8 mg/kg fw for reconstituted flakes), i.e. a reduction of around 90% (Mäder et al., 2009). Rytel et al. (2018) reported a reduction of 83% in the multi-step production of dehydrated potato (puree).

Rytel (2012b) studied the effects of industrial potato processing to obtain potato granules (dehydrated) and the changes in GA content (α-solanine and α-chaconine) at the different stages. The processing included steam peeling, slicing, blanching, cooling in water, steaming and a 2-stage drying (pneumatic and fluidization drying) to finally obtain a granulated product. After steam peeling, the TGA content decreased by 50% on average (from 160 to 81 mg/kg dw) (see **Table 38**). The authors observed a somewhat larger decrease for α-solanine (55%) than for α-chaconine (46%). After blanching a further decrease of 26% compared to the peeled potatoes was observed (from 81 to 59.6 mg/kg dw). In this case a larger decrease in α-chaconine content (29%) compared to α-solanine (21%) was observed. Cooling in water did not result in significant changes in the TGA content, but the subsequent steaming step decreased TGA by 35% on average (from 59.6 to 38.1 mg/kg dw). The last processing step of drying resulted in a further TGA decline from 38.1 to 31.1 mg/kg dw). The final granulated product contained 26.6 mg/kg dw, i.e. 16% of the original TGA content in unpeeled potatoes.

[34] Reprinted with permission from Tajner-Czopek A, Rytel E, Aniolowska M and Hamouz K, 2014. The influence of French fries processing on the glycoalkaloid content in coloured-fleshed potatoes. European Food Research and Technology, 238, 895-904. Copyright 2009, American Chemical Society.

3612 **Table 38.** Reduction in the total content of total glycoalkaloids (TGA) in *S. tuberosum* tubers as a result of peeling (as part of a commercial or experimental
3613 processing).

Study [a]	Peeling method	Number of cultivars	Analytical method	TGA initial content in unpeeled tuber (mg/kg)	TGA content in peeled tuber (mg/kg)	Reduction in TGA content (%)
Sizer et al. (1980)	Hand peeling (1–2 mm) [b]	1	Non-aqueous tritation	42.3 (fw)	23.7 (fw)	44%
Hellenäs et al. (1995)	Manual peeling [b]	1	HPLC-UV	Cultivar 'Magnum Bonum': 336 (fw)	243 (fw)	30%
Panovská et al. (1997)	Manual peeling [b]	7	HPLC-UV	178 (fw) Mean of 7 cultivars	82 (fw) Mean of 7 cultivars	54% Mean of 7 cultivars
Ostrý et al. (2010)	Hand peeling (1 mm) [b]	3 [d]	HPTLC	56–215 (fw) Across cultivars	47–177 (fw) Across cultivars	44% Across cultivars
Lachman et al. (2013)	Standard kitchen peeler (1–2 mm) [b]	6	LC-MS/MS	267 (dw) Mean of 6 cultivars	93 (dw) Mean of 6 cultivars	65% Mean of 6 cultivars
Rytel et al. (2018)	Manual peeling [b]	2 (coloured-fleshed)	HPLC-UV	Cultivar 'Valfi': 269 (dw) 'Blaue Elise': 238 (dw)	Cultivar 'Valfi': 97 (dw) 'Blaue Elise': 63 (dw)	69% Mean of 2 cultivars
Rytel et al. (2005)	n.r. (taken from a French fries production line) [c]	2	HPLC-UV	Cultivar 'Santana': 209 (dw) 'Innowator': 186 (dw)	Cultivar 'Santana': 127 (dw) 'Innowator': 141 (dw)	32% Mean of 2 cultivars
Pęksa et al. (2006)	n.r. (taken from a chip processing line) [c]	2	HPLC-UV	Cultivar 'Karlena': 129 (dw) 'Saturna': 165 (dw)	Cultivar 'Karlena': 101 (dw) 'Saturna': 118 (dw)	25% Mean of 2 cultivars
Mäder et al. (2009)	Steam-peeling [c]	1	HPTLC	294 (dw)	67 (dw)	77%
Rytel (2012a)	Laboratory carborundum peeler [c]	4	HPLC-UV	254 (dw) Mean of 4 cultivars	175 (dw) Mean of 4 cultivars	31% Mean of 4 cultivars
Rytel (2012b)	Steam-peeling [c]	1	HPLC-UV	160 (dw)	81 (dw)	50%
Tajner-Czopek et al. (2014)	Mechanical peeler [c]	4 (coloured-fleshed)	HPLC-UV	50 (fw) Mean of 4 cultivars	25 (fw) Mean of 4 cultivars	50% Mean of 4 cultivars

3614 TGA: total glycoalkaloids. dw: dry weight. fw: fresh weight. n.r.: not reported. LC: liquid chromatography. UV: ultraviolet. MS: mass spectrometry. MS/MS: tandem MS. HR: high resolution. HP: high
3615 performance. HPTLC: high performance thin layer chromatography.
3616 (a): Study year is in this case the year of publication.
3617 (b): Household-like peeling.
3618 (c): Industrial-like peeling.
3619 (d): A total of 20 samples was tested.

3620 **Table 39**. Reduction in the total content of total glycoalkaloids (TGA) in *S. tuberosum* tubers as a result of boiling, blanching and/or steaming (as part of a
3621 commercial or experimental processing). Note: the reduction is given as the percent reduction in the TGA content after peeling.

Study [a]	Processing step(s)	Number of cultivars	Analytical method	TGA initial content in peeled tuber (mg/kg)	TGA content in cooked tuber (mg/kg)	Reduction in TGA content (%)
Takagi et al. (1990)	Boiling in water	3	HPLC-UV	91–135 [d]	90–126 [d]	5% Mean of 3 cultivars
Mulinacci et al. (2008)	Boiling in water [c]	3 (red- and blue-fleshed)	HPLC-UV	Cultivar 'Kennebec': 46 (fw) 'Vitelotte': 79 (fw) 'Highland Burgundy': 180 (fw)	Cultivar 'Kennebec': 29 (fw) 'Vitelotte': 66 (fw) 'Highland Burgundy': 153 (fw)	23% Mean of 3 cultivars
Ostrý et al. (2010)	Boiling in salted water until edible stage	3 [e]	HPTLC	117–217 (fw) Across cultivars	102–180 (fw) Across cultivars	19% Across cultivars
Lachman et al. (2013)	Boiling	6	LC-MS/MS	92.6 (dw) Mean of 6 cultivars	79.7 (dw) Mean of 6 cultivars	14% Mean of 6 cultivars
Rytel et al. (2018) [b]	Blanching of potato strips	2 (blue-fleshed)	HPLC-UV	Cultivar 'Valfi': 97 (dw) 'Blaue Elise': 63 (dw)	Cultivar 'Valfi': 60 (dw) 'Blaue Elise': 44 (dw)	34% Mean of 2 cultivars
Rytel et al. (2018) [b]	Blanching of potato halves	2 (blue-fleshed)	HPLC-UV	Cultivar 'Valfi': 97 (dw) 'Blaue Elise': 63 (dw)	Cultivar 'Valfi': 79 (dw) 'Blaue Elise': 51 (dw)	19% Mean of 2 cultivars
Rytel et al. (2018) [b]	Blanching + steaming of potato halves	2 (blue-fleshed)	HPLC-UV	Cultivar 'Valfi': 97 (dw) 'Blaue Elise': 63 (dw)	Cultivar 'Valfi': 64 (dw) 'Blaue Elise': 42 (dw)	34% Mean of 2 cultivars
Rytel et al. (2005)	Blanching (2 stage)	2	HPLC-UV	Cultivar 'Santana': 127 (dw) 'Innowator': 141 (dw)	Cultivar 'Santana': 55 (dw) 'Innowator': 48 (dw)	62% Mean of 2 cultivars
Pęksa et al. (2006)	Blanching	2	HPLC-UV	Cultivar 'Karlena': 101 (dw) 'Saturna': 118 (dw)	Cultivar 'Karlena': 38 (dw) 'Saturna': 37 (dw)	65% Mean of 2 cultivars
Mäder et al. (2009)	Blanching + steam cooking	1	HPTLC	67 (dw)	34 (dw)	49%
Rytel (2012a)	Blanching	4	HPLC-UV	175 (dw) Mean of 4 cultivars	126 (dw) Mean of 4 cultivars	28% Mean of 4 cultivars
Rytel (2012a)	Steaming and cooling	4	HPLC-UV	175 (dw) Mean of 4 cultivars	101 (dw) Mean of 4 cultivars	42% Mean of 4 cultivars
Rytel (2012b)	Blanching + steaming	1	HPLC-UV	81 (dw)	38 (dw)	53%

Tajner-Czopek et al. (2014)	Blanching	4 (coloured-fleshed)	HPLC-UV	25 (fw) Mean of 4 cultivars	- (f)	~16% Mean of 4 cultivars

dw: dry weight. fw: fresh weight. LC: liquid chromatography. UV: ultraviolet. MS: mass spectrometry. MS/MS: tandem MS. HR: high resolution. HP: high performance. HPTLC: high performance thin layer chromatography.
(a): Study year is in this case the year of publication.
(b): Study year is 2014–2015.
(c): Experiments were conducted with unpeeled potatoes.
(d): Results were presented for α-solanine and α-chaconine.
(e): A total of 12 samples was tested.
(f): Results provided in figures for α-solanine and α-chaconine.

Table 40. Reduction in the total content of total glycoalkaloids (TGA) in *S. tuberosum* tubers as a result of frying for the production of French fries or potato crisps. Note: the reduction is given as the percent reduction in the TGA content after peeling.

Study (a)	Processing step(s)	Number of cultivars	Analytical method	TGA initial content in peeled tuber (mg/kg)	TGAs content in fried potato product (mg/kg)	Reduction in TGA content (%)
Takagi et al. (1990)	Frying (5 min at 150°C)	3	HPLC-UV	222–243 (c),(d)	218–231 (c),(d)	3% Mean of 3 cultivars
Takagi et al. (1990)	Frying (5 min at 170°C)	4	HPLC-UV	185–373 (c),(d)	169–294 (c),(d)	21% Mean of 4 cultivars
Takagi et al. (1990)	Frying (10 min at 210°C)	3	HPLC-UV	114–234 (c),(d)	64–149 (c),(d)	38% Mean of 3 cultivars
Rytel et al. (2005)	Frying at 170–180°C (2 stage) (French fries processing)	2	HPLC-UV	Cultivar 'Santana': 127 (dw) 'Innowator': 141 (dw)	Cultivar 'Santana': 14 (dw) 'Innowator': 6 (dw)	92% Mean of 2 cultivars
Pęksa et al. (2006)	Frying at 180°C (chip processing (b))	2	HPLC-UV	Cultivar 'Karlena': 101 (dw) 'Saturna': 118 (dw)	Cultivar 'Karlena': 24 (dw) 'Saturna': 23 (dw)	79% Mean of 2 cultivars
Tajner-Czopek et al. (2014)	Frying at 175°C (2 stage) (French fries processing)	4	HPLC-UV	25 (fw) mean of cultivars	- (d)	~48% Mean of 4 cultivars
Rytel et al. (2018) (b)	Frying (2-stage, French fries processing)	2 (blue-fleshed)	HPLC-UV	Cultivar 'Valfi': 97 (dw) 'Blaue Elise': 63 (dw)	Cultivar 'Valfi': 9.5 (dw) 'Blaue Elise': 7.0 (dw)	89% Mean of 2 cultivars
Rytel et al. (2018) (b)	Frying (potato crisps procesing)	2 (blue-fleshed)	HPLC-UV	Cultivar 'Valfi': 97 (dw) 'Blaue Elise': 63 (dw)	Cultivar 'Valfi': 53 (dw) 'Blaue Elise': 28 (dw)	50% Mean of 2 cultivars

TGA: total glycoalkaloids. dw: dry weight. fw: fresh weight. LC: liquid chromatography. UV: ultraviolet. HP: high performance.
(a): Study year is in this case the year of publication.
(b): It is not clear whether the final products refers to French fries or to potato crisps.

(c): Results were presented for α-solanine and α-chaconine.
(d): Results provided in figures for α-solanine and α-chaconine.

Table 41. Reduction in the total content of total glycoalkaloids (TGA) in *S. tuberosum* tubers as a result of baking or microwaving. Note: the reduction is given as the percent reduction compared to the original GA content in the unpeeled potato.

Study [a]	Processing step(s)	Number of cultivars	Analytical method	TGA initial content in fresh tuber (mg/kg)	TGA content in processed tuber (mg/kg)	Reduction in TGA content (%)
Bushway and Ponnampalam (1981)	Baking [b]	2	HPLC-UV	103–161 (fw)	99–113 (fw)	20% Mean of 2 cultivars
Lachman et al. (2013)	Baking [b]	6	LC-MS/MS	267 (dw) Mean of 6 cultivars	135 (dw) Mean of 6 cultivars	49% Mean of 6 cultivars
Bushway and Ponnampalam (1981)	Microwaving [b]	2	HPLC-UV	103–161 (fw)	124–133 (fw)	3% Mean of 2 cultivars
Takagi et al. (1990)	Microwaving [c]	3	HPLC-UV	86–239 [d]	72–208 [d]	15% Mean of 3 cultivars
Lachman et al. (2013)	Microwaving [b]	6	LC-MS/MS	267 (dw) Mean of 6 cultivars	146 (dw) Mean of 6 cultivars	45% Mean of 6 cultivars
Mulinacci et al. (2008)	Microwaving [b]	3 (red- and blue-fleshed)	HPLC-UV	Cultivar 'Kennebec': 46 (fw) 'Vitelotte': 79 (fw) 'Highland Burgundy': 180 (fw)	Cultivar 'Kennebec': 38 (fw) 'Vitelotte': 75 (fw) 'Highland Burgundy': 168 (fw)	7% Mean of 3 cultivars [c]

TGA: total glycoalkaloids. LC: liquid chromatography. UV: ultraviolet. MS/MS: tandem mass spectrometry. HP: high performance.
(a): Study year is in this case the year of publication.
(b): Experiments were conducted with unpeeled potatoes.
(c): Experiments were conducted with peeled potatoes, cut into cubes of 5-10 mm.
(d): Results were presented for α-solanine and α-chaconine.

Potato peels

Bushway and Ponnampalam (1981) studied the effect of frying (4 min at 177°C) on the TGA content in raw potato peels. A reduction of 16–22% (678–763 mg/kg to 567–594 mg/kg, corrected for moisture loss) was observed.

Similar results were obtained by Bushway et al. (1983). Starting from raw potato peels, containing GAs in the range of 18.0 to 1,068 mg/kg across cultivars, the levels of GAs in baked potato peels were 24 to 1,098 mg/kg across cultivars, in fried peels 32.7 to 1,649 mg/kg and in baked-fried 41.7 to 1,557 mg/kg (all concentrations uncorrected for moisture loss). The authors attributed this difference to the greater moisture loss during frying than during baking.

3.2.3.1.3. Processing of potatoes for feed

Whole potatoes intended for use as feeds for ruminants normally undergo no further processing [35] and therefore no effects on GA concentrations would be expected. When the supply of potatoes exceeds demand, they may also be ensiled (either alone or with the addition of a green forage), but no reports have been identified on the effects of ensiling on the GA contents.

For pigs, it is generally recommended that potatoes are cooked or steamed before being fed (Whittemore, 1977), and the effects of these processes on GA concentrations when applied to the production of foods, as reported above, may also be expected to apply to feed.

A number of by-products of potato processing are used as feeds for livestock and companion animals. These include potato protein and potato peelings. Where the peels have been removed by steaming, it should be noted that GAs are stable to decomposition at cooking temperatures below 150°C, and therefore potato peelings obtained by steaming might represent a risk to livestock, depending on the extent of greening in the tubers (Bushway and Ponnampalam, 1981; Takagi et al., 1990).

Drying of industrial potato peels resulted in an increase of GA levels (Hossain et al., 2016). The levels of α-solanine increased from ~100 mg/kg dw in the fresh peels to nearly 400 mg/kg dw after drying either by air-, vacuum oven- or freeze-drying. Similar behaviour was observed for α-chaconine, increasing from ~50 to ~600 mg/kg dw. Air drying showed the highest increase in GA levels. The levels of solanidine and demissidine appeared not to be affected by the drying process.

Potato peelings produced as a by product from industrial processing were stored for 4 days at room temperature to investigate the effect on the GA content. A slow degradation of the GAs was observed with calculated half lifes in the range of 2.9 to 9.5 days, depending on the size of the potato peel wastes (Sepelevs et al., 2019).

3.2.4.2. GAs from food plants other than *S. tuberosum*

The α-tomatine concentration decreases during fruit ripening (see **Section 1.3.3.2**). The concentration in the ripe fruit furthermore differs depending on the cultivar. While a number of published investigations on the effect of different processing steps on the GA concentration in potatoes and potato products exist, those regarding tomato and tomato products are scarce. Examples of the content of α-tomatine in different tomato products after processing are shown in **Table 36**.

Friedman and Levin (1995) reported that microwaving of green tomatoes only caused a 7% loss of α-tomatine, while preparation of fried green tomatoes caused a 22% loss, when adjusting for added ingredients and moisture changes. No significant reduction was noted for the content of α-

[35] They may be chopped or sliced to reduce the risk of choking.

dehydrotomatine (measured as 'solanine' [36]) upon preparation of green tomato marmalade, while preparation of pickled green tomatoes resulted in a loss of 11% (Kibler et al., 1985). Kyzlink et al. (1981) found that cooking for 5 min of unripe green tomatoes resulted in a loss of 0–15% α-tomatine. Freeze-drying of the produced tomato homogenate and subsequent storage for 4 weeks at room temperature, resulted in a reduction of the α-tomatine content of 82–85% (Kyzlink et al., 1981). Voldrich et al. (1992) also noted that the α-tomatine content of tomatoes and tomato products (fresh or fermented) declines upon storage. Two batches of unripe green tomatoes did not contain measurable amounts of α-tomatine after been stored at 4°C for 10 weeks (starting content 52 and 83 mg/kg fw, respectively). Similar results were found for products of green tomato salad, fermented green tomato and tomato puree that had been stored for 2 months (Voldrich et al., 1992). Stoyanova et al. (1988) reported for pickled immature tomatoes a reduction in α-tomatine content from an initial 360–400 mg/kg fw to 240–340 mg/kg fw after 10 days and to 120–160 mg/kg fw after 180 days. After 6 months of storage the tomatoes had lost their initial bitter taste.

A single study has been published on the effect of cooking and grilling on the content of GAs in 3 cultivars of aubergine. Boiling for 10 min reduced the TGA content by 6 to 35% due to leaching, while grilling for 5 min resulted in 8% to 107% increase in TGA content, what the authors contributed to a water loss of the grilled product (Lo Scalzo et al., 2016).

3.2.4.3. Summary on influence of storage and processing on the levels of GAs

For potatoes, factors such as the duration of storage, temperature, light exposure, as well as tuber damage can affect the GA content during storage of potato tubers. Varying results have been obtained in the different studies depending on the conditions and the potato cultivar studied. In general, longer storage times, exposure to light and tuber damage have been reported to induce the formation of GAs. The influence of temperature is dependent on the potato cultivar and other conditions tested in the studies.

Processing of potatoes has been reported to reduce the content of GAs in the final processed product. In general, the peeling of potatoes reduced the GA content by 25% up to 75% depending on the peeling method used. Boiling in water and blanching reduced the GA content in peeled potatoes in the range of 5% to 65%. Frying in oil lowered the content from 20% up to 90% compared to the peeled potato tuber. Microwave and oven baking of unpeeled potatoes have also been reported to cause a reduction in the GA content by 3% to 45% and by 20% to 50%, respectively.

No information has been found about the chemical nature of the GAs degradation products.

For tomatoes, the ripening process leads to a decrease of the content of α-tomatine and α-dehydrotomatine. Factors that may lead to an increase of α-tomatine and α-dehydrotomatine after harvesting have not been identified. Processing of tomatoes may result in modest losses of α-tomatine of up to 15%. Storage at low temperatures may slow down the ripening of tomatoes and the concomitant decrease of α-tomatine, but does not fully stop it. Storage of tomato products also results in a loss of α-tomatine over time.

For aubergine, the ripening process has been reported to result in an increase of the GA content. Other factors that may influence the formation of GAs have not been reported. Boiling in water may reduce the TGA content by 6 to 35%, while grilling may lead to an increased TGA content in the grilled product, due to a lower water content.

[36] The authors report the presence of 'solanine' in tomato and tomato products. It should be noted that the analytical method (colourimetry) used in this publication cannot distinguish between α-solanine, α-chaconine and α-dehydrotomatine. It is therefore likely that the authors have measured α-dehydrotomatine.

3.3. Exposure assessment

3.3.1. Current dietary acute exposure assessment for humans

The CONTAM Panel assessed the dietary acute exposure to potato TGAs (sum of α-solanine and α-chaconine) in food following the methodology described in **Section 2.5**.

A scenario including only days in which there was consumption of main-crop potatoes was considered the most relevant for acute risk assessment. The summary statistics of the probabilistic acute dietary exposure assessment to potato TGAs across European dietary surveys by age group are shown in **Table 42**. Detailed results for each survey are shown in **Annex A6**.

The mean exposure across surveys ranged from 26.0 µg/kg bw per day in adults to 193.4 µg/kg bw per day in toddlers. The 95th percentile exposure ranged from 88.2 µg/kg bw per day in adults to 617.9 µg/kg bw per day in Toddlers (up to 1,057.9 µg/kg bw per day in the upper limit of the 95% confidence interval).

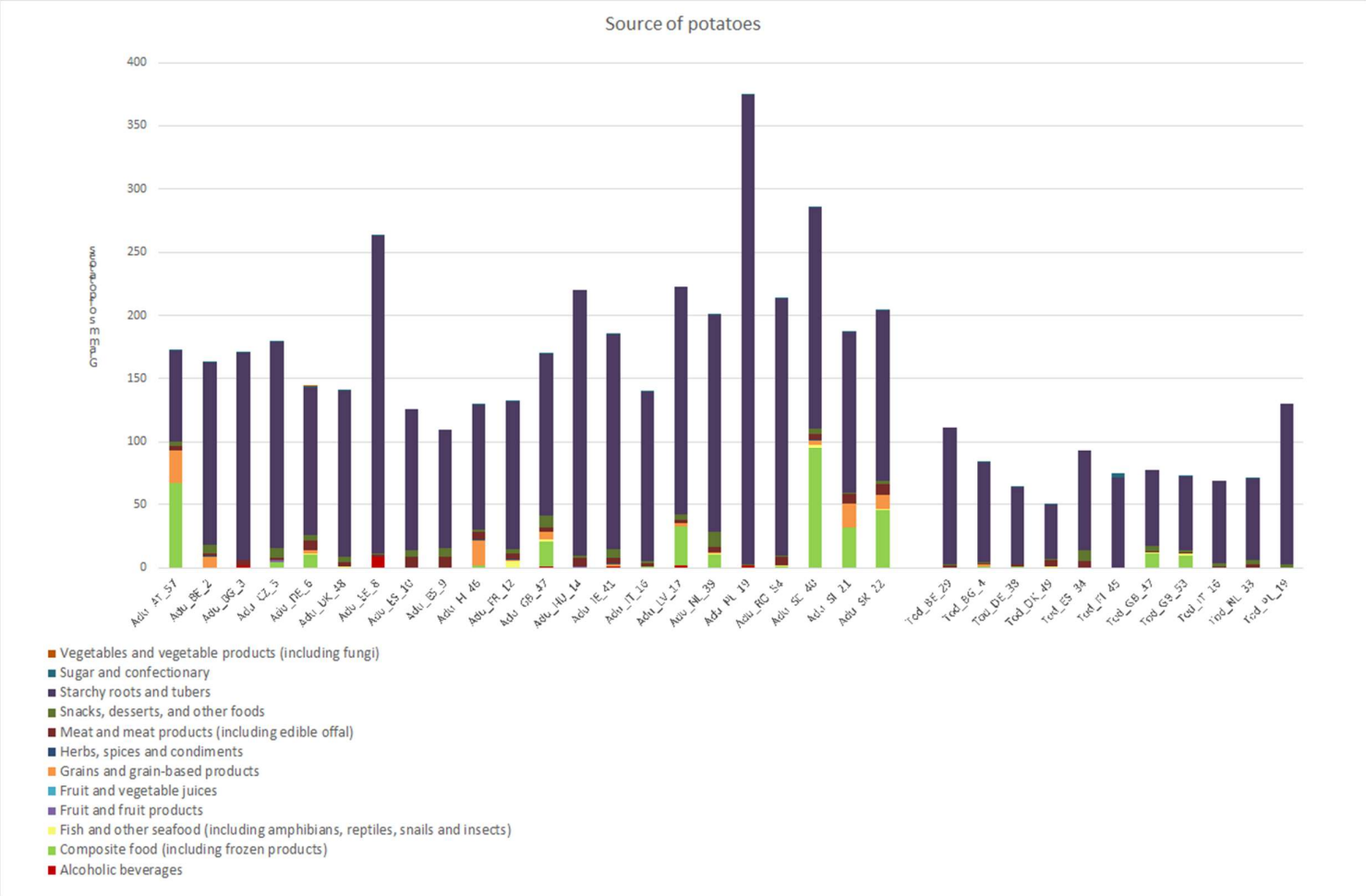
Table 42. Summary statistics of the probabilistic acute dietary exposure assessment to potato TGAs across European dietary surveys (µg/kg bw per day) by age group including only days with consumption. The corresponding 95% confidence intervals are presented in the brackets.

Age class	Range across durveys of acute dietary exposure (µg/kg bw per day)			
	Mean		P95 [(a)]	
	Minimum	Maximum	Minimum	Maximum
Infants	35.4 (19.3 - 68.8)	133.0 (124.1 - 143.8)	349.1 (319.5 - 382.6)	422.0 (376.9 - 468.4)
Toddlers	66.6 (63.1 - 70.6)	193.4 (140.7 - 284.1)	254.8 (237.3 - 272.9)	617.9 (369.4 - 1,057.9)
Children	54.7 (49.0 - 62.1)	167.1 (143.9 - 194.0)	178.7 (150.8 - 210.4)	518.6 (364.0 - 747.2)
Adolescents	35.5 (32.5 - 39.2)	122.8 (109.4 - 138.0)	116.3 (102.4 - 133.0)	377.4 (314.9 - 447.1)
Adults	26.0 (22.7 - 30.1)	91.7 (86.6 - 97.6)	88.2 (73.7 - 106.7)	277.2 (255.6 - 302.4)
Elderly	29.1 (21.9 - 40.8)	80.0 (68.1 - 95.8)	96.7 (89.1 - 104.7)	240.4 (189.3 - 309.6)
Very elderly	32.3 (29.2 - 36.1)	79.6 (62.0 - 108.0)	96.5 (84.1 - 110.6)	250.0 (170.3 - 385.9)

bw: body weight; P95: 95th percentile
(a): One dietary survey had less than 60 days in the Infants and Toddlers age groups, therefore, these were not included in calculation of the 95th percentile exposure.

Figure 8 shows the mean amounts (grams per day) of main-crop potatoes consumed by adults or toddlers via different food categories. Method details are given in **Section 2.5**. Data were available for 22 European countries. In the age group Adults the largest part of main crop potatoes is consumed as 'Starchy roots and tubers', followed by the food categories 'Composite food' and 'Grains and grain-based products'. Food surveys for toddlers show that the food category 'Starchy roots and tubers' is predominant with no significant differences among the 11 European countries, for which data are available. These findings indicate that the main dietary GA source via potatoes and potato-derived products for adults and toddlers in Europe are 'Starchy roots and tubers' while other food categories appear to be of considerably less importance.

3755 **Figure 8.** Mean daily amounts of main-crop potatoes (grams per day) calculated on consumption days by food source category for Adults and Todders across
3756 the different surveys.



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3.3.2. Previously reported dietary exposure assessments

Ruprich et al. (2009) estimated the acute and chronic exposure to GAs using probabilistic modelling for Sweden, Czech Republic and the Netherlands. The authors combined the data on occurrence of α-solanine and α-chaconine in potato tubers available from the Czech Republic (n = 200 analytical results, peeled raw potatoes, year 2004–2005) and from Sweden (n = 239 analytical results, raw unpeeled potatoes, year 1997) to perform the overall exposure assessment for the three countries. Since the data from the Czech Republic referred to peeled potatoes, a factor of 1.25 was applied to correct for GAs losses during peeling. The final dataset contained 439 results for raw unpeeled potatoes. In addition, for the acute exposure estimation, four other scenarios were considered to simulate a situation in which raw unpeeled potatoes contained limited quantities of GAs, by deleting concentrations over 200, 150, 100 or 50 mg/kg. This resulted in four occurrence datasets with the following median (average, range) concentrations:

(i) Complete dataset (n = 439): 64.0 (70.4, 6.3–302.9) mg/kg
(ii) Cut off 200 mg/kg (n = 421): 62.5 (63.5, 6.3–197.8) mg/kg
(iii) Cut off 150 mg/kg (n = 400): 60.2 (58.0, 6.3–149.9) mg/kg
(iv) Cut off 100 mg/kg (n = 333): 48.1 (45.5, 6.3–99.8 mg/kg
(v) Cut off 50 mg/kg (n = 173): 15.1 (18.2, 6.3–49.4) mg/kg

In addition, processing factors were included in the assessment to correct for losses of GAs during processing of potatoes. Factors of 0.8, 0.64 and 0.17, respectively, were considered for peeling, for peeling and cooking, and for frying or baking in fat. Food consumption data was retrieved from the national databases and for the Czech Republic comprised subjects from 4 to 90 years old, for Sweden subjects from 17 to 74 years old, and from The Netherlands subjects from 1 to 97 years old. The consumption data on potato products were recalculated at raw agricultural commodity level. The probabilistic estimation was done using the Monte Carlo Risk Assessment software.

Mean (and upper 97.5% confidence limit) acute exposure estimates, when all occurrence data were considered were 80 (85), 80 (87) and 78 (84) µg/kg bw per day for the Czech Republic, Sweden and the Netherlands, respectively. The calculated P95 (and upper 97.5% confidence limit) acute dietary exposures were 358 (383), 339 (366), 319 (344) µg/kg bw per day for the Czech Republic, Sweden and the Netherlands, respectively. The estimates were lower when the different cut off scenarios were used (200, 150, 100 and 50 mg/kg cut off). For the Czech Republic the mean acute estimates for these scenarios were 72, 66, 52 and 21 µg/kg bw per day, respectively, for Sweden they were 73, 67, 53 and 21 µg/kg bw per day, respectively, and for the Netherlands they were 71, 64, 51 and 20 µg/kg bw per day, respectively. The P95 acute exposures using the different cut off scenarios were for the Czech Republic: 318, 288, 228 and 106 µg/kg bw per day, respectively; for Sweden: 302, 273, 217 and 100 µg/kg bw per day, respectively and for the Netherlands 283, 258, 205 and 94 µg/kg bw per day, respectively. Median (and upper 97.5% confidence limit) chronic estimates of intake were 74 (79), 73 (79) and 154 (166) µg/kg bw per day. The authors noted that the differences in intake estimates between countries were due not only to different consumption habits but also due to the structure and complexity of the food consumption database used in each case. Considering 1 mg/kg bw per day as the critical exposure dose for adverse effects, the authors concluded that 0.01% of the population in all three countries exceeded this value. No chronic estimates were above that level.

Pariera Dinkins and Paterson (2008) estimated the acute oral exposure to GAs (sum of α-solanine and α-chaconine) considering the occurrence in the inner tissue of tubers from plants defoliated by Colorado potato beetles and with no defoliation (control) for different age groups within the US population. The authors used a dataset for peeled potatoes with mean (standard deviation) GAs concentrations of 174.5

± 8.59 mg/kg dw for the control and 258.8 ± 23.3 mg/kg dw for Colorado potato beetle defoliation samples. Estimates of acute exposure were higher in the scenario with Colorado potato beetle defoliation samples compared to the control scenario. The median exposure estimates for the US population were 239 and 354 µg/kg bw per day for the controls scenario and for the Colorado potato beetle defoliation scenario, respectively. Acute oral exposures were also estimated for seven population subgroups including infants, children 7–12 years old, youth 13–19-year-old, women >13 years old, women >20 years old, and males >20 years old). The median of exposure in the control scenario ranged from 190 to 515 µg/kg bw per day, while in the potato beetle defoliation scenario it ranged from 282 to 760 µg/kg bw per day. The estimated P95 acute exposure ranged from 406 to 1,131 µg/kg bw per day in the control scenario and from 603 to 1,676 µg/kg bw per day in the potato beetle defoliation scenario. Highest exposures were calculated for the age group children 1–6 years. It should be noted that it is not indicated whether reduction factors for heat processing steps were applied to calculate the exposure.

A detailed comparison between these studies and the current exposure assessment is not possible, since the three studies used different data sets, age groups and exposure models. Nevertheless, all studies indicate that there is a likelyhood that the LOAEL of 1 mg/kg bw per day is exceeded in a number of cases, with the largest probability for the younger age groups.

3.3.3. Current dietary exposure assessment for farm animals, horses and companion animals

While potatoes are not widely used as feeds for livestock, in some situations they may represent an important component of the ration, for example when production exceeds market requirements for human consumption. Potato by-products, particularly potato protein and potato starch, are used to a limited extent in compound feeds for certain non-ruminant livestock and companion animals. However, insufficient data were available on levels of GAs in either potatoes or potato by-products used as animal feeds.

3.4. Risk characterisation

3.4.1. Human health risk characterisation

3.4.1.1. GA from edible parts of *S. tuberosum*

The available data on acute toxicity were considered insufficient to establish a HBGV, and instead the CONTAM Panel used an MOE approach to assess a possible health concern from acute exposure to potato TGAs via food. An MOE higher than 10 indicates that there is no health concern. This MOE of 10 takes into account the interindividual variability in toxicodynamics (a factor of 3.2), and extrapolation from a LOAEL to a NOAEL (a factor of 3).

The LOAEL of 1 mg potato TGA/kg bw in humans was used as reference point to characterise the risk following acute exposure.

The CONTAM Panel calculated the acute dietary exposure using a probabilistic approach and applying reduction factors to estimate the GA levels in the final food consumed as described in **Section 2.5**, **3.2.1** and **3.3.1**. Comparison of the acute exposure estimates (see **Table 43**) to the reference point of 1 mg potato TGAs/kg bw per day resulted in the MOE values that are presented in **Table 43** for acute mean and P95 exposure values across surveys and age groups.

3843 **Table 43.** Margin of exposure (MOE) values for the range of acute mean and P95 exposure assessment
3844 across different surveys.

MOE	Mean exposure		P95 exposure	
	Minimum	Maximum	Minimum	Maximum
Infants	28	8	3	2
Toddlers	15	5	4	2
Children	18	6	6	2
Adolescents	28	8	9	3
Adults	39	11	11	4
Elderly	34	13	10	4
Very elderly	31	13	10	4

3845 The MOEs for the younger age groups indicate a health concern for the food consumption surveys with
3846 the highest mean exposure, as well as for the P95 exposure in all surveys. For adult age groups, the
3847 MOEs indicate a health concern only for the food consumption surveys with the highest P95 exposures.

3848 To estimate the consequences of these findings, the Panel calculated the mean percentage of days
3849 across surveys per age group on which the potato TGA intake may be below the MOE of 10 (**Table 44**).
3850 The highest number of survey days with intake of potatoes below the MOE of 10 was estimated for
3851 toddlers (56%) followed by children (50%). For the other age groups the estimated intake was
3852 calculated to be below the MOE of 10 in 23 to 40% of the survey days.

3853 **Table 44.** Summary statistics of the % of survey days with an intake of potato TGAs below the margin
3854 of exposure (MOE) of 10 calculated only for days of potato consumption. The corresponding 95%
3855 confidence intervals (95% CI) are given in the brackets.

Age class	Range of mean % of days below the MOE of 10 [(a)]	
	Minimum	Maximum
Infants	5 (0 - 21)	40 (38 - 42)
Toddlers	19 (18 - 20)	56 (45 - 67)
Children	14 (12 - 17)	50 (45 - 54)
Adolescents	7 (5 - 8)	38 (34 - 41)
Adults	4 (2 - 6)	28 (26 - 29)
Elderly	4 (0 - 9)	23 (18 - 28)
Very elderly	5 (3 - 6)	23 (16 - 31)

3856 (a) The mean percentage of days below the MOE of 10 per age group for each survey was calculated by averaging
3857 the percentage of days below the reference for each subject and then averaged over all subjects within the age group
3858 for each survey. The process was iterated 1,000 times as for the exposure estimates, and a mean of the mean
3859 percentage of days below the reference per age group was calculated together with a 95% confidence interval.

3860 The Panel also calculated the mean percentage of days across surveys per age group on which the
3861 potato TGA intake may be above the LOAEL of 1 mg/kg bw. Exposure was estimated to be above the
3862 LOAEL of 1 mg/kg bw for toddlers and children on maximally 2% and 1% of the survey days,
3863 respectively, and on less than 1% of the survey days for the other age groups (see **Appendix H**).

3864

3.4.1.2. GAs from edible parts of food plants other than *S. tuberosum*

No occurrence data on tomato or aubergine-derived GAs or aglycones were submitted to EFSA. The data from the literature indicate that the concentration of tomato GAs is lower than of potato GAs in the various Solanum species (see **Section 3.2.2.1.2**). There are few data on the occurrence of GAs in aubergines (see **Section 3.2.2.1.3**). Overall, the CONTAM Panel concluded that the data from the literature are too limited to attempt an exposure assessment for tomato or aubergine GAs.

In addition, the lack of any human study and insufficient experimental toxicity data for tomato and aubergine GAs (no long-term studies, no dose-response in repeated dose studies) impairs the identification of a reference point.

To conclude, there are insufficient data to perform a risk characterisation for tomato and aubergine GAs.

3.4.2. Farm animals, horses and companion animal risk characterisation

Since no studies on the adverse effects of GAs in farm animals, horses or companion animals were identified, which could be used for the identification of a reference points, no risk characterisation could be performed.

3.5. Uncertainty analysis

The evaluation of the inherent uncertainties in the assessment of exposure to GAs has been performed following the guidance of the Opinion of the Scientific Committee related to Uncertainties in the Dietary Exposure Assessment (EFSA, 2007). In addition, the report on 'Characterizing and Communicating Uncertainty in Exposure Assessment' has been considered (WHO/IPCS, 2008). The CONTAM Panel took note of the new guidance on uncertainties of the Scientific Committee (EFSA Scientific Committee, 2018), but it was not implemented in this opinion.

The CONTAM Panel assessed the risk for human health following acute exposure to potato GAs. The data available for repeated dose toxicity and on reproduction in experimental animals did not allow to draw conclusions or to establish a reference point for a chronic exposure risk assessment. In this respect, the Panel noted that: (i) the repeated dose studies on the aglycone solanidine in rodents showed adverse effects being different from those induced by the respective potato GAs, (ii) the developmental effects of potato GAs and aglycones have not been studied over a dose-range, for the entire gestational period until weaning, and in species other than hamster, (iii) from the limited data available in humans relatively long serum half-lives were reported for potato GAs suggesting a potential for bioaccumulation, (iv) no conclusive toxicity data on repeated exposure of humans with GAs and aglycones are available.

Due to limited information no risk assessment was possible for effects of potato GAs and their aglycones for farm animals, horses and companion animals.

Similarly, a risk assessment for the effects of tomato and aubergine GAs and their aglycones for humans could not be performed.

Therefore, the evaluation of uncertainties in this section focuses only on the assessment of the risk following acute exposure to potato GAs and aglycones in humans.

3.5.1. Assessment objectives

The objectives of the assessment are clarified in **Section 1.2** on Interpretation of the Terms of Reference.

3.5.2. Exposure scenario / exposure model

Occurrence data in food

In most studies only α-solanine and α-chaconine have been looked for and reported, while other, minor GAs as well as β- and γ-forms of solanine and chaconine, have not been investigated. However, these would represent around 5% of the total potato GAs content, and thus represent a minor uncertainty.

In response to EFSA's request to submit occurrence data on GAs, only 3 countries provided data, thus country-based differences in the levels of GA in main-crop potatoes is not evenly represented in the available data, introducing an uncertainty on the representativeness of the overall statistics. Differences in GA levels in potatoes, due to the use of different cultivars, location, different growing conditions, and year to year variability, across European countries may result in over- or underestimation of exposure for certain food consumption surveys.

For 95% of the samples the information concerning specifically the peeling of the analysed potatoes was not reported. However, most of the samples were reported as unprocessed while for 156 samples this information was not available. For the latter samples, the CONTAM Panel considered it unlikely they were peeled, and thus they were assumed to have been analysed unpeeled. In case these (or part of) 156 samples were peeled, this might have caused an underestimation of the GA occurrence.

The analytical results used for the exposure assessment were obtained using different analytical methods, different detection techniques and with varying LODs/LOQs. The ratio between α-solanine and α-chaconine was found to differ between data submissions, indicating differences between the analytical methodologies. This is more likely to result in an underestimation than in an overestimation of the exposure.

Food processing / reduction factors

Important sources of uncertainty in the exposure assessment are related to the different assumptions made, such as that the occurrence values obtained from sampled potatoes are representative of consumed potatoes, and to the assignment of reduction factors associated with processing. Literature studies, however, report a wide range of reduction factors, introducing uncertainty on the actual reduction during the various processing steps. To take into account the reduction by peeling and heat processing, several assumptions were made (see **Section 2.5**). These assumptions contribute to the uncertainty of the exposure assessment and they could have led to an over- or underestimation of the exposure.

It is not known to which extent these food processing steps result in only a partial degradation of GAs (to β- and γ-forms of solanine and chaconine) or degradation to the aglycone (solanidine). This may result in a slight underestimation of the GA content present in food products as consumed.

Food Consumption data

Uncertainties and limitations related to the use of the EFSA Comprehensive Food Consumption Database were described by EFSA (EFSA, 2011a) and are not further detailed in this Scientific Opinion. These relate to the use of different dietary survey methodologies, standard portion sizes, representativeness of samples included in surveys, or to the inclusion of consumption surveys covering only few days to estimate high percentiles of chronic exposure.

Uncertainties and limitations related to the use of the RPC model related to the process of converting the consumed amount of composite foods to the amount of raw primary commodity were also described by EFSA (EFSA, 2019) and are not further detailed in this Scientific Opinion.

3.5.3. Hazard identification and characterisation

Toxicokinetics

The available studies did not allow for a full characterization of the absorption, distribution, metabolism and excretion profile for α-solanine and α-chaconine. Differences in absorption and excretion were observed for these two GAs in rats and hamsters, with indications of higher absorption and slower excretion rates in the latter species. There is uncertainty on the most representative species for the human risk assessment.

Interactions

The possibility that components different from GAs in potatoes, such as spirostane saponins, may enhance the toxicity of GA has also to be considered. As discussed by McMillan and Thompson (1979) in their report on an outbreak of poisoning in schoolboys associated with the consumption of toxic potatoes, saponins might promote gastrointestinal absorption of GAs or contribute to irritation of the gastrointestinal tract. This might contribute to an overestimation of the risk from the potato GAs alone.

Observations in Humans

A NOAEL for local effects in the gastrointestinal tract of potato GAs could not be identified.

Data on adverse effects of potato GAs were only available for adults and school children, but are lacking for infants, toddlers and young children.

Available data suggest interindividual differences in the susceptibility to the adverse effects of potato GAs in the gastrointestinal tract.

Intake figures of potato GAs for case reports were in general estimates which causes uncertainties on the shape of the dose-response relationship and which may lead to over or under estimation.

Non-specific symptoms like nausea and gastrointestinal discomfort may not always have been associated with intake of potato GAs. This might lead to under-reporting and contribute to the uncertainty in the NOAEL/LOAEL.

3.5.4. Summary of uncertainties

In **Table 45**, a summary of the uncertainty evaluation is presented, highlighting the main sources of uncertainty and indicating an estimate of whether the respective source of uncertainty might have led to an over- or underestimation of the exposure or the resulting risk.

Table 45. Summary of qualitative evaluation of the impact of uncertainties on the risk assessment of acute exposure to potato GAs in food.

Sources of uncertainty	Direction [(a)]
Occurrence	
No GA levels determined in potatoes of specific cultivars grown at different locations and conditions; variability between countries and years	+/-
In case of no information it was assumed that potatoes were analysed as unpeeled	-
Estimated reduction of potato GAs concentration to account for loss during food processing (peeling and heat treatment)	+/-
Consumption data	

Different methodologies / representativeness / underreporting / misreporting / no portion size standard	+/-
Conversion of food consumed to raw primary commodity	+/-
Hazard identification and characterisation	
Uncertainty in the exposure levels of GAs estimated from case reports	+/-
Uncertainties on the shape of the dose-response relationship due to limited data available in humans	+/-
Putative interactions on the local irritating effect of potato GA with saponins	+
Possible interindividual differences in humans with regards to gastrointestinal symptoms	+/-
Non-specific symptoms of intoxications in humans not associated with potato GAs intake, resulting in under-reporting and uncertainties for derivation of LOAEL/NOAEL	+/-

(a):+=uncertainty with potential to cause over-estimation of exposure/risk; - =uncertainty with potential to cause under-estimation of exposure/risk.

The CONTAM Panel considered that the impact of the uncertainties on the risk assessment of acute exposure to potato GAs in food is moderate. Overall, the identified uncertainties may either cause an over- or underestimation of the risk.

4. Conclusions

4.1. Hazard identification and characterisation

4.1.1. Toxicokinetics

- The potato glycoalkaloids (GAs), α-solanine and α-chaconine, show a relatively low oral bioavailability in experimental animals, with differences between species. Hamsters exhibit higher absorption and slower excretion rates for both substances when compared to rats. Due to the limited information, the metabolic profiles of potato GAs in experimental animals could not be characterised.
- In humans, α-solanine and α-chaconine are systemically absorbed following ingestion with relatively long serum half-lives reported suggesting a possible accumulation.
- Levels of solanidine were regularly detected in the blood of human volunteers in several studies suggesting hydrolysis of GAs. The blood clearance of solanidine appears to be slow.
- No further information is available on metabolism and excretion of potato GAs in humans.
- Toxicokinetic data on tomato and aubergine GAs and their aglycones could not be identified, neither for experimental animals nor for humans.
- Information on the toxicokinetics of GAs in farm and companion animals was limited to ruminants, for which the data suggest an extensive conversion of α-solanine and α-chaconine to aglycones in rumen, and a low potential of solanidine to transfer into cows' milk.

4.1.2. Toxicity in experimental animals

- In acute, subacute and subchronic studies similar/identical doses of α-solanine and α-chaconine elicited comparable symptoms in several experimental animal species. This indicates a similar potency of α-solanine and α-chaconine with regard to toxicity.

- In repeated oral dose studies in rodents, reduced body and liver weight were the most common effects for potato GAs, while the aglycone solanidine increased absolute and relative liver weight.
- The tomato GA, α-tomatine, and the aglycone, tomatidine, showed no adverse effects in rodents.
- In mice, the aubergine GA, α-solasonine, increased the body weight gain. However, its aglycone, solasodine, lowered the body weight and caused gastric gland degeneration and liver toxicity, indicating different effects of the aglycone when compared to the GA.
- Malformations in the central nervous system occurred in hamsters at relatively high doses of potato GAs and the aglycone, solanidine, when applied for only one day or for a short, very restricted time period during gestation. No no-observed-adverse-effect level (NOAEL) or lowest-observed-adverse-effect level (LOAEL) could be identified from these studies.
- Reduced postnatal survival of pups due to insufficient milk production was reported when pregnant Holtzman rats had been exposed to α-solanine.
- Exposure of male dogs to the aubergine aglycone solasodine decreased epididymal weight and cauda epididymal epithelial height. Also an epididymal lumen depleted of sperm was reported. Similar effects of solasodine were observed in Rhesus monkeys.
- From the limited number of studies available, there was no evidence for genotoxicity of the potato GAs, α-solanine and α-chaconine, and the aglycone, solanidine, as well as the aubergine GA α-solamargine.
- No long-term chronic toxicity/carcinogenicity study for potato, tomato or aubergine GAs or for the respective aglycones could be identified.

4.1.3. Observations in humans

- Acute toxic effects following ingestion of potato GAs include gastrointestinal symptoms of varying severity such as vomiting, diarrhoea and abdominal pain, which may occur from a total potato GAs (potato TGA) intake of 1 mg/kg bw or more.
- In severe cases, paralysis, respiratory insufficiency, cardiac failure, coma and death have been reported. Doses in the range of 3–6 mg potato TGAs/kg bw are considered to be potentially lethal for humans.

4.1.4. Adverse effects in farm animals, horses and companion animals

- No data on the potential adverse effects of potato GAs in horses, companion animals (cats and dogs) or fur animals were identified.
- Due to an insufficient database on the adverse effects of GAs in ruminants, pigs, poultry, rabbits and fish, an acute reference dose could not be derived.

4.1.5. Mode of action

- Adverse effects of GAs may be due to their ability to complex with membrane 3β-hydroxy sterols, thereby causing disruption and loss of integrity of cell membranes. After oral exposure, these effects may affect the mucosa of the gastrointestinal tract and cause the symptoms observed in intoxicated humans, such as nausea, vomiting and diarrhoea.
- GAs inhibit acetylcholinesterase (AChE) and serum butyrylcholinesterase (BuChE) by a reversible, competitive mode of action. The relative potency of inhibition of α-solanine and α-chaconine

appear to be similar. The aglycones exert weak or no inhibitory effects. The excess of acetylcholine at the neuronal and neuromuscular junctions upon inhibition of the enzymes might also contribute to the symptoms described for intoxications with GAs.

4.1.6. Margin of exposure (MOE) approach

- The CONTAM Panel considered that the use of rodent data on acute toxicity was not appropriate to establish a reference point for acute exposure to potato GAs in humans. The CONTAM Panel selected the LOAEL of 1 mg potato TGA/kg bw per day as the reference point for the acute risk characterisation, based on human data from case reports, outbreaks and studies in volunteers.
- The available data on acute toxicity were considered insufficient to establish a health-based guidance value (HBGV). Instead, the MOE approach was used to assess a possible health concern from acute exposure to potato TGAs via food.
- The main symptoms following acute exposure to potato GAs may be mainly due to local irritation of the gastrointestinal mucosa rather than inhibition of AChE activity. Therefore, the CONTAM Panel considered that the possible interindividual variability in toxicodynamics is more relevant than the interindividual variability in toxicokinetics.
- An MOE higher than 10 indicates that there is no health concern. This MOE of 10 takes into account the extrapolation from a LOAEL to a NOAEL (a factor of 3) and the interindividual variability in toxicodynamics (a factor of 3.2).
- The experimental data for repeated dose toxicity are not sufficient to identify a reference point for chronic exposure to potato GAs. In humans, no evidence of health problems associated with repeated or long-term intake of GAs via potatoes has been identified.
- No data for determining a reference point for tomato or aubergine GAs or aglycones are available.

4.2. Occurrence and exposure

4.2.1. Food

- Occurrence data were only available for α-solanine and α-chaconine, and mostly for 'Main crop potatoes' and 'New potatoes'. Few data were available for processed food categories. No occurrence data were available for GAs and their aglycones in tomato and aubergine.
- The mean UB occurrence for 'Main crop potatoes' and 'New potatoes' was 52.0 mg/kg and the P95 occurrence was 117.0 mg/kg. The minimum and maximum reported concentrations were 1.1 mg/kg and 550.3 mg/kg, respectively.
- The CONTAM Panel decided to use the occurrence data in the raw primary commodities due to lack of available data for processed foods.
- The CONTAM Panel assessed the acute dietary exposure to potato TGAs (sum of α-solanine and α-chaconine) using a probabilistic approach.
- Reduction factors were estimated for the major food processing steps, comprising peeling (factor between 0.25 and 0.75) and heat processing of potatoes (factors between 0.2 and 0.9 for frying and deep frying, and between 0.05 and 0.65 for all other cooking methods), and these were applied to the occurrence data. It was assumed that 90% of the potatoes are consumed as peeled.

- The mean upper bound exposure to potato TGAs across surveys ranged from 26.0 µg/kg bw per day in adults to 193.4 µg/kg bw per day in toddlers. The 95th percentile exposure ranged from 88.2 µg/kg bw per day in adults to 617.9 µg/kg bw per day in toddlers (up to 1,057.9 µg/kg bw per day in the upper limit of the 95% confidence interval).

4.2.2. Feed

- Potatoes and by-products thereof can be used as feed but data on potato GAs in feed were insufficient to perform an exposure assessment.

4.3. Risk characterisation

4.3.1. Human health risk characterisation

- For potato GAs, the MOEs for the younger age groups indicate a health concern for the food consumption surveys with the highest mean exposure, as well as for the P95 exposure in all surveys.

- For adult age groups, the MOEs indicate a health concern only for the food consumption surveys with the highest P95 exposures.

- The Panel calculated the mean percentage of days with potato consumption across surveys per age group on which the potato TGA intake may be below the MOE of 10. The highest number of survey days was estimated for toddlers (56%) followed by children (50%). For the other age groups the estimated intake was calculated to be below the MOE of 10 in 23 to 40% of the survey days.

- For tomato and aubergine GAs, the risk to human health could not be characterised due to the lack of occurrence data in food and the limited information on the adverse effects in experimental animals and humans.

4.3.2. Farm animals, horses and companion animal health risk characterisation

- No risk characterisation could be performed due to insufficient occurrence data of GAs for feed and the lack of, or limited, data on the potential adverse effects of GAs in farm animals, horses and companion animals.

5. Recommendations

The following needs have been identified to improve the risk assessment for humans and reduce the uncertainties:

- Research on the occurrence of GAs and their aglycones and other potentially toxicologically relevant secondary plant metabolites in the potato cultivars available on the market and on new potato cultivars resulting from breeding experiments.

- Occurrence data on GAs and their aglycones in potato processed products, including foods for infants.

- Occurrence data on GAs and their aglycones in tomato and aubergine and products thereof.

- Data on the toxicokinetics of potato, tomato and aubergine GAs and aglycones in experimental animals and humans.

- Data on repeated dose toxicity, including reproductive and developmental toxicity of potato, tomato and aubergine GAs and aglycones in experimental animals.
- Studies in humans linking dietary exposure, biomarkers of exposure and adverse effects.

The following needs have been identified to improve the risk assessment for farm animals, horses and companion animals and reduce the uncertainties:

- Occurrence data on potato GAs and their aglycones in feed.
- Studies on the kinetics and the potential adverse effects from feed material containing GAs of potato GAs in farm animals, horses and companion animals.

Documentation provided to EFSA

1. Sicherheitsbewertung gentechnisch veränderter Lebensmittel: Glykoalkaloide in transgenen Kartoffeln. PhD thesis by Gessner Gerhard. Technisched Universität Műnchen. 2000. Submitted to EFSA by the German Federation for Food Law and Food Science (BLL) following the Consultation with stakeholders (see **Section 2.2.1**).

2. Occurrence data of GAs in 'fresh potato', 'fresh potato with skin for industrial use' and 'potato crisps'. Data submitted by the European Snasks Association (ESA) following the Consultation with stakeholders (see **Section 2.2.1**). The occurrence data was not further submitted following the requirements of the EFSA Guidance on Standard Sample Description for Food and Feed (EFSA, 2010a), and they could not be used for the current dietary exposure assessment.

3. Comments on GAs in potato and potato products and occurrence data in potatoes and potato products such as 'croquettes', 'French fries', 'potato flakes', 'potato pancakes' and 'potato wedges'. Data submitted by the European Potato Processors' Association (EUPPA) following the Consultation with stakeholders (see **Section 2.2.1**). The occurrence data was not further submitted following the requirements of the EFSA Guidance on Standard Sample Description for Food and Feed (EFSA, 2010a), and they could not be used for the current dietary exposure assessment.

4. Comments on GAs in potato and potato products. Data submitted by the European Potato Trade Associations (Europatat) following the Consultation with stakeholders (see **Section 2.2.1**).

5. Occurrence data of GAs in potato and potato products such as 'potato dietary fibre', 'potato proteins', 'potato pulp', 'potato juice', 'potato starch' and 'potatoes used for starch production'. Data submitted by the European Starch Industry Association following the Consultation with stakeholders (see **Section 2.2.1** and **3.2.1**).

References

Abbasi KS, Masud T, Qayyum A, Ahmad A, Mehmood A Bibi Y and Sher A, 2016. Photo-induced changes in quality attributes of potato tubers during storage. Journal of Applied Botany and Food Quality, 89, 315-321.

Abdel-Gawad A, Omer EA and Ahmed SS, 1993. Effect of storage on the glycoalkaloids and carbohydrate contents in different potato varieties. Egyptian Journal of Horticulture, 19, 81-92.

Abreu P, Relva A, Matthew S, Gomes Z and Morais Z, 2007. High-performance liquid chromatographic determination of glycoalkaloids in potatoes from conventional, integrated, and organic crop systems. Food Control, 18, 40-44.

Afify AEMR, El-Beltagi HS, Aly AA and El-Ansary AE,2012. The impact of γ-irradiation, essential oils and iodine on biochemical components and metabolism of potato tubers during storage. Notulae Botanicae Horti Agrobotanici, 40, 129-139.

Ahmed SS and Műller K, 1978. Effect of wond-damages on the glycol-alkaloid content in potato tubers and chips. Lebenmittel Wiss und Technol, 11, 144-146.

Álvarez MA, Rodríguez Talou J, Paniego NB and Giulietti AM, 1994. Solasodine production in transformed organ cultures (roots and shoots) of *Solanum eleagnifolium* Cav. Biotechnology Letters, 16, 393-396.

Al Chami L, Mendez R, Chataing B, O'Callaghan J, Usubillaga A and LaCruz L, 2003. Toxicological effects of alpha-solamargine in experimental animals. Phytotherapy Research, 17, 254-258. doi:10.1002/ptr.1122

Al Sinani SSS and Eltayeb EA, 2017. The steroidal glycoalkaloids solamargine and solasonine in Solanum plants. South African Journal of Botany, 112, 253-269.

Alberti B, 1932. Zum Nachweis von Solanin. Zeitschrift fuer Untersuchung der Lebensmittel, 64, 260-262.

Aldous CN, Sharma RP and Salunkhe DK, 1980. Effects of α-chaconine on brain biogenic amines, electroencephalogram, cardiac rates and respiratory response in rats. Journal of Food Safety, 2, 20-33.

Alfa J and Heyl E, 1923. Kartoffeln 1922-er ernte mit ausserordentlich hohem solaningehalt. Zeitschrift für Untersuchung der Nahrungs- und Genußmittel, sowie der Gebrauchsgegenstände, 46, 306–309.

Almeida AE, Cardoso CRP, Almeida DV, Moreira RRD, Silva M and Varanda EA, 2010. Mutagenic activity of glycoalkaloids from *Solanum palinacanthum* Dunal (Solanacea) found in the Brazilian cerrado. Latin American Journal of Pharmacy, 29, 122-126.

Allen JR, Marlar Dr RJ, Chesney CF, Helgeson JP, Kelman A, Weckel KG, Traisman E and White Jr JW, 1977. Teratogenicity Studies on Late Blighted Potatoes in Nonhuman Primates (Macaca rnulatta and Saguinus labiatus). Experimental Teratology, 15, 17-23.

Alozie SO, Sharma RP and Salunkhe DK, 1978. Inhibition of rat cholinesterase isoenzymes in vitro and in vivo by the potato alkaloid, α-chaconine. Journal of Food Biochemistry, 2, 259-276.

Alozie SO, Sharma RP and Salunkhe DK, 1979a. Physiological disposition, subcellular distribution and tissue binding of [3H]-α-chaconine. Journal of Food Safety, 1, 257-273.

Alozie SO, Sharma RP and Salunkhe DK, 1979b. Excretion of alpha-chaconine-3H, a steroidal glycoalkaloid from *Solanum tuberosum* L. and its metabolites in hamsters. Pharmacological Research Communications, 11, 483-490.

Amer FS, Reddivari L, Madiwale GP, Stone M, Holm DG and Vanamala J, 2014. Effect of Genotype and Storage on Glycoalkaloid and Acrylamide Content and Sensory Attributes of Potato Chips. American Journal of Potato Research, 91, 632-641.

Anses (Agence Nationale de Sécurité Sanitaire de l'Alimentation, de l'Environnment et du Travail), 2014. Appui scientifique et technique de l'Anses, Saisine n°2014-SA-0115 Note d'appui scientifique et technique de l'Agence nationale de sécurité sanitaire de l'alimentation, de l'environnement et du travail relatif aux intoxications liées à la consommation de tomates cerise en provenance du Maroc. Maisons-Alfort. 13 pp. Available online: https://www.anses.fr/fr/system/files/ERCA2014sa0115.pdf

Arena MP, Govers C, Lotti C, Ricciardi L, Wichers HJ and Mes JJ, 2018. The effect of tomatine on gene expression and cell monolayer integrity in Caco-2. Molecules, 23, 644.

AOAC (Association of Official Agricultural Chemists), 2000. AOAC official method 997.13: Glycoalkaloids (a-Solanine and a-Chaconine) in potato tubers. 3.

Azim A, Shaikh HA and Ahmad R, 1982. Effect of feeding greened potatoes on different visceral organs and blood plasma of rabbits. Journal of the Science of Food and Agriculture, 33, 1275-1279.

Azim A, Shaikh HA and Ahmad R, 1983. Toxic effects of high glycoalkaloid feeding on the protein digestibility and growth of rabbits. Journal of Pharmacy, 2, 15-24.

Azim A, Shaikh HA and Ahmad R, 1984. Toxic effects of high glycoalkaloid feeding on the RBC counts and hemoglobin concentration of rabbit blood. Journal of Pharmacy, 3, 43-49.

Bagheri M, Bushehri AAS, Hassandokht and Naghavi MR, 2017. Evaluation of solasonine content and expression patterns of SGT1 gene in different tissues of two Iranian Eggplant (*Solanum melongena* L.) genotypes. Food Technology and Biotechnology, 55, 236-242.

Bajaj KL, Kaur G and Chadha ML, 1979. Glycoalkaloid content and other chemical constituents of the fruits of some egg plant (*Solanum melongena*, L.) varieties. Journal of Plant Foods, 3, 163-168.

Baker DC, Keeler RF and Gaffield W, 1989. Pathology in hamsters administered Solanum plant species that contain steroidal alkaloids. Toxicon, 27, 1331-1337.

Baker DC, Keeler RF and Gaffield WP, 1998. Mechanism of death in Syrian hamstyers gavaged potto sprout material. Toxicologic Pathology, 16, 333-339.

Baldina S, Picarella ME, Troise AD, Pucci A, Ruggieri V, Ferracane R, Barone A, Fogliano V and Mazzucato A, 2016. Metabolite profiling of Italian tomato landrces with different fruit types. Frontiers in Plant Science, 7, 664.

Barceloux DG, 2009. Potatoes, tomatoes, and solanine toxicity (*Solanum tuberosum* L., Solanum [lycopersicum L.).]. Disease-a-month: DM, 55, 391-402. doi:10.1016/j.disamonth.2009.03.009

Bártová V, Diviš J, Bárta J, Brabcová A and Švajnerova M, 2013. Variation of nitrogenous components in potato (Solanum tuberosum L.) tubers produced under organic and conventional crop management. Euroepan Journal of Agronomy, 49, 20-31.

Bejarano L, mingolet E, Devaux A, Espinola N, Carrasaco E and Larondelle Y, 2000. Glycoalkaloids in potato tubers: the effect of variety and drought stress on thea-solanine and a-chaconine contents of potatoes. Journal of the Science of Food and Agriculture, 80, 2096-2100.

Bergers WWA, 1981. Investigation of the contents of phenolic and alkaloidal compounds of gamma irradiated potatoes during storage. Food Chemistry, 7, 47-61.

Beausoleil JL, Spergel JM and Pawlowski NA, 2001. Anaphylaxis to raw potato. Annals of Allergy, Asthma and Immunology, 86, 68-70. doi:10.1016/s1081-1206(10)62359-9

Beier RC and Nigg HN, 1992. Natural Toxicants in Foods. Phytochemical Resources for Medicine and Agriculture, 247-367.

Benilova IV, Soldatkin AP, Martelet C and Jaffrezic-Renault N, 2006. Nonfaradaic impedance probing of potato glycoalkaloids interaction with butyrylcholinesterase immobilized onto gold electrode. Electroanalysis, 18, 1950-1956. doi: 10.1002/elan.200603601

BfR (Bundesinstitut fűr Risikobewertung), 2018a. BfR Opinion No 010/2018 of 23 April 2018. Table potatoes should contain low levels of glycoalkaloids (solanine) (Abstract in English). Available at: http://www.bfr.bund.de/cm/349/table-potatoes-should-contain-low-levels-of-glycoalkaloids-solanine.pdf

BfR (Bundesinstitut fűr Risikobewertung), 2018b. Stellungnahme Nr. 010/2018 des BfR vom 23. April 2018. Speisekartoffeln sollten niedrige Gehalte an Glykoalkaloiden (Solanin) enthalten (in German). Available at: http://www.bfr.bund.de/cm/343/speisekartoffeln-sollten-niedrige-gehalte-an-glykoalkaloiden-solanin-enthalten.pdf

Bhat R and Pallyath G, 2016. Fruits of tropical climates: dietary importance and health benefits, in 'Encyclopedia of food and health' (Caballero B, Finglas PM and Toldra F, eds.). Elsevier, Amsterdam. P. 144.
Bintcliffe EJB, Clydesdale A and Draper SR, 1982. Effect of genotype, site and season on the glycoalkaloid content of potato tubers. Journal of the National Institute of Agricultural Botanic, 16, 86-91.
Bitsch R et al 1974. Quality retention of potatoes during transport and storage as influenced by various packaging materials. II. Effects of various packaging materials on solanine formation. Nahrung, 18, 395-408.
Blankemeyer JT, Atherton R and Friedman M, 1995. Effect of potato glycoalkaloids α-chaconine and α-solanine on sodium active transport in frog skin. Journal of Agricultural and Food Chemistry, 43, 636-639.
Blankemeyer JT, White JB, Stringer BK and Friedman M, 1997. Effect of α-tomatine and tomatidine on membrane potential of frog embryos and active transport of ions in frog skin. Food and Chemical Toxicology, 35, 639-646.
Bodart P, Kabengera C, Noirfalise A, Hubert P and Angenot L, 2000. Determination of α-solanine and α-chaconine in potatoes by high-performance thin-layer chromatography/densitometry. Journal of AOAC International, 83, 1468-1473.
Bolin FM, 1962. Green potatoes can kill sheep. North Dakota Farm Research, 22(7), 15.
Bömer A and Mattis H, 1923. Über hohe Solaningehalte bei Kartoffeln. Zeitschrift für Untersuchung der Nahrungs- und Genussmittel sowie der Gebrauchsgegenstaende, 45, 288-291.
Bömer A and Mattis H, 1924. Der Solaningehalt in der Kartoffel (The solanine content of potatoes). Zeitschrift für Untersuchung der Nahrungs- und Genussmittel sowie der Gebrauchsgegenstaende, 47, 97-127.
Branzell C and Hellenäs K-E, 1999. Undvik höga halter glykoalkaloider: Välj potatis av rätt sort! Vår Föda, 3/99, p 4-8.
Brimijoin S, Chen VP, Pang YP, Geng L and Gao Y, 2016. Physiological roles for butyrylcholinesterase: A BChE-ghrelin axis. Chemico-Biological Interactions, 259, 271-275.
Brimijoin S, Gao Y, Geng L and Chen VP, 2018. Treating Cocaine Addiction, Obesity, and Emotional Disorders by Viral Gene Transfer of Butyrylcholinesterase. Frontiers in Pharmacology, 9, 112.
Brown D and Keeler RF, 1978. Structure-activity relation of steroid teratogens. 3. Solanidan epimers. Journal of Agricultural and Food Chemistry, 26, 566-569.
Bushway RJ and Ponnampalam R, 1981. α-Chaconine and α-solanine content of potato products and their stability during several modes of cooking. Journal of Agricultural and Food Chemistry, 29, 814-817.
Bushway RJ, Bureau JL and McGann DF, 1983. Alpha-chaconine and alpha-solanine content of potato peels and potato peel products. Journal of Food Science, 8, 84-86.
Bushway RJ, McGann DF and Bushway AA, 1984. Gas chromatographic method for the determination of solanidine and its application to a study of feed-milk transfer in the cow. Journal of Agricultural and Food Chemistry, 32, 548-551.
Bushway RJ, Perkins LB, Paradis LR and Vanderpan S, 1994. High-Performance Liquid Chromatographic Determination of the Tomato Glycoalkaloid, Tomatine, in Green and Red Tomatoes. Journal of Agricultural and Food Chemistry, 42, 2824-2829.
Calf OW, Huber H, Peters JL, Weinhold A and van Dam NM, 2018. Glycoalkaloid composition explains variation in slug resistance in Solanum dulcamara. Oecologia, 187, 495-506.
Canada Diseases Weekly Report, 1984. Solanine food poisoning associated with a school lunch program - Alberta. May 1984. Canada Diseases Weekly Report, Health and Welfare Canada, 71, 10-18. Available at: http://publications.gc.ca/collections/collection_2016/aspc-phac/H12-21-1-10-18.pdf
Caprioli G, Logrippo S, Cahill MG and James KJ, 2014. High-performance liquid chromatography LTQ-Orbitrap mass spectrometry method for tomatidine and non-target metabolites quantification in organic and normal tomatoes, International Journal of Food Sciences and Nutrition, 65, 942-947.
Caruso G, Pokluda R, Sekara A, Kalisz A, Jezdinsky, A, Kopta T and Grabowska A, 2017. Agricultural practices, biology and quality of eggplant cultivated in Central Europe. A review. Horticultural Science, 44, 201-212.
Cavlovic P, Mankotia M, Pantazopoulos P and Scott PM, 2003. Liquid chromatographic determination of alpha-solasonine in frozen green peas as an indicator of the presence of nightshade berries. Journal of AOAC International, 86, 759-763.
Cayen MN, 1971. Effect of dietary tomatine on cholesterol metabolism in the rat. Journal of Lipid Research, 12, 482-490.
Ceriola F, Portis E, Toppino L, Barchi L, Acciarri N, Ciriaci T, Sala T, Rotino GL and Lankri S, 2013. The population structure and diversity of eggplant from Asia and the Mediterranean Basin. PLoS ONE 8: e73702.
Chaube S and Swinyard CA, 1976. Teratological and toxicological studies of alkaloidal and phenolic compounds from *Solanum tuberosum* L. Toxicology and Applied Pharmacology, 36, 227-237.
Chaube S, Swinyard CA and Daines RH, 1973. Failure to induce malformations in fetal rats by feeding blighted potatoes to their mothers. The Lancet, 1 (7798): 329-330.
Chen X, Ding Y and Kan J, 2018. Changes in the content and influence factors of α-solanine in potato during storage. Emirates Journal of Food and Agriculture, 30, 10-16.
Choi E-M, 2007. Suppressive effects of potato (*Solanum tuberlosum*) on type II [conagen-induced arthritis in DBA/1J mice]. Food Science and Biotechnology, 16, 43-48.
Choi E and Koo S, 2005. Anti-nociceptive and anti-inflammatory effects of the ethanolic extract [of potato (*Solanum tuberosum*). Food and Agricultural Immunology, 16, 29-39. doi:10.1080/09540100500064320

4305 Choi EM, Lee KH and Koo SJ, 2005. Effects of ethanol extract of potato (*Solanum tuberlosum*) on Freund's complete
4306 adjuvant-induced model of chronic arthritis in mice. Food Science and Biotechnology, 14, 228-232.
4307 Choi S-H, Lee SH, Kim HJ, Lee IS, Kozukue N, Levin CE and Friedman M, 2010. Changes in free amino acid, phenolic,
4308 chlorophyll, carotenoid, and glycoalkaloid contents in tomatoes during 11 stages of growth and inhibition of
4309 cervical and lung human cancer cells by green tomato extracts. Journal of gricultural and Food Chemistry, 58,
4310 7547-7556.
4311 Cichon MJ, Riedl KM and Schwartz SJ, 2017. A metabolomic evaluation of the phytochemical composition of tomato
4312 juices being used in human clinical trials. Food Chemistry, 28, 270-78.
4313 Cieslik E and Praznik W, 1998. Changes of glycoalkaloid content in potato tubers of selected varieties during
4314 vegetation and storage. Polish Journal of Food and Nutrition Sciences, 7, 417-422.
4315 Claringbold, 1982. Kinetics and retention of solanidine in man. Xenobiotica, 12, 293-302.
4316 Clarke G, Fall CHD, Lincoln DW and Merrick LP, 1978. Effects of cholinoceptor antagonists on suckling-induced and
4317 experimentally evoked release of oxytocin. British Journal of Pharmacology, 63, 519-527.
4318 Clement E and Verbist JF, 1980. The determination of solanine in *Solanum tuberosum* L. tubers: a comparative
4319 study of nine colorimetric methods. Lebensmittel-Wissenschaft und -Technologie, 13, 202-206.
4320 Čolović MB, Krstić DZ, Lazarević-Pašti TD, Bondžić AM and Vasić VM, 2013. Acetylcholinesterase Inhibitors:
4321 Pharmacology and Toxicology. Current Neuropharmacology, 2013, 11, 315-335.
4322 Conner HW, 1937. Effect of light on solanine synthesis in the potato tuber. Plant Physiology, 12, 79-98.
4323 Cooper MR and Johnson AW, 1984. Poisonous plants in Britain and their effects on Animals and Man. Reference
4324 Book 161, Ministry of Agriculture, Fisheries and Food. HMSO, UK.
4325 Cortical, 1888. Accidents d'intoxication survenus au 139c d'infanterie à Lyon, le 11 et 12 Juillet 1888, et imputés à
4326 la consommation de pommes de terre de mauvaise qualité. Archives de Medicine et de Pharmacie militaire,
4327 4, 3. As cited by Rothe (1919).
4328 Crawshaw R, 2001. Co-product feeds: animal feeds from the food and drinks industries Wiley-Blackwell, Nottingham
4329 University, pp.
4330 Dale MFB, Griffiths DW, Bain H and Goodman BA, 1997. The effect of gamma irradiation on glycoalkaloid and
4331 chlorophyll synthesis in seven potato cultivars. Journal of the Science of Food and Agriculture, 75, 141-147.
4332 Dale MFB, Griffiths DW and Bain HF, 1998. Effect of bruising on the total glycoalkaloid and chlorogenic acid content
4333 of potato (Solanum tuberosum) tubers of five cultivars. Journal of the Science of Food and Agriculture, 77,
4334 499-505.
4335 Dalvi RR, 1985. Comparative assessment of the effect of solanine administered orally and intraperitoneally on
4336 hepatic dysfunction in male rats. Nihon Juigaku Zasshi. The Japanese Journal of Veterinary Science, 47, 657-
4337 659.
4338 Damon SR, 1928. Food Infections and Food Intoxications. Chapter XI Potato poisoning. Williams & Wilkins,
4339 Baltimore, Maryland, 153-7.
4340 Dao L and Friedman M, 1994. Chlorophyll, Chlorogenic Acid, Glycoalkaloid, and Protease Inhibitor Content of Fresh
4341 and Green Potatoes. Journal of Agricultural and Food Chemistry, 42, 633-639.
4342 Dastmalchi K, Perez Rodriguez M, Lin J, Yoo B and Stark RE, 2019. Temporal resistance of potato tubers:
4343 Antibacterial assays and metabolite profiling of wound-healing tissue extracts from contrasting cultivars.
4344 Phytochemistry, 159, 75-89.
4345 Davies AMC and Blincow PJ, 1984. Glycoalkaloid content of potatoes and potato products sold in the UK. Journal
4346 of the Science of Food and Agriculture, 35, 553-557.
4347 Dean BB, 1994. Managing the potato production system. Food Products Press, New York. ISBN: 1560220252. p
4348 56.
4349 Deußer H, Guignard C, Hoffmann L and Evers D, 2012. Polyphenol and glycoalkaloid contents in potato cultivars
4350 grown in Luxembourg. Food Chemistry, 135, 2814–2824.
4351 Diehl JF and Wedler A, 1977. Traditional versus "organic" farming. Comparative studies concerning the quality of
4352 the harvested products. Berichte der Bundesforschungsanstalt fuer Ernaehrung(1): 144 pp.
4353 Distl M and Wink M, 2009. Identification and quantification of steroidal alkaloids from wild tuber-bearing Solanum
4354 species by HPLC and LC-ESI-Ms. Potato Research, 52, 79-104.
4355 Dixit VP, Gupta RS and Gupta S, 1989. Antifertility plant products: testicular cell population dynamics following
4356 solasodine ($C_{27}H_{43}O_2N$) administration in Rresus monkey (*Macaca mulatta*). Andrologia, 21, 542-546.
4357 Dixit VP and Gupta RS, 1982. Antispermatogenic, antiandrogenic properties of solasodine ($C_{27}H_{43}O_2N$) obtained
4358 from Solanum xanthocarpum berries on the male genital tract of dog (*Cannis familiars*): A histophysiological
4359 approach. International Journal of Andrology, 5, 295–307.
4360 Dolan LC, Matulka RA and Burdock GA, 2010. Naturally occurring food toxins. Toxins, 2, 2289-2332.
4361 doi:10.3390/toxins2092289
4362 Dornow Food Technology, 2019. Applications - steam peeling mechanical peeling. What peeling method is used
4363 where? Accessed on 4 February 2019 at http://www.dornow.de/de/englisch/Q-Prospekte/Q104e5.pdf
4364 Duysen EG, Stribley JA, Fry DL, Hinrichs SH and Lockridge O, 2002. Rescue of the acetylcholinesterase knockout
4365 mouse by feeding a liquid diet; phenotype of the adult acetylcholinesterase deficient mouse. Developmental
4366 Brain Research, 137, 43-54.

4367 Eanes RC, Tek N, Kirsoy O, Frary A, Doganlar S and Almeida AE, 2008. Development of Practical HPLC Methods for
4368 the Separation and Determination of Eggplant Steroidal Glycoalkaloids and their Aglycones. Journal of Liquid
4369 Chromatography and Related Technologies, 31, 984-1000.
4370 Edwards EJ and Cobb AH, 1997. Effect of temperature on glycoalkaloid and chlorophyll accumulation in potatoes
4371 (Solanum tuberosum L cv King Edward) stored at low photon flux density, including preliminary modeling
4372 using an artificial neural network. Journal of Agricultural and Food Chemistry, 45, 1032-1038.
4373 Edwards EJ and Cobb AH, 1999. The effect of prior storage on the potential of potato tubers (Solanum tuberosum
4374 L) to accumulate glycoalkaloids and chlorophylls during light exposure, including artificial neural network
4375 modelling. Journal of the Science of Food and Agriculture, 79, 1289-1297.
4376 Edwards EJ, Saint RE and Cobb AH, 1998. Is there a link between greening and light-enhanced glycoalkaloid
4377 accumulation in potato (*Solanum tuberosum* L) tubers? Journal of the Science of Food and Agriculture, 76,
4378 327-333.
4379 EFSA (European Food Safety Authority), 2005. Opinion of the Scientific Committee on a request from EFSA related
4380 to a harmonised approach for risk assessment of substances which are both genotoxic and carcinogenic. EFSA
4381 Journal 2005;282, 1–31. https://doi.org/10.2903/j.efsa.2005.282
4382 EFSA (European Food Safety Authority), 2007. Guidance of the Scientific Committee on a request from EFSA related
4383 to Uncertainties in Dietary Exposure Assessment. EFSA Journal 2007;4(12):438, 54 pp.
4384 https://doi.org/10.2903/j.efsa.2007.438
4385 EFSA (European Food Safety Authority), 2009. Guidance of the Scientific Committee on transparency in the scientific
4386 aspects of risk assessments carried out by EFSA. Part 2: general principles. EFSA Journal 2009;1051, 1–22.
4387 https://doi.org/10.2903/j.efsa.2009.1051
4388 EFSA (European Food Safety Authority), 2010a. Standard sample description for food and feed. EFSA Journal
4389 2010;8(1):1457, 54 pp. https://doi.org/10.2903/j.efsa.2010.1457
4390 EFSA (European Food Safety Authority), 2010b. Management of left-censored data in dietary exposure assessment
4391 of chemical substances. EFSA Journal 2010;8(3):1557, 96 pp. https://doi.org/10.2903/j.efsa.2010.1557
4392 EFSA (European Food Safety Authority), 2011a. Use of the EFSA Comprehensive European Food Consumption
4393 Database in Intakes Assessment. EFSA Journal 2011;9(3):2097, 34 pp.
4394 https://doi.org/10.2903/j.efsa.2011.2097
4395 EFSA (European Food Safety Authority), 2011b. Report on the development of a food classification and description
4396 system for exposure assessment and guidance on its implementation and use. EFSA Journal 2011;9(12):2489.
4397 [84 pp.] https://doi.org/10.2903/j.efsa.2011.2489
4398 EFSA (European Food Safety Authority), 2015. The food classification and description system FoodEx2 (revision 2).
4399 EFSA supporting publication 2015:EN-804. 90pp. https://doi.org/10.2903/sp.efsa.2015.EN-804
4400 EFSA (European Food Safety Authority), Dujardin B and Kirwan L, 2019.Technical report on the raw primary
4401 commodity (RPC) model: strengthening EFSA's capacity to assess dietary exposure at different levels of the
4402 food chain, from raw primary commodities to foods as consumed. EFSA supporting publication 2019:EN-1532.
4403 30pp. https://doi.org/10.2903/sp.efsa.2019.EN-1532
4404 EFSA Scientific Committee 2011. Scientific Opinion on genotoxicity testing strategies applicable to food and feed
4405 safety assessment. EFSA Journal 2011;9(9):2379, 69 pp. https://doi.org/10.2903/j.efsa.2011.2379
4406 EFSA Scientific Committee 2012a. Guidance on selected default values to be used by the EFSA Scientific Committee,
4407 Scientific Panels and Units in the absence of actual measured data. EFSA Journal 2012;10(3):2579, 32 pp.
4408 https://doi.org/10.2903/j.efsa.2012.2579
4409 EFSA Scientific Committee, 2012b. Scientific Opinion on Risk Assessment Terminology. EFSA Journal
4410 2012;10(5):2664, 43 pp. https://doi.org/10.2903/j.efsa.2012.2664
4411 EFSA Scientific Committee, Hardy A, Benford D, Halldorsson T, Jeger MJ, Knutsen KH, More S, Mortensen A, Naegeli
4412 H, Noteborn H, Ockleford C, Ricci A, Rychen G, Silano V, Solecki R, Turck D, Aerts M, Bodin L, Davis A, Edler
4413 L, Gundert-Remy U, Sand S, Slob W, Bottex B, Abrahantes JC, Marques DC, Kass G and Schlatter JR, 2017a.
4414 Update: use of the benchmark dose approach in risk assessment. EFSA Journal, 15, n/a-n/a.
4415 https://doi.org/10.2903/j.efsa.2017.465
4416 EFSA Scientific Committee, Hardy A, Benford D, Halldorsson T, Jeger MJ,Knutsen HK, More S, Naegeli H, Noteborn
4417 H, Ockleford C, Ricci A, Rychen G, Schlatter JR, Silano V,Solecki R, Turck D, Benfenati E, Chaudhry QM, Craig
4418 P, Frampton G, Greiner M, Hart A, Hogstrand C,Lambre C, Luttik R, Makowski D, Siani A, Wahlstroem H,
4419 Aguilera J, Dorne J-L, Fernandez Dumont A,Hempen M, Valtueña Martínez S, Martino L, Smeraldi C, Terron A,
4420 Georgiadis N and Younes M, 2017b. Scientific Opinion on the guidance on the use of the weight of evidence
4421 approach in scientific assessments. EFSA Journal, 15(8): 4971, 69 pp.
4422 https://doi.org/10.2903/j.efsa.2017.4971
4423 EFSA SC (EFSA Scientific Committee), Hardy A, Benford D, Halldorsson T, Jeger M, Knutsen HK, More S, Naegeli H,
4424 Noteborn H, Ockleford C, Ricci A, Rychen G, Silano V, Solecki R, Turck D, Younes M, Aquilina G, Crebelli R,
4425 Gurtler R, Hirsch-Ernst KI, Mosesso P, Nielsen E, van Benthem J, Carfi M, Georgiadis N, Maurici D, Parra Morte
4426 J and Schlatter J, 2017c. Scientific Opinion on the clarification of some aspects related to genotoxicity
4427 assessment. EFSA Journal 2017c;15(12):5113, 25 pp. https://doi.org/10.2903/j.efsa.2017.5113
4428 EFSA Scientific Committee, Hardy A, Benford D, Halldorsson T, Jeger MJ,Knutsen HK, More S, Naegeli H, Noteborn
4429 H, Ockleford C, Ricci A, Rychen G, Schlatter JR, Silano V,Solecki R, Turck D, Younes M, Bresson J-L, Griffin J,
4430 Hougaard Benekou S, van Loveren H, Luttik R,Messean A, Penninks A, Ru G, Stegeman JA, van der Werf W,

Westendorf J, Woutersen RA, Barizzone F, Bottex B, Lanzoni A, Georgiadis N and Alexander J, 2017d. Guidance on the assessment of the biologicalrelevance of data in scientific assessments. EFSA Journal 2017;15(8):4970, 73 pp. https://doi.org/10.2903/j.efsa.2017.4970

EFSA SC (EFSA Scientific Committee), 2018. Benford D,Halldorsson T, Jeger MJ, Knutsen HK, More S, Naegeli H, Noteborn H, Ockleford C, Ricci A, Rychen G,Schlatter JR, Silano V, Solecki R, Turck D, Younes M, Craig P, Hart A, Von Goetz N, Koutsoumanis K,Mortensen A, Ossendorp B, Martino L, Merten C, Mosbach-Schulz O and Hardy A, 2018. Guidance on Uncertainty Analysis in Scientific Assessments. EFSA Journal 2018;16(1):5123, 39 pp. https://doi.org/10.2903/j.efsa.2018.5123

Eldridge AC and Hockridge ME, 1983. High-performance liquid chromatographic separation of Eastern black nightshade (Solanum ptycanthum) glycoalkaloids. Journal of Agricultural and Food Chemistry, 31, 1218-1220.

Elkahoui S, Bartley GE, Yokoyama WH and Friedman M, 2018. Dietary Supplementation of Potato Peel Powders Prepared from Conventional and Organic Russet and Non-organic Gold and Red Potatoes Reduces Weight Gain in Mice on a High-Fat Diet. Journal of Agricultural and Food Chemistry, 66, 6064-6072.

El-Said SM, 2013. Removal of pharmacological undersirable compounds from potato tuber. Research and Reviews in BioSciences, 7, 129-135.

Eltayeb EA and Roddick JG, 1984. Changes in the Alkaloid Content of Developing Fruits of Tomato (Lycopersicon esculentum Mill.): I. Analyses of cultivars and mutants with different ripening characteristics. Journal of Experimental Botany, 35, 252-260.

Emmanuel S, Ignacimuthu S, Perumalsamy R and Amalraj T, 2006. Antiinflammatory activity of *Solanum trilobatum*. Fitoterapia, 77, 611-612.

Essers AJ, Alink GM, Speijers GJ, Alexander J, Bouwmeister PJ, van den Brandt PA, Ciere S, Gry J, Herrman J, Kuiper HA, Mortby E, Renwick AG, Shrimpton DH, Vainio H, Vittozzi L and Koeman JH, 1998. Food plant toxicants and safety Risk assessment and regulation of inherent toxicants in plant foods. Environmental Toxicology and Pharmacology, 5, 155-172.

Ewing WN, 2002. The Feeds Directory Branded Products Guide. Context, Ashby de la Zouch, UK.

FAOSTAT (Food and Agriculture Organization of the United Nations), 2016. FAOSTAT database on tomato production. Rome, Italy: FAO. Retrieved in 2016 from http://www.fao.org/faostat/en/#data

FAOSTAT (Food and Agriculture Organization of the United Nations), 2018a. FAOSTAT database on potato production. Rome, Italy: FAO. Retrieved in 2018 from http://www.fao.org/faostat/en/#data

FAOSTAT (Food and Agriculture Organization of the United Nations), 2018b. FAOSTAT database on aubergine production. Rome, Italy: FAO. Retrieved in 2018 from http://www.fao.org/faostat/en/#data

Faridnia F, Burritt DJ, Bremer PJ and Oey I, 2015. Innovative approach to determine the effect pf pulsed electric fields on the microstructure of whole potato tubers: Use of cell viability, microscopic images and ionic leakage measurements. Food Research International, 77, 556-564.

Fiems LO, De Boever JL, Vanacker JM and De Brabander DL, 2013. Effect of cull potatoes in the diet for finishing Belgian Blue double-muscled cows. Animal, 7, 93-100.

Finotti E, Bertone A and Vivanti V, 2006. Balance between nutrients and anti-nutrients in nine Italian potato cultivars. Food Chemistry, 99, 698–701.

Fitzpatrick TJ, McDermott JA and Osman SF, 1978. Evaluation of injured commercial potato samples for total glycoalkaloid content. Journal of Food Science, 43, 1617-1618.

Fontaine TD, Irving GW, Poole RM and Dooluttle SP, 1948. Isolation and partial characterization of crystalline tomatine, an antibiotic agent from the tomato plant. Archives of Biochemistry, 18, 467-475.

Friedman M, 1992. Composition and safety evaluation of potato berries, potato and tomato seeds, potatoes, and potato alkaloids. Food Safety Assessment, Chapter 35, pp 429-462.

Friedman M, 2002. Tomato glycoalkaloids: role in the plant and in the diet. Journal of Agricultural and Food Chemistry, 50, 5751-5780.

Friedman M, 2004. Analysis of biologically active compounds in potatoes (*Solanum tuberosum*), tomatoes (*Lycopersicon esculentum*), and jimson weed (*Datura stramonium*) seeds. Review. Journal of Chromatography A, 1054, 143-155.

Friedman M, 2006. Potato glycoalkaloids and metabolites: roles in the plant and in the diet. Journal of Agricultural and Food Chemistry, 54, 8655-8681.

Friedman M, 2015. Chemistry and anticarcinogenic mechanisms of glycoalkaloids produced by eggplants, potatoes, and tomatoes. Journal of Agricultural and Food Chemistry, 63, 3323-3337.

Friedman M and Dao L, 1992. Distribution of glycoalkaloids in potato plants and commercial potato products. Journal of Agricultural and Food Chemistry, 40, 419-423.

Friedman M and Henika PR, 1992. Absence of genotoxicity of potato alkaloids alpha-chaconine, alpha-solanine and solanidine in the Ames Salmonella and adult and foetal erythrocyte micronucleus assays. Food and Chemical Toxicology, 30, 689-694.

Friedman M and Levin CE, 1992. Reversed-phase high-performance liquid chromatographic separation of potato glycoalkaloids and hydrolysis products on acidic columns. Journal of Agricultural and Food Chemistry, 40, 2157-2163.

Friedman M and Levin CE, 1995. Tomatine Content in Tomato and Tomato Products Determined by HPLC with Pulsed Amperometric Detection. Journal of Agricultural and Food Chemistry, 1995, 43, 1507-1511.

4494 Friedman M and Levin CE, 1998. Dehydrotomatine content in tomatoes. Journal of Agricultual and Food Chemistry,
4495 46, 4571-4576.
4496 Friedman M and McDonald GM, 1995. Acid-catalyzed partial hydrolysis of carbohydrate groups of the potato
4497 glycoalkaloid a-chaconine in alcoholic solutions. Journal of Agricultural and Food Chemistry, 43, 1501-1506.
4498 Friedman M and McDonald GM, 1997. Potato glycoalkaloids: chemistry, analysis, safety, and plant physiology.
4499 Critical Reviews in Plant Sciences, 16, 55-132.
4500 Friedman M, Levin CE and McDonald GM, 1994. a-Tomatine determination in tomatoes by HPLC using pulsed
4501 amperometric detection. Journal of Agricultural and Food Chemistry, 42, 1959-1964.
4502 Friedman M, Henika PR and Mackey BE, 1996. Feeding of potato, tomato and eggplant alkaloids affects food
4503 consumption and body and liver weights in mice. Journal of Nutrition, 126, 989-999.
4504 Friedman M, McDonald GM and Filadelfi-Kesyi M, 1997. Potato glycoalkaloids: chemistry, analysis, safety, and plant
4505 physiology. Critical Reviews in Plant Sciences, 16, 55-132.
4506 Friedman M, Bautista FF, Stanker LH and Larkin KA, 1998. Analysis of potato glycoalkaloids by a new ELISA Kit.
4507 Journal of Agricultural and Food Chemistry, 46, 5097-5102.
4508 Friedman M, Fitch TE and Yokoyama WE, 2000a. Lowering of plasma LDL cholesterol in hamsters by the tomato
4509 glycoalkaloid tomatine. Food and Chemical Toxicology, 38, 549-553.
4510 Friedman M, Fitch TE, Levin CE and Yokoyama WH, 2000b. Feeding Tomatoes to HamstersReduces their Plasma
4511 Low-density Lipoprotein Cholesterol and Triglycerides. Journal of Food Science, 65, 897-900.
4512 Friedman M, Henika PR and Mackey BE, 2003a. Effect of feeding solanidine, solasodine and tomatidine to non-
4513 pregnant and pregnant mice. Food and Chemical Toxicology, 41, 61-71.
4514 Friedman M, Roitman JN and Kozukue N, 2003b. Glycoalkaloid and Calystegine Contents of Eight Potato Cultivars.
4515 Journal of Agricultural and Food Chemistry, 51, 2964–2973.
4516 Friedman M, Kozukue N, Kim H-J, Choi S-H and Mizuno M, 2017. Glycoalkaloid, phenolic, and flavonoid content and
4517 antioxidative activities of conventional nonorganic and organic potato peel powders from commercial gold,
4518 red, and Russet potatoes. Journal of Food Composition and Analysis, 62, 69-75.
4519 Fujiwara Y, Takaki A, Uehara Y, Ikeda T, Okawa M, Yamauchi K, Ono M, Yoshimitsu H, Nohara T, 2004. Tomato
4520 steroidal alkaloid glycosides, esculeosides A and B, from ripe fruits. Tetrahedron, 60, 4915–4920.
4521 Furui H, Inakuma T, Ishiguro Y and Kiso M, 1997. Absorptiometric measurement of tomatine in tomatoes. Nippon
4522 Nogei Kagaku Kaishi, 71, 777-782.
4523 Gaffield W and Keeler RF, 1993. Implication of C-5, C-6 unsaturation as a key structural factor in steroidal alkaloid-
4524 induced mammalian teratogenesis. Experientia, 49, 922-924.
4525 Gaffield W and Keeler RF, 1996a. Induction of terata in hamsters by solanidane alkaloids derived from *Solanum*
4526 *tuberosum*. Chemical Research in Toxicology, 9, 426-433.
4527 Gaffield W and Keeler RF, 1996b. Craniofacial malformations induced in hamsters by steroidal alkaloids. Journal of
4528 Natural Toxins, 5, 25-38.
4529 Gattermann R, Fritzsche P, Weinandy R and Neumann K, 2002. Comparative studies of body mass, body
4530 measurements and organ weights of wildderived and laboratory golden hamsters (*Mesocricetus auratus*).
4531 Laboratory Animals, 36, 445–454.
4532 Gee JM, Wortley GM, Johnson IT, Price KR, Rutten AA, Houben GF and Penninks AH, 1996. Effects of saponins and
4533 glycoalkaloids on the permeability and viability of mammalian intestinal cells and on the integrity of tissue
4534 preparations in vitro. Toxicology In Vitro, 10, 117-128.
4535 Ginzberg I, Tokuhisa JG and Veilleux RE, 2009. Potato Steroidal Glycoalkaloids: Biosynthesis and Genetic
4536 Manipulation. Potato Research, 52, 1-15.
4537 Glynn A, Igra AM, Sand S, Ilbäck NG, Hellenäs KE, Rósen J and Aspenström-Fagerlund B, 2017. Are additive effects
4538 of dietary surfactants on intestinal tight junction integrity an overlooked human health risk? - A mixture study
4539 on Caco-2 monolayers. Food and Chemical Toxicology, 106, 314-323.
4540 Gonmori K, Meguro H, Lu YQi, Yoshioka N, Hori K and Kikkawa S, 1993. The risk of solanine poisoning in a folk
4541 remedy and solanine production in potato. Research and Practice in Forensic Medicine, 36, 5. Only abstract
4542 in English.
4543 Grandison L, Gelato M and Meites J, 1974. Inhibition of prolactin secretion by cholinergic drugs. Proceedings of the
4544 Society for Experimental Biology and Medicine, 145, 1236-1239.
4545 Griebel C, 1923. Solaninreiche gesundheitsschädliche Kartoffeln. Zeitschrift fuer Untersuchung der Nahrungs- und
4546 Genussmittel sowie der Gebrauchsgegenstaende, 45, 175-183.
4547 Griffiths DW, Bain H and Dale FB, 1998. Effect of Storage Temperature on Potato (Solanum tuberosum L.) Tuber
4548 Glycoalkaloid Content and the Subsequent Accumulation of Glycoalkaloids and Chlorophyll in Response to
4549 Light Exposure. Journal of Agricultural adn Food chemistry, 46, 5262-5268.
4550 Gosselin B and Mondy NI, 1989. Effect of packaging materials on the chemical composition of potatoes. Journal of
4551 Food Science, 54, 629-631.
4552 Gosselin B, Mondy NI and Evans WD, 1988. Effect of method of irrigation on the total glycoalkaloid and nitrate-
4553 nitrogen content of Rosa potatoes. American Potato Journal, 65, 99-103.
4554 Groen K, Pereboom-de Fauw DP, Besamusca P, Beekhof PK, Speijers GJ and Derks HJ, 1993. Bioavailability and
4555 disposition of 3H-solanine in rat and hamster. Xenobiotica, 23, 995-1005.
4556 Gull DD, 1960. Chlorophyll and solanine changes in tubers of Solanum Tuberosum induced by fluorescent light, and
4557 a study of solanine toxicology by bioassay technique. PhD, Cornell University, Michigan, 149, 11-139.

4558 Gunning OV, 1950. Suspected potato poisoning in a mare. British Veterinary Journal, 106, 32-33.
4559 Gupta RS and Dixit VP, 2002. Effects of short term treatment of solasodine on cauda epididymis in dogs. Indian
4560 Journal of Experimental Biology, 40, 169-173.
4561 Hamouz K, Pazderu K, Lachman J, Orsak M, Pivec V, Hejtmankova K, Tomasek J and Cizek M, et al 2014. Effect of
4562 cultivar, flesh colour, location and year of cultivation on the glycoalkaloid content in potato tubers. Plant Soil
4563 and Environment, 60, 512-517.
4564 Hansen AA, 1925. Two fatal cases of potato poisoning. Science, 61, 340-341.
4565 Hajšlová J, Schulzová V, Slanina P, Janné K, Hellenäs KE and Andersson CE, 2005. Quality of organically and
4566 conventionally grown potatoes: Four-year study of micronutrients, metals, secondary metabolites, enzymic
4567 browning and organoleptic properties. Food Additives and Contaminants, 22, 514-534.
4568 Harris FW and Cockburn T, 1918. Alleged poisoning by potatoes. Analyst, 43, 133-137.
4569 Harvey MH, McMillan M, Morgan MR and Chan HW, 1985a. Solanidine is present in sera of healthy individuals in
4570 amounts dependent on their dietary potato consumption. Human Toxicology, 4, 187-194.
4571 Harvey MH, Morris BA, McMillan M and Marks V, 1985b. Measurement of potato steroidal alkaloids in human serum
4572 and saliva by radioimmunoassay. Human Toxicology, 4, 503-512.
4573 Harvey MH, Morris BA, McMillan M and Marks V, 1986. Potato steroidal alkaloids and neural tube defects: serum
4574 concentrations fail to demonstrate a causal relation. Human Toxicology, 5, 249-253.
4575 Hellenäs KE, 1986. A simplified procedure for quantification of potato glycoalkaloids in tuber extracts by HPLC;
4576 comparison with ELISA and a colorimetric method. Journal of the Science of Food and Agriculture, 37, 776-
4577 782.
4578 Hellenäs KE, Branzell C, Johnsson H and Slanina P, 1995a. Glycoalkaloid content of early potato varieties. Journal
4579 of the Science of Food and Agriculture, 67, 125-128.
4580 Hellenäs K-E, Branzell C, Johnsson H and Slanina P, 1995b. High levels of glycoalkaloids in the established Swedish
4581 potato variety Magnum Bonum. Journal of the Science of Food and Agriculture, 68, 249-255.
4582 Hellenäs KE, Nyman A, Slanina P, Lööf L and Gabrielsson J, 1992. Determination of potato glycoalkaloids and their
4583 aglycone in blood serum by high-performance liquid chromatography. Application to pharmacokinetic studies
4584 in humans. Journal of Chromatography, 573, 69-78.
4585 Harris H and Whittaker M, 1959. Differential response of human serum cholinesterase types to an inhibitor in potato.
4586 Nature, 183, 1808-1809.
4587 Harris H and Whittaker M, 1962. Differential inhibition of the serum cholinesterase phenotypes by solanine and
4588 solanidine. Annals of Human Genetics, 26, 73-76.
4589 Heppner KM, Tong J, Kirchner H, Nass R and Tschöp MH, 2011. The ghrelin O-acyltransferase-ghrelin system: a
4590 novel regulator of glucose metabolism. Current Opinion in Endocrinology, Diabetes and Obesity, 18, 50-55.
4591 Herb SF, Fitzpatrick TJ and Osman SF, 1975. Separation of potato glycoalkaloids by gas chromatography. Journal
4592 of Agricultural and Food Chemistry, 23, 520-523.
4593 Hlywka JJ, Stephenson GR, Sears MK and Yada RY, 1994. Effects of insect damage on glycoalkaloid content in
4594 potatoes (Solanum tuberosum). Journal of Agricultural and Food Chemistry, 42, 2545-2550.
4595 Hopkins J, 1995. The glycoalkaloids: naturally of interest (but a hot potato?). Food and Chemical Toxicology, 33,
4596 323-328.
4597 Hossain MB, Rai DK and Brunton NP, 2015. Optimisation and validation of ultra-high performance liquid
4598 chromatographic-tandem mass spectrometry method for qualitative and quantitative analysis of potato
4599 steroidal alkaloids. Journal of Chromatography B, 997, 110–115.
4600 Hossain MB, Brunton NP and Rai DK, 2016. Effect of drying methods on the steroidal alkaloid content of potato
4601 peels, shoots and berries. Molecules, 21, 403.
4602 Houben RJ and Brunt K, 1994. Determination of glycoalkaloids in potato tubers by reversed-phase high-performance
4603 liquid chromatography. Journal of Chromatography A, 661, 169-174.
4604 Hugenholtz F and de Vos WM, 2018. Mouse models for human intestinal microbiota research: a critical evaluation.
4605 Cellular and Molecular Life Sciences, 75, 149-160.
4606 Iablokov V, Sydora BC, Foshaug R, Meddings J, Driedger D, Churchill T and Fedorak RN, 2010. Naturally occurring
4607 glycoalkaloids in potatoes aggravate intestinal inflammation in two mouse models of inflammatory bowel
4608 disease. Digestive Diseases and Sciences, 55, 3078-3085.
4609 Ieri F, Innocenti M, Andrenelli L, Vecchio V and Mulinacci N, 2011. Rapid HPLC/DAD/MS method to determine
4610 phenolic acids, glycoalkaloids and anthocyanins in pigmented potatoes (*Solanum tuberosum* L.) and
4611 correlations with variety and geographical origin. Food Chemistry, 125, 750–759.
4612 Iijima Y, Fujiwara Y, Tokita T, Ikeda T, Nohara T, Aoki K, Shibata D, 2009. Involvement of ethylene in the
4613 accumulation of esculeoside a during fruit ripening of tomato (*Solanum lycopersicum*). Journal of Agricultural
4614 and Food Chemistry, 57, 3247–3252.
4615 Iijima Y, Watanabe B, Sasaki R, Takenaka M, Ono H, Sakurai N, Umemoto N, Suzuki H, Shibata D and Aoki K, 2013.
4616 Steroidal glycoalkaloid profiling and structures of glycoalkaloids in wild tomato fruit. Phytochemistry, 95, 145-
4617 157.
4618 Jandrić Z, Rathor MN, Švarc-Gajić J, Maestroni BM, Sasanya JJ, Djurica R and Cannavan A, 2011. Development of
4619 a liquid chromatography-tandem mass spectrometric method for the simultaneous determination of tropane
4620 alkaloids and glycoalkaloids in crops. Food Additives and Contaminants, 28, 1205–1219

4621 JECFA (Joint FAO/WHO Expert Committee on Food Additives), 1992. Evaluation of certain food additives and
4622 naturally occurring toxicants. Thirty-ninth report of the Joint FAO/WHO Expert Committee on Food Additives.
4623 World Health Organisation Technical Report Series, 828, 1-49.
4624 JECFA (Joint FAO/WHO Expert Committee on Food Additives), 1993. Toxicological evaluation of certain food
4625 additives and naturally occurring toxicants. WHO Food Additives Series, Vol. 30. World Health Organization,
4626 Geneva.
4627 Jin CY, Liu H, Xu D, Zeng FK, Zhao YC, Zhang H and Liu G, 2018. Glycoalkaloids and phenolic compounds in three
4628 commercial potato cultivars grown in Hebei, China. Food Science and Human Wellness, 7, 156-162.
4629 Jones PG and Fenwick GR, 1981. The glycoalkaloid content of some edible solanaceous fruits and potato products.
4630 Journal of the Science of Food and Agriculture, 32, 419-421.
4631 Karaway MA, 2014. Removal of pharmacological undesirable compounds from potato tuber. Research and Reviews
4632 in BioSciences, 8, 73-77.
4633 Karmakar K, Islam A, Chhanda SA, Tuhin TI, Muslim T and Rahman A, 2015. Secondary metabolites from the fruits
4634 of *Solanum torvum* SW. Journal of Pharmacognosy and Phytochemistry, 4, 160-163.
4635 Keeler RF, Young S and Brown D, 1976. Spina bifida, exencephaly, and cranial bleb produced in hamsters by the
4636 solanum alkaloid solasodine. Research Comunications in Chemical Pathology and Pharmacology, 13, 723-730.
4637 Keeler RF, Baker DC and Gaffield W, 1990. Spirosolane-containing Solanum species and induction of congenital
4638 craniofacial malformations Toxicon, 28, 873-884.
4639 Kees M, Beckel N and Sharp C, 2015. Successful treatment of Solanum dulcamara intoxication in a Labrador retriever
4640 puppy. Canadian Veterinary Journal, 56, 1283–1286.
4641 Kerr CA, Goodband RD, Smith II JW, Musser RE, Bergstro¨m JR, Nessmith WG Jr, Tokach MD and Nelssen JL, 1998.
4642 Evaluation of Potato Proteins on the Growth Performance of Early-Weaned Pigs. Journal of Animal Science,
4643 76, 3024–3033.
4644 Keukens EAJ, Vrije Td, Fabrie CHJP, Demel t' RA, Jongen WMF and Kruijff Bd, 1992. Dual specificity of sterol-
4645 mediated glycoalkaloid induced membrane disruption Biochunica et Biophysica Acta, 1110, 127-136.
4646 Keukens EAJ, Hop MECM and Jongen WMF, 1994. Rapid High-Performance Liquid Chromatographic Method for the
4647 Quantification of a-Tomatine in Tomato. Journal of Agricultural and Food Chemistry, 42, 2475-2477.
4648 Keukens EAJ, deVrije T, vandenBoom C, deWaard P, Plasman HH, Thiel F, Chupin V, Jongen WMF, deKruijff B,
4649 1995. Molecular basis of glycoalkaloid induced membrane disruption. Biochimica et Biophysica Acta, 1240,
4650 216–228.
4651 Keukens EAJ, deVrije T, Jansen LAM, deBoer H, Janssen M, deKroon AIPM, Jongen WMF and deKruijff B, 1996.
4652 Glycoalkaloids selectively permeabilize cholesterol containing biomembranes. Biochimica et Biophysica Acta,
4653 1279, 243–250.
4654 Khaserao S and Somani R, 2017. Evaluation of anti-obesity activity of solasodine in high fat diet-induced obesity in
4655 rat. International Journal of Pharmacy and Pharmaceutical Sciences, 9, 23-29.
4656 Khodayari S, Alizadeh AM, Kouhpayeh SA, Mohsenikia M, Karimi AA, Khodayari H, Abdolahi AR, Sarbiyaii A, Erfani
4657 S, Kazemi S and Zakeri H, 2013. The acute and chronic toxicity effects of alpha-solanine in mice. Journal of
4658 Babol University of Medical Sciences, 15, 24-31.
4659 Kibler R, Lang H and Ziegler W, 1985. Einfluß küchentechnischer Maßnahmen auf den Solaningehalt grüner
4660 Tomatenfrüchte. Deutsche Lebensmittel-Rundschau, 81, 111-113.
4661 King RR and McQueen RE, 1981. Transformations of potato glycoalkaloids by rumen microorganisms. Journal of
4662 Agricultural and Food Chemistry, 29, 1101-1103.
4663 Kline BE, von Elbe H, Dahle NA and Morris Kupchan S, 1961. Toxic effects of potao sprouts and of solanine fed to
4664 pregnant rats. Proceedings of the Society for Experimental Biology and Medicine, 107.
4665 Knapp S, 2013. Typification of *Solanum* species (Solanaceae) described by Casamiro Gómez Ortega. Anales del
4666 Jardín Botánico de Madrid, 70(1), 56-61.
4667 Knuthsen P, Jensen U, Schmidt B and Larsen IK, 2009. Glycoalkaloids in potatoes: Content of glycoalkaloids in
4668 potatoes for consumption. Journal of Food Composition and Analysis, 22, 577-581.
4669 Koh E, Kaffka S and Mitchell AE, 2013. A long-term comparison of the influence of organic and conventional crop
4670 management practices on the content of the glycoalkaloid alpha-tomatine in tomatoes. Journal of the Science
4671 of Food Agriculture, 93, 1537–1542.
4672 Koleva II, van Beek TA, Soffers AEMF, Dusemund B and Rietjens IMCM, 2012. Alkaloids in the human food chain -
4673 Natural occurrence and possible adverse effects. Molecular Nutrition and Food Research, 56, 30-52.
4674 König H and Staffe A, 1953. Beitrag zur Wirkung des Solanins auf Blutbild und -Katalase beim Schaf (Action of
4675 solanine upon blood picture and blood catalase of sheep). Deutsche Tierärztliche Wochenschrift, 60, 150-153.
4676 Kozukue N, Kozukue E and Mizuno S, 1987. Glycoalkaloids in potato plants and tubers. HortScience, 22, 294–296.
4677 Kozukue N and Mizuno S, 1990. Effects of Light Exposure and Storage Temperature on Greening and Glycoalkaloid
4678 Content in Potato Tubers. Journal of the Japanese Society for Horticultural Science, 59, 673-677.
4679 Kozukue N and Friedman M, 2003. Tomatine, chlorophyll, β-carotene and lycopene content in tomatoes during
4680 growth and maturation. Journal of the Science of Food and Agriculture, 83, 195-200.
4681 Kozukue N, Han JS, Lee KR and Friedman M, 2004. Dehydrotomatine and alpha-tomatine content in tomato fruits
4682 and vegetative plant tissues. Journal of Agricultural and Food Chemistry, 52, 2079-2083.
4683 Kuhn R and Löw I, 1954. Die Konstitution des Solanins. Angewandte Chemie, 66, 639-640.

4684 Kyzlink V, Míková K and Jelínek R, 1981. Tomatine, solanine and embryotoxicity of unripe tomatoes. Sbornik Vysoké
4685 Školy Chemicko-Technologické v Praze, Potraviny E51, 69-83.
4686 Lachman J, Hamouz K, Musilová J, Hejtmánková K, Kotíková Z, Pazderů K, Domkářová J, Pivec V and Cimr J, 2013.
4687 Effect of peeling and three cooking methods on the content of selected phytochemicals in potato tubers with
4688 various colour of flesh. Food Chemistry, 138, 1189-1197.
4689 Lampitt LH, Bushill JH, Rooke HS and Jackson EM, 1943. Solanine, glycoside of the potato. II. Distribution in the
4690 potato plant. Journal of the Society of Chemical Industry, London, 62, 48-51.
4691 Langkilde S, Schroder M, Stewart D, Meyer O, Conner S, Davies H and Poulsen M, 2008. Acute toxicity of high
4692 doses of the glycoalkaloids, alpha-solanine and alpha-chaconine, in the Syrian Golden hamster. Journal of
4693 Agricultural and Food Chemistry, 56, 8753-8760.
4694 Langkilde S, Mandimika T, Schroder M, Meyer O, Slob W, Peijnenburg A and Poulsen M, 2009. A 28-day repeat
4695 dose toxicity study of steroidal glycoalkaloids, alpha-solanine and alpha-chaconine in the Syrian Golden
4696 hamster. Food and Chemical Toxicology, 47, 1099-1108.
4697 Langkilde S, Schroder M, Frank T, Shepherd LVT, Conner S, Davies HV, Meyer O, Danier J, Rychlik M, Belknap WR,
4698 McCue KF, Engel K-H, Stewart D, Knudsen I and Poulsen M, 2012. Compositional and toxicological analysis of
4699 a GM potato line with reduced alpha-solanine content - a 90-day feeding study in the Syrian Golden hamster.
4700 Regulatory Toxicology and Pharmacology, 64, 177-185.
4701 Layer PG, Klaczinski J, Salfelder A, Sperling LE, Thangaraj G, Tuschl C and Vogel-Höpker A, 2013. Cholinesterases
4702 in development: AChE as a firewall to inhibit cell proliferation and support differentiation. Chemico-Biological
4703 Interactions, 203, 269-276.
4704 Lee MR, 2006. The Solanaceae: foods and poisons. The journal of the Royal College of Physicians of Edinburgh,
4705 36, 162-169.
4706 Lee C and De Luca C, 2019. Potato propagation North Dacota State University. Plant Propagation PLSC 368.
4707 Available at: (Accessed January 2019)
4708 https://www.ndsu.edu/pubweb/chiwonlee/plsc368/student/papers06/carina%20de%20luca/carinadeluca.ht
4709 m
4710 Lee KG, Lee SG, Lee HH, Lee HJ, Shin JS, Kim NJ, An HJ, Nam JH, Jang DS and Lee KT, 2015. α-Chaconine isolated
4711 from a Solanum tuberosum L. cv Jayoung suppresses lipopolysaccharide-induced pro-inflammatory mediators
4712 via AP-1 inactivation in RAW 264.7 macrophages and protects mice from endotoxin shock. Chem Biol Interact,
4713 235, 85-94.
4714 Lelario F, De Maria S, Rivelli AR, Russo D, Milella L, Bufo SA and Scrano L, 2019. A Complete Survey of Glycoalkaloids
4715 Using LC-FTICR-MS and IRMPD in a Commercial Variety and a Local Landrace of Eggplant (*Solanum*
4716 *melongena L.*) and their Anticholinesterase and Antioxidant Activities. Toxins, 11, 230.
4717 Leonardi C, Ambrosino P, Esposito F and Fogliano V, 2000. Antioxidative activity and carotenoid and tomatine
4718 contents in different [typologies of fresh consumption tomatoes.]. Journal of Agricultural and Food Chemistry,
4719 48, 4723-4727.
4720 Lepper W, 1949. Solanine content of 58 varieties of potatoes. Zeitschrift für Lebensmittel-Untersuchung und -
4721 Forschung, 89, 264-273.
4722 Li H-J, Jiang Y and Li P, 2006. Chemistry, bioactivity and geographical diversity of steroidal alkaloids from the
4723 Liliaceae family. Natural Products Reports, 23, 735-752.
4724 Liebenow H, 1970. Solanum nigrum as a nitrate-containing plant and the determination of its alkaloid content.
4725 Wissenschaftliche Zeitschrift der Humboldt-Universität zu Berlin, Mathematisch-Naturwissenschaftliche Reihe,
4726 19, 59-71.
4727 Liljemark A and Widoff E, 1960. Greening and solanine development of white potato in fluorescent light. American
4728 Potato Journal, 37, 379-389.
4729 Lionetto MG, Caricato R, Calisi A, Giordano ME and Schettino T, 2013. Acetylcholinesterase as a Biomarker in
4730 Environmental and Occupational Medicine: New Insights and Future Perspectives. Biomedical Research
4731 International, 321213.
4732 Liu W, Zhang N, Li B, Fan S, Zhao R, Li Li-P, Wu G-H and Zhao Y, 2014. Determination of α-chaconine and α-
4733 solanine in commercial potato crisps by QuEChERS extraction and UPLC-MS/MS. Chemical Papers, 68, 1498-
4734 1504.
4735 Lo Scalzo R, Fibinani M, Francese G, D´Alessandro A, Rotino GL, Conte P and Mennella G, 2016. Cooking influence
4736 on physicochemical fruit characteristics of eggplant (*Solanum melongena* L.). Food Chemistry, 194, 835-42.
4737 Loewenstein-Lichtenstein Y, Glick D, Gluzman N, Sternfeld M, Zakut H and Soreq H, 1996. Overlapping drug
4738 interaction sites of human butyrylcholinesterase dissected by site-directed mutagenesis. Molecular
4739 Pharmacology, 50, 1423-1431.
4740 Love SL, Herrman TJ, Thompsonjohns A and Baker TP, 1994. Effect and interaction of crop management factors
4741 on the glycoalkaloid concentration of potato tubers. Potato Research, 37, 77-85.
4742 Machado RMD, Toledo MCF and García LC, 2006. Effect of light and temperature on the formation of glycoalkaloids
4743 in potato tubers. Food Control, 18, 503-508.
4744 Machida-Hirano R, 2015. Diversity of potato genetic resources. Breeding Science, 65, 26-40.
4745 Mäder J, Rawel H and Kroh LW, 2009. Composition of phenolic compounds and glycoalkaloids α- solanine and α-
4746 chaconine during commercial potato processing. Journal of Agricultural and Food Chemistry, 57, 6292-6297.

4747 Majamaa H, Seppala U, Palosuo T, Turjanmaa K, Kalkkinen N and Reunala T, 2001. Positive skin and oral challenge
4748 responses to potato and occurrence of immunoglobulin E antibodies to patatin (Sol t 1) in infants with atopic
4749 dermatitis. Pediatric Allergy and Immunology, 12, 283-288.
4750 Manrique-Moreno M, Londoño-Londoño J, Jemioła-Rzemińska M, Strzałka K, Villena F, Avello M and Suwalsky M,
4751 2014. Structural effects of the Solanum steroids solasodine, diosgenin and solanine on human erythrocytes
4752 and molecular models of eukaryotic membranes. Biochimica et Biophysica Acta, 1838, 266-277.
4753 Martínez-Valverde I, Periago MJ, Provan G and Chesson A, 2002. Phenolic compounds, lycopene and antioxidant
4754 activity in commercial varieties of tomato (*Lycopersicum esculentum*). Journal of the Science of Food and
4755 Agriculture, 82, 323-330.
4756 Matthew JA, Morgan MRA, McNerney R, Chan HW-S and Coxon DT, 1983. Determination of solanidine in human
4757 plasma by radioimmunoassay. Food and Chemical Toxicology, 21, 637-640.
4758 Maurya A, Manika N, Verma RK, Singh SC and Srivastava SK, 2013. Simple and Reliable Methods for the
4759 Determination of Three Steroidal Glycosides in the Eight Species of Solanum by Reversed-phase HPLC Coupled
4760 with Diode Array Detection Phytochemical Analysis, 24, 87–92.
4761 McGehee DS, Krasowski MD, Fung DL, Wilson B, Gronert GA and Moss J, 2000. Cholinesterase inhibition by potato
4762 glycoalkaloids slows mivacurium metabolism. Anesthesiology, 93, 510-9.
4763 McMillan M and Thompson JC, 1979. An outbreak of suspected solanine poisoning in schoolboys: Examinations of
4764 criteria of solanine poisoning. The Quarterly Journal of Medicine, 48, 227-243.
4765 Mennella G, Lo Scalzo R, Fibiani M, D'Alessandro A, Francese G, Toppino L, Acciarri N, de Almeida AE and Rotino
4766 GL, 2012. Chemical and bioactive quality traits during fruit ripening in eggplant (*S. melongena* L.) and allied
4767 species. Journal of Agricultural and Food Chemistry, 60, 11821-11831.
4768 Mensinga TT, Sips AJAM, Rompelberg CJM, van Twillert K, Meulenbelt J, van den Top HJ and van Egmond HP,
4769 2005. Potato glycoalkaloids and adverse effects in humans: an ascending dose study. Regulatory Toxicology
4770 and Pharmacology, 41, 66-72.
4771 Mesulam MM, Guillozet A, Shaw P, Levey A, Duysen EG and Lockridge O, 2002. Acetylcholinesterase knockouts
4772 establish central cholinergic pathways and can use butyrylcholinesterase to hydrolyze acetylcholine.
4773 Neuroscience, 110, 627-639.
4774 Miceli A, Sabatino L, Moncada A, Vetrano F and D'Anna F, 2014. Nursery and field evaluation of eggplant grafted
4775 onto unrooted cuttings of Solanum torvum Sw. Scientia Horticulturae, 178, 203-210.
4776 Michalska L, Nagel G, Swiniarski E and Zydowo MM, 1985. The effect of alpha-solanine on the active calcium
4777 transport in rat intestine. Gen Pharmacol, 16, 69-70.
4778 Milner SE, Brunton NP, Jones PW, O'Brien NM, Collins SG and Maguire AR, 2011. Bioactivities of glycoalkaloids and
4779 their aglycones from Solanum species. Journal of Agricultural and Food Chemistry, 59, 3454-3484.
4780 Milligan JB, 1941. Suspected potato poisoning in cattle. Veterinary Record, 53, 512
4781 Moco S, Capanoglu E, Tikunov Y, Bino RJ, Boyacioglu D, Hall RD, Vervoort J and de Vos RCH, 2007. Tissue
4782 specialization at the metabolite level is perceived during the development of tomato fruit. Journal of
4783 Experimental Botany, 58, 4131-46.
4784 Mohsenikia M, Alizadeh AM, Khodayari S, Khodayari H, kouhpayeh SA, Karimi A, Zamani M, Azizian S and
4785 Mohagheghi MA, 2013. The protective and therapeutic effects of alpha-solanine on mice breast cancer.
4786 European Journal of Pharmacology, 718, 1-9.
4787 Mondy NI and Gosselin B, 1988. Effect of peeling on total phenols, total glycoalkaloids, discoloration and flavor of
4788 cooked potatoes. Journal of Food Science, 53, 756-759.
4789 Mondy NI, Leja M and Gosselin B, 1987. Changes in total phenolic, total glycoalkaloid, and ascorbic acid content of
4790 potatoes as a result of bruising. Journal of Food Science, 52, 631-633.
4791 Mondy NI and Munshi CB, 1990. Effect of nitrogen fertilization on glycoalkaloid and nitrate content of potatoes.
4792 Journal of Agricultural and Food Chemistry, 38, 565-567.
4793 Morgan MRA, Coxon DT, Bramham S, Chan HWS, Van Gelder WMJ and Allison MJ, 1985. Determination of the
4794 glycoalkaloid content of potato tubers by three methods including enzyme-linked immunosorbent assay.
4795 Journal of the Science of Food and Agriculture, 36, 282-288.
4796 Mori K, Asano K, Tamiya S, Nakao T and Mori M, 2015. Challenges of breeding potato cultivars to grow in various
4797 environments and to meet different demands. Breed Science, 65, 3-16.
4798 Morris T, 1859. Case of poisoning by potatoes-berries. British Medical Journal, 719-720. As cited by Van Gelder,
4799 1989.
4800 Morris SC and Lee TH, 1984. The toxicity and teratogenicity of Solanaceae glycoalkaloids, particularly those of the
4801 potato (Solanum tuberosum): a review. Food Technology in Australia, 36, 118-124.
4802 Morris SC and Petterman JB, 1985. Genetic and environmental effects on levels of glycoalkaloids in cultivars of
4803 potato (Solanum tuberosum L.). Food Chemistry, 18, 271-282.
4804 Muncke, 1845. Med. Ann. Heidelberg XI:298. As cited by Bömer and Matthis, 1924.
4805 Muneta P, Shen W-W, Jennings S, Everson D and Edwarsa L, 1977. Potassium hydroxide for peeling potatoes.
4806 American Potato Journal, 54, 83-90.
4807 Mulinacci N, Ieri F, Giaccherini C, Innocenti M, Andrenelli L, Canova G, Saracchi M and Casiraghi MC, 2008. Effect
4808 of cooking on the anthocyanins, phenolic acids, glycoalkaloids, and resistant starch content in two pigmented
4809 cultivars of *Solanum tuberosum* L. Journal of Agricultural Food Chemistry, 56, 11830-11837.
4810 Mulder P, 2019. Personal communication provided via email.

4811 Mulatu B, Applebaum SW, Zerem Z and Coll M, 2006. Tomato fruit size, maturity and a-tomatine content influence
4812 the performance of larvae of potato tuber moth Phthorimaea operculella (Lepidoptera: Gelechiidae). Bulletin
4813 of Entomological research, 96, 173-178.
4814 Munari CC, de Oliveira PF, de Souza Lima IM, de Paula Lima Martins S, de Carvalho da Costa J, Bastos JK and
4815 Tavares DC, 2012. Evaluation of cytotoxic, genotoxic and antigenotoxic potential of Solanum lycocarpum fruits
4816 glicoalkaloid extract in V79 cells. Food Chemistry and Toxicology, 50, 3696-701.
4817 Munari CC, de Oliveira PF, Leandro LF, Pimenta LM, Ferreira NH, da Costa J de C, Bastos JK, Tavares DC, 2014. In
4818 vivo assessment of genotoxic, antigenotoxic and anticarcinogenic activities of *Solanum lycocarpum* fruits
4819 glycoalkaloidic extract. PLoS One, 9, 1-8.
4820 Najm AA, Haidi MRHS, Fazeli F, Darzi MT and Rahi A, 2012. Effect of Integrated Management of Nitrogen Fertilizer
4821 and Cattle Manure on the Leaf Chlorophyll, Yield, and Tuber Glycoalkaloids of Agria Potato. Communications
4822 in Soil Science and Plant Analysis, 43, 912-923.
4823 Nara A, Saka K, Yamada C, Kodama T and Takagi T, 2019. Forensic analysis using ultra-high-performance liquid
4824 chromatography–tandem mass spectrometry with solid-phase extraction of α-solanine and α-chaconine in
4825 whole blood. Forensic Toxicology, 37, 197-206.
4826 Nema PK, Ramayya N, Duncan E and Niranjan K, 2008. Potato glycoalkaloids: formation and strategies for
4827 mitigation. Journal of the Science of Food and Agriculture, 88, 1869-1881.
4828 Ness E, Joner PE and Dahle HK, 1984. Alpha-solanine tested for mutagenicity with Ames test. Acta veterinaria
4829 Scandinavica, 25, 145-147.
4830 Nevin NC and Merrett JD, 1975. Potato avoidance during pregnancy in women with a previous infant with either
4831 anencephaly and/or spina bifida. British Journal of Preventive and Social Medicine, 29, 111-115.
4832 Ni W, Tian T, Zhang L, Wang L and Ren A, 2018. Maternal periconceptional consumption of sprouted potato and
4833 risks of neural tube defects and orofacial clefts. Nutrition Journal, 28,17-112
4834 Nie X, Zhang G, Lv S and Guo H, 2018. Steroidal glycoalkaloids in potato foods as affected by cooking methods.
4835 International Journal of Food Properties, 21, 1.
4836 Nie X, Li C, Zhang G, Shao Z, Wang X, Shi H and Guo H, 2019. Light exposure and wounding: synergistic effects
4837 on steroidal glycoalkaloid accumulation in potato tubers during storage. International Journal of Food Science
4838 and Technology, 54, 2939-2948.
4839 Nigg HN, Ramos LE, Graham EM, Sterling J, Brown S and Cornell JA, 1996. Inhibition of human plasma and serum
4840 butyrylcholinesterase (EC 3.1.1.8) by alpha-chaconine and alpha-solanine. Fundamental and Applied
4841 Toxicology, 33, 272-281.
4842 Nishie K, Gumbmann MR and Keyl AC, 1971. Pharmacology of solanine. Toxicology and Applied Pharmacology, 19,
4843 81-92.
4844 Nishie K, Norred WP and Swain AP, 1975. Pharmacology and toxicology of chaconine and tomatine. Research
4845 Communications in Chemical Pathology and Pharmacology, 12, 657-668.
4846 Nohara T, Ono M, Ikeda T, Fujiwara Y and El-Aasr, 2010. The tomato saponin, Esculeoside A. Journal of Natural
4847 Products, 73, 1734-1741.
4848 Nordic Working Group on Food Toxicology and Risk Assessment, 1990. Assessment of health-risks related to
4849 glycoalkaloids ('Solanine') in potatoes: a nordic view. Compiled by Slanina P. Vår Föda, 43, Supplement 1, 14
4850 pp.
4851 Norman TE, Chaffin MK, Norton PL, Coleman MC, Stoughton WB and Mays T, 2012. Concurrent ivermectin and
4852 Solanum spp. toxicosis in a herd of horses. Journal of Veterinary Internal Medicine, 26, 1439–1442.
4853 Norred WP, Nishie K and Osman SF, 1976. Excretion, distribution and metabolic fate of 3H-alpha-chaconine.
4854 Research Communications in Chemical Pathology and Pharmacology, 13, 161-171.
4855 Ogg AG Jr, Rogers BS and Schilling EE, 1981. Characterization of black nightshade (Solanum-nigrum) and related
4856 species in the United States. Weed Science, 29, 27-32.
4857 Ogg AG and Rogers BS, 1989. Taxonomy, distribution, biology, and control and black nightshade (Solanum nigrum)
4858 and related species in the United States and Canada. Reviews of Weed Science, 4, 25–58.
4859 Orgell WH, Vaidya KA and Dahm PA, 1958. Inhibition of human plasma cholinesterase in vitro by extracts of
4860 solanaceous plants. Science, 128, 1136-1137.
4861 Orgell WH, 1963. Inhibition of human plasma cholinesterase in vitro by alkaloids, glycosides, and other natural
4862 substances. Lloydia, 26, 36-43.
4863 Oshiro N, Kuniyoshi K, Nakamura A, Araki Y, Tamanaha K and Inafuku Y, 2008. A Case of Food Poisoning Due to
4864 Ingestion of Eggplant, *Solanum melongena*, Grafted on Devil's Trumpet, *Datura metel*. Shokuhin Eiseigaku
4865 Zasshi, 49, 376-379.
4866 Ostrý V, Ruprich J and Skarkova J, 2010. Glycoalkaloids in potato tubers: the effect of peeling and cooking in salted
4867 water. Acta Alimentaria, 39, 130-135.
4868 Panovská Z, Hajšlová J, Kosinková P and Čepl J, 1997. Glycoalkaloid content of potatoes sold in Czechia. Nahrung,
4869 41, 146- 149.
4870 Papathanasiou F, Mitchell SH and Harvey BMR, 1999. Variation in glycoalkaloid concentration of potato tubers
4871 harvested from mature plants. Journal of the Science of Food and Agriculture, 79, 32–36.
4872 Paradiso R, Arena C, Rouphael Y, d'Acquino L, Makris K, Vitaglione P and De Pascale S, 2019. Growth,
4873 photosynthetic activity and tuber quality of two potato cultivars in controlled environment as affected by light
4874 source. Plant Biosystems, 153, 725-735.

Pariera Dinkins CL and Peterson RKD, 2008. A human dietary risk assessment associated with glycoalkaloid responses of potato to Colorado potato beetle defoliation. Food and Chemical Toxicology, 46, 2837-2840.
Parnell A, Bhuva VS and Bintcliffe EJB, 1984. The glycoalkaloid content of potato varieties. Journal of the National Institute of Agricultural Botanic, 16, 535-541.
Park S, Park MY, Song G and Lim W, 2009. Alpha-solanine inhibits cell proliferation via mitochondrial dysfunction and inhibin synthesis in mouse testis In vitro and In vivo. Chemosphere, 235, 271-279
Patel B, Schutte R, Sporns P, Doyle J, Jewel L and Fedorak RN, 2002. Potato glycoalkaloids adversely affect intestinal permeability and aggravate inflammatory bowel disease. Inflammatory Bowel Diseases, 8, 340–346.
Patil BC, Sharma RP, Salunkhe DK and Salunkhe K, 1972. Evaluation of solanine toxicity. Food and Cosmetics Toxicology, 10, 395-398.
Pęksa A, Golubowska G, Aniolowski K, Lisinska G and Rytel, 2006. Changes of glycoalkaloids and nitrate content in potatoes during chip processing. Food Chemistry, 97, 151-156.
Percival G, 1999. Light-induced glycoalkaloid accumulation of potato tubers (Solanum tuberosum L). Journal of the Science of Food and Agriculture, 79, 1305-1310.
Pereira EFR, Aracava Y, DeTolla LJ Jr, Beecham EJ, Basinger GW Jr, Wakayama EJ and Albuquerque EX, 2014. Animal Models That Best Reproduce the Clinical Manifestations of Human Intoxication with Organophosphorus Compounds. Journal of Pharmacology and Experimental Therapy, 350, 313–321.
Petersson EV, Arif U, Schulzova V, Krtková V, Hajšlová J, Meijer J, Andersson HC, Jonsson L and Sitbon F, 2013. Glycoalkaloid and Calystegine Levels in Table Potato Cultivars Subjected to Wounding, Light, and Heat Treatments. Journal of Agricultural and Food Chemistry, 61, 5893−5902.
Pfuhl E, 1899. Regarding an outbreak of illness due to poisoning by solanine in potatoes. Deutsche Medizinische Wochenschrift, 25, 753-754.
Phillips BJ, Hughes JA, Phillips JC, Walters DG, Anderson D and Tahourdin CSM, 1996. A study of the toxic hazard that might be associated with the consumption of green potato tops. Food and Chemical Toxicology, 34, 439-448.
Pohanska M, 2014. Inhibitors of Acetylcholinesterase and Butyrylcholinesterase Meet Immunity. International Journal of Molecular Sciences, 15, 9809-9825.
Pokrovskii AA, 1956. Effect of alkaloids of growing potatoes on cholinesterase. Biokhimiia, 21, 683-688.
Pond WG and Maner JH, 1984. Swine production and nutrition. AVI Publishing Co., Westport, Connecticut: AVI Pub. Co, 1984. ISBN: 0870554506. Bookmark: https://trove.nla.gov.au/version/26200281
Poswillo DE, Sopher D and Mitchell S, 1972. Experimental induction of foetal malformation with "blighted" potato: A preliminary report. Nature, 239, 462-464.
Poswillo DE, Sopher D, Mitchell SJ, Coxon DT, Curtis RF and Price KR, 1973a. Investigations into the Teratogenic Potential of Imperfect Potatoes. Teratology, 8, 339-348.
Poswillo DE, Sopher D and Mitchell S, 1973b. Further investigations into the Teratogenic Potential of Imperfect potatoes. Nature, 224, 367-368.
Reale M, Costantini E, Di Nicola M, D'Angelo C, Franchi S, D'Aurora M, Di Bari M, Orlando V, Galizia S, Ruggieri S, Stuppia L, Gasperini C, Tata AM and Gatta V, 2018. Butyrylcholinesterase and Acetylcholinesterase polymorphisms in Multiple Sclerosis patients: implication in peripheral inflammation. Scientific Reports, 8, 1319.
Reelakh LA and Keem A, 1958. Sov. Med. 22, 129. As cited by McMillan and Thompson (1979).
Renwick JH, 1972. Hypothesis: anencephaly and spina bifida are usually preventable by avoidance of a specific but unidentified substance present in certain potato tubers. British Journal of Preventive and Social Medicine, 26, 67-88.
Renwick JH, Calringbold WDB, Earthy ME, Few JD and McLean ACS, 1984. Neural-Tube Defects Produced in Syrian Hamsters by Potato Glycoalkaloids. Terotology, 30, 371-381.
Rice SG, Nowak L, Duysen EG, Lockridge O, Lahiri DK and Reyes PF, 2007. Neuropathological and immunochemical studies of brain parenchyma in acetylcholinesterase knockout mice: Implications in Alzheimer's disease. Journal of Alzheimers Disease, 11, 481-489.
Roddick JG, 1996. Steroidal Glycoalkaloids: Nature and Consequences of Bioactivity. Advances in Experimental Medicine and Biology, 404, 277-295.
Roddick JG, 1989. The acetylcholinesterase-inhibitory activity of steroidal glycoalkaloids and their aglycones. Phytochemistry, 28, 2631-2634.
Romanucci V, Pisanti A, Di Fabio G, Davinelli S, Scapagnini G, Guaragna A and Zarrelli A, 2016. Toxin levels in different variety of potatoes: Alarming contents of α-chaconine. Phytochemistry Letters, 16, 103-107.
Romanucci V, Di Fabio G, Di Marino C, Davinelli S, Scapagnini G and Zarrelli A, 2018. Evaluation of new strategies to reduce the total content of α-solanine and α-chaconine in potatoes. Phytochemistry Letters, 23, 116-119.
Rosenfeld HJ, Sundell HA and Ringstad PLM, 1995. Influence of packaging materials and temperature on the glycoalkaloid content of potato tubers. Food Research International, 28, 481-484.
Ross H, Pasemann P and Nitzsche W, 1978. Der Glykoalkaloidgehalt von Kartoff elsorten in seiner Abhangigkeit von Anbauort und -jahr und seiner Beziehung zum Geschmack. Z. Pflanzenziichtg. 80, 64-79.
Rothe JC, 1919. Über Erkrankungen nach Genuss von solaninhaltigen Kartoffeln. Zeitsch Hyg, 88, 1-12.
Ruddick JA, Harwig J and Scott PM, 1974. Nonteratogenicity in rats of blighted potatoes and compounds contained in them. Teratology, 9, 165-168.

Ruprich J, Rehurkova I, Boon PE, Svensson K, Moussavian S, Van der Voet H, Bosgra S, Van Klaveren JD and Busk L, 2009. Probabilistic modelling of exposure doses and implications for health risk characterization: Glycoalkaloids from potatoes. Food and Chemical Toxicology, 47, 2899-2905.

Rytel E, Gołubowska G, Linińska G, Pęksa A and Aniołowski K, 2005. Changes in glycoalkaloid and nitrate contents in potatoes during French fries processing. Journal of the Science of Food and Agriculture, 85, 879-882.

Rytel E, 2012a. Changes in the levels of glycoalkaloids and nitrates after the dehydration of cooked potatoes. American Journal of Potato Research, 89, 501-507.

Rytel E, 2012b. The effect of industrial potato processing on the concentrations of glycoalkaloids and nitrates in potato granules. Food Control, 28, 380-384.

Rytel E, Tajner-Czopek A, Kita A, Kucharska AZ, Sokół-Łeztowska A and Hamouz K, 2018. Content of anthocyanins and glycoalkaloids in blue-fleshed potatoes and changes in the content of a-solanine and a-chaconine during manufacture of fried and dried products. International Journal of Food Science and Technology, 53, 719–727.

Sabatino L, Iapichino G, D'Anna F, Palazzolo E, Mennella G and Rotino GL, 2018. Hybrids and allied species as potential rootstocks for eggplant: Effect of grafting on vigour, yield and overall fruit quality traits. Scientia Horticulturae, 228, 81-90.

Sackmann W, Kern H and Wiesmann E, 1959. Untersuchungen über die biologischen Wirkungen von Tomatin and Solanin (Studies on the biological effects of tomatin and solanine). Schweizerische Zeitschrift fur Pathologie und Bakteriologie. Revue Suisse de Pathologie et de Bacteriologie, 22, 557-63.

Sánchez-Mata MC, Yokoyama WE, Hong Y-J and Prohens J, 2010. Alpha-solasonine and alpha-solamargine contents of gboma (Solanum macrocarpon L.) and scarlet (Solanum aethiopicum L.) eggplants. Journal of Agricultural and Food Chemistry, 58, 5502-5508.

Sander H, 1956. Studien über Bildung und Abbau von Tomatin in der Tomatenpflanze (The formation and degradation of tomatin in the tomato plant). Planta, 47, 374-400.

Sander H, 1958. Tomatin ein möglicher Ausgangsstoff für die Lycopinbildung. Naturwissenschaften, 45, 59-59.

Sarquis JI, Coria NA, Aguilar I and Rivera A, 2000. Glycoalkaloid content in Solanum species and hybrids from a breeding program for resistance to late blight (Phytophthora infestans). American Journal of Potato Research, 77, 295-302.

Satoh T, 1967. Glycemic effects of solanine in rats. Japanese Journal of Pharmacology, 17, 652-658.

Schmiedeberg O, 1892. Über die toxikologische Bedeutung des Solaningehaltes der Kartoffeln. Archiv für exp. Pathologie und Pharmakologie, Bd. XXXVI, S. 373. As cited by Bömer and Matthis (1924).

Schwarz A, Pinto E, Haraguchi M, de Oliveira CA, Bernardi MM and de Souza Spinosa H, 2007. Phytochemical study of *Solanum lycocarpum* (St. Hil) unripe fruit and its effect on rat gestation. Phytotherapy Research, 21, 1025-1028.

Sepelevs I, Nakurte I and Galoburda R, 2019. Biologically active compound stability in the industrial potato processing by-products. FOODBALT 2019, 131-136, ref. 40.

Sharma RP, Taylor MJ and Bourcier DR, 1983. Subcellular distribution of alpha-chaconine in mouse hepatocytes. Drug and Chemical Toxicology, 6, 219-234.

Sharma RP, Willhite CC, Shupe JL and Salunkhe DK, 1979. Acute toxicity and histopathological effects of certain glycoalkaloids and extracts of *Alternaria solani* or *Phytophthora infestans* in mice. Toxicology Letters, 3, 349-355.

Sharma RP, Willhite CC, WU MT and Salunkhe DK, 1978. Teratogenic potential of blighted potato concentrate in rabbits, hamster and miniature swine. Teratology, 18, 55-62

Shin JS, Lee KG, Lee HH, Lee HJ, An HJ, Nam JH, Jang DS and Lee KT, 2016. alpha-Solanine Isolated From Solanum Tuberosum L. cv Jayoung Abrogates LPS-Induced Inflammatory Responses Via NF-kappaB Inactivation in RAW 264.7 Macrophages and Endotoxin-Induced Shock Model in Mice. Journal of Cellular Biochemistry, 117, 2327-2339.

Sinden SL and Webb RE, 1972. Effect of variety and location on the glycoalkaloid content of potatoes. American Potato Journal, 49, 334-338.

Sinden SL and Webb RE, 1974. Effect of environment ion glycoalkaloid content of six potato varieties at 39 locations. Technical Bulleting No 1472. Agricultural Research Service. United States Department of Agriculture.

Sinden SL, Sanford LL and Webb RE, 1984. Genetic and environmental control of potato glycoalkaloids. American Potato Journal, 61, 141-155.

Sizer CE, Maga JA and Craven CJ, 1980. Total glycoalkaloids in potatoes and potato chips. Journal of Agricultural and Food Chemistry, 28, 578-579.

Skarkova J, Ostry V and Ruprich J, 2008. Determination of Glycoalkaloids (α-solanine and α-chaconine) in Peeled Potato Tubers Using High-Performance Thin-Layer Chromatography. Journal of Planar Chromatography - Modern TLC, 21, 113-117.

Skrabule I, Muceniece R and Kirhnere I, 2013. Evaluation of Vitamins and Glycoalkaloids in Potato Genotypes Grown Under Organic and Conventional Farming Systems. Potato Research, 56, 259-276.

Smith DB, Roddick JG and Jones JL, 1996. Potato glycoalkaloids: some unanswered questions. Trends in Food Science and Technology, 7, 126-131.

Smith SW, Giesbrecht E, Thompson M, Nelson LS and Hoffman RS, 2008. Solanaceous steroidal glycoalkaloids and poisoning by Solanum torvum, the normally edible susumber berry. Toxicon, 52, 667-676.

5002 Sotelo A and Serrano B, 2000. High-Performance Liquid Chromatographic Determination of the Glycoalkaloids r-
5003 Solanine and r-Chaconine in 12 Commercial Varieties of Mexican Potato. Journal of Agricultural and Food
5004 Chemistry, 48, 2472-2475.
5005 Stanker LH, Kamps-Holtzapple C and Friedman M, 1994. Development and characterization of monoclonal
5006 antibodies that differentiate between potato and tomato glycoalkaloids and aglycons. Journal of Agricultural
5007 and Food Chemistry, 42, 2360-2366.
5008 Stein HH, 2002. Experience of feeding pigs without antibiotics: A European Perspective. Animal Biotechnology, 13,
5009 85-95.
5010 Sternfeld M, Rachmilewitz J, Loewenstein-Lichtenstein Y, Andres C, Timberg R, BenAri S, Glick D, Soreq H and
5011 Zakut H, 1997. Normal and atypical butyrylcholinesterases in placental development, function, and
5012 malfunction. Cellular and Molecular Neurobiology, 17, 315-332.
5013 Stoyanova et al 1988. Changes in the content of alkaloidal glycosides during processing of tomatoes. Pishchevaya
5014 Promyshlennost (Moscow, Russian Federation), 10, 55-56.
5015 Sucha L, Hroch M, Rezacova M, Rudolf E, Havelek R, Sispera L, Cmielova J, Kohlerova R, Bezrouk A, Tomsik P,
5016 2013. The cytotoxic effect of α-tomatine in MCF-7 human adenocarcinoma breast cancer cells depends on its
5017 interaction with cholesterol in incubation media and does not involve apoptosis induction. Oncology Reports,
5018 30, 2593-2602.
5019 Surjawan I, Dougherty MP, Bushway RJ, Bushway AA, Briggs JL and Camire ME, 2001. Sulfur Compounds Reduce
5020 Potato Toxins during Extrusion Cooking. Journal of Agricultural and Food Chemistry, 49, 2835-2838.
5021 Swain AP, Fitzpatrick TJ, Talley EA, Herb SF and Osman SF, 1978. Anzymatic hydrolysis of a-chaconine and cc-
5022 solanine. Phytochemistry, 17, 800-801.
5023 Swedish NFA (Swedish National Food Agency), 2016. Samordnade kontrollprojekt 2015 – Glykoalkaloider, kadmium
5024 och bly i potatis. Livsmedelsverkets rapportserie nr 23/2015.
5025 Swinyard CA and Chaube S, 1973. Are potatoes teratogenic for experimental animals. Teratology, 8, 349-357.
5026 Taher D, Solberg SØ, Prohens J, Chou Y-Y, Rakha M and Wu T-H, 2017. World vegetable center eggplant collection:
5027 origin, composition, seed dissemination and utilization in breeding. Frontiers in Plant Science, 8, Article 1484.
5028 Tajner-Czopek A, Rytel E, Aniolowska M and Hamouz K, 2014. The influence of French fries processing on the
5029 glycoalkaloid content in coloured-fleshed potatoes. European Food Research and Technology, 238, 895-904.
5030 Takagi K, ToyodaM, Fujiyama Y and Saito Y, 1990. Effect of cooking on the contents of α-chaconine and α-solanine
5031 in potatoes. Shokuhin Eiseigaku Zasshi, 31, 67-73.
5032 Tamasi G, Pardini A, Bonechi C, Donati A, Pessina F and Marcolongo P, 2019. Characterization of nutraceutical
5033 components in tomato pulp, skin and locular gel. European Food Research and Technology, 245, 907–918.
5034 Temperton H, 1943. The effects of feeding green and sprouted potatoes to laying pullets. Veterinary Record, 55,
5035 359.
5036 Tepe EJ, Anderson GJ, Spooner DM and Bohs L, 2016. Relationships among wild relatives of the tomato, potato
5037 and pepino. Taxon, 65, 262-276.
5038 Tice RR, 1998. α-Chaconine [20562-03-2] and α-Solanine [20562-02-1]. Review of toxicological literature.
5039 Integrated Laboratory Systems, 103.
5040 Tömösközi-Farkas R, Daood HG, Polgár ZS and Hajós GY, 2006. Determination of Glycoalcaloids in Hungarian
5041 Potatoes by HPLC. Chromatographia Supplement, 63, 115-118.
5042 Tömösközi-Farkas R, Polgár ZS, Nagy-Gasztonyi M, Horváth V, Renkecz T, Simon K, Boross F, Fabulya Z and Daood
5043 H, 2014. Changes of potentially antinutritive components in 8 Hungarian potatoes under organic and
5044 conventional farming. Acta Alimentaria, 43, 676-683.
5045 Tsiaoussis J, Hatzidaki E, Docea AO, Nikolouzakis, TK, Petrakis D, Burykina, T, Mamoulakis C, Makrigiannakis A and
5046 Tsatsakis A, 2018. Molecular and clinical aspects of embryotoxicity induced by acetylcholinesterase inhibitors.
5047 Toxicology, 409, 137-143.
5048 Turner NJ and Kuhnlein HV, 1983. Camas (Camasia spp.) and Riceroot (Fritillaria spp.): two Liliaceous 'root' foods
5049 of the Northwest Coast Indians. Ecology of Food and Nutrition, 13, 199-219.
5050 Tusche T, Wuertz S, Susenbeth A and Schulz C, 2011. Feeding fish according to organic aquaculture guidelines
5051 EC710/2009: Influence of potato protein concentrates containing various glycoalkaloid levels on health status
5052 and growth performance of rainbow trout (*Oncorhynchus mykiss*). Aquaculture, 319, 122-131.
5053 Tuśnio A, Pastuszewska B, Taciak M, Barszcz M and Skomiał J, 2013a. The response of rats to solanidine
5054 glycoalkaloids and trypsin inhibitor present in potato protein concentrates, and to glycoalkaloids provided by
5055 potato sprouts. Journal of Animal and Feed Sciences, 22, 130-136.
5056 Tuśnio A, Pastuszewska B, Taciak M, Mieczkowska A and Smulikowska S, 2013b. Response of growing chicken to
5057 potato protein concentrates providing different amounts of solanidine glycoalkaloids and trypsin inhibitor.
5058 Archiv fűr Geflűgelkunde, 77, 51-58.
5059 University of Chemistry and Technology Prague, 2019. Extensive literature search and selection for relevance of
5060 studies related to the chemistry and toxicity of glycoalkaloids and quinolizidine alkaloids in food and feed.
5061 https://doi.org/10.2903/sp.efsa.2019.en-1348
5062 Urban J, Hamouz K, Lachman J, Pulkrábek J and Pazderů K, 2018. Effect of genotype, flesh colour and environment
5063 on the glyco-alkaloid content in potato tubers from integrated agriculture. Plant, Soil and Environment, 64,
5064 186–91.

5065 USDA (United States Department of Agriculture), 2011. Plnts poisonous to livestock in the Western States.
5066 Agricultural Research Service Information Bulletin 415.
5067 Valcárcel J, Reilly K, Gaffney M and O'Brien N, 2014. Effect of Genotype and Environment on the Glycoalkaloid
5068 Content of Rare, Heritage, and Commercial Potato Varieties. Journal of Food Science, 79, 1039-1048.
5069 Valkonen JPT, Keskitalo M, Vasara T and Pietila L, 1996. Potato glycoalkaloids: a burden or a blessing? Critical
5070 Reviews in Plant Sciences, 15, 1-20.
5071 Van Denburgh RW, Hiller L-K- and Koller DC, 1986. Ultrastructural Changes in Potato Tuber Pith Cells during Brown
5072 Center Development. Plant Physiology, 81, 167-170.
5073 Van Gelder WMJ, 1985. Determination of the total C27-steroidal alkaloid composition of Solanum species by high-
5074 resolution gas chromatography. Journal of Chromatography A, 331, 285-293.
5075 Van Gelder WMJ, 1989. Steroidal glycoalkaloids in Solanum species: consequences for potato breeding and food
5076 safety. Van Gelder, Dissertation. Centrale landbouwcatalogus 000 0352 3277, ISN 506943.
5077 Van Gelder WMJ, 1991. Chemistry, toxicology, and occurrence of steroidal glycoalkaloids: potential contaminants
5078 of the potato. In Poisonous Plant Contamination of Edible Plants; Rizk, A. F. M., Ed.; CRC Press: Boca Raton,
5079 FL, 1991; pp 121-143. pp.
5080 Van Gelder WMJ, Vinket JH and Scheffer JJC, 1988. Steroidal glycoalkaloids in tubers and leaves of *Solanum* species
5081 used in potato breeding. Euphytica, 39, 147-158.
5082 Van Gelder WMJ and De Ponti OMB, 1987. α-Tomatine and other steroidal glycoalkaloids in fruits of tomato lines
5083 resistant to the glasshouse whitefly (*Trialeurodes vaporariorum* Westw.). Euphytica, 36, 555-561.
5084 Viollier R, 1941. Lebensmittelkontrolle und Berichte von Untersuchungsämtern. Zeitschrift fuer Untersuchung der
5085 Lebensmittel, 192.
5086 Vogt H and Stute K, 1969. Dried ground potatoes in poultry rations. II. Solanine determination and effects of
5087 solanine. Archiv fűr Geflűgelkunde, 33, 316-320.
5088 Vogel-Höpker A, Sperling LE and Layer PG, 2012. Co-opting Functions of Cholinesterases in Neural, Limb and Stem
5089 Cell Development. Protein and Peptide Letters, 19, 155-164.
5090 Voldrich M, Ondroušek S and Dobiáš J, 1992. Steroid glycoalkaloids in fresh and processed tomatoes. Potrav. Vêdy,
5091 10, 23-30.
5092 Vorontsova MS, Stern S, Bohs L and Knapp S, 2013. African spiny Solanum subgenus Leptostemonum, Solanaceae):
5093 a thorny phylogenetic tangle. Botanical Journal of the Linnean Society, 173, 176-193.
5094 Wang F, Yang YF and Li PZ, 2008. A case-control study on the risk factors of neural tube defects in Shanxi province.
5095 Zhonghua Liu Xing Bing Xue Za Zhi, 29, 771-4. In Chinese, abstract in English.
5096 Whittemore CT, 1977. The potato (*Solanum tuberosum*) as a source of nutrients for pigs, calves and fowl — A
5097 review. Animal Feed Science and Technology, 2, 171-190.
5098 WHO/IPCS (World Health Organisation/International Programme on Chemical Safety), 2008. Uncertainty and data
5099 quality in exposure assessment. Harmonisation project document No. 6. ISBN 978924156376. Available
5100 online: https://apps.who.int/iris/handle/10665/44017
5101 WHO/IPCS (World Health Organisation/International Programme on Chemical Safety), 2009. Principles and
5102 methods for the risk assessment of chemicals in food. Environmental Health Criteria 240. International
5103 Programme on Chemical Safe. ISBN: 9789241572408. Available online:
5104 https://www.who.int/foodsafety/publications/chemical-food/en/
5105 Willimott SG, 1933. An investigation of solanine poisoning. Analyst, 58, 431-439.
5106 Wilson GS, 1959. A small outbreak of solanine poisoning. Monthly bulletin of the Ministry of Health and the Public
5107 Health Laboratory Service, 18, 207-210.
5108 Wilson RH, Poley GW and De Eds F, 1961. Some pharmacologic and toxicologic properties of tomatine and its
5109 derivatives. Toxicology and Applied Pharmacology, 3, 39-48.
5110 Wilson AM, McGann DF and Bushway RJ, 1983. Effect of growth-location and length of storage on glycoalkaloid
5111 content of roadside-stand potatoes as stored by consumers. Journal of Food Protection, 46, 119-121.
5112 Wood F and Young D, 1974. TGA in potatoes. Agriculture Canada, 1533, 3.
5113 Xie W, Stribley JA, Chatonnet A, Wilder PJ, Rizzino A, McComb RD, Taylor P, Hinrichs SH and Lockridge O,
5114 2000. Postnatal developmental delay and supersensitivity to organophosphate in gene-targeted mice lacking
5115 acetylcholinesterase. Journal of Pharmacology and Experimental Therapeutics, 293, 896-902.
5116 Zarzecka K and Gugala M, 2007. Changes in the content of glycoalkaloids in potato tubers according to soil tillage
5117 and weed control methods. Plant, Soil and Environment, 53, 247-251.
5118 Zhang L, Li M, Zhang G, Wu L, Cai D and Wu Z, 2018. Inhibiting Sprouting and Decreasing α-Solanine Amount of
5119 Stored Potatoes Using Hydrophobic Nanosilica. ACS Sustainable Chemistry and Engineering, 6, 10517-10525.
5120 Zitnak A, 1961. The occurrence and distribution of free alkaloid solanidine in netted gem potatoes. Canadian Journal
5121 of Biochemistry and Physiology, 39, 1257-1265.
5122 Zwenger C and Kind A, 1861. Ueber das Solanin und dessen Spaltungs-producte. Ann. Chem. Pharm., 118, 129-
5123 151.

5124 ## Abbreviations

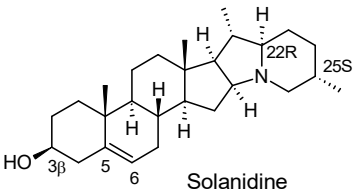
ACh	Acetylcholine
AChE	Acetylcholinesterase
ALT	Alanine aminotransferase
Anses	French Agency for Food, Environmental and Occupational Health and Safety
AOAC	Association of Official Analytical Chemists
AST	Aspartate aminotransferase
AUC_{0-24}	Areas Under the Curve (0–24 h)
$AUC_{0-\infty}$	Areas Under the Curve (from point zero to infinity)
BfR	The German Federal Institute for Risk Assessment
BuChE	Butyrylcholinesterase
bw	Body weight
ChEs	Cholinesterases
CI	Confidence interval
C_{max}	Maximum serum concentrations
CNS	Central nervous system
CONTAM Panel	Panel on Contaminants in the Food Chain
dw	Dry weight
ECG	Electrocardiogram
EEG	electroencephalogram
EFSA	European Food Safety Authority
ELISA	Enzyme-linked immunosorbent assay
EU	European Union
F	female
FAO	Food and Agricultural Organization
FDA	Food and Drug Administration
FID	Flame ionization detector
fw	Fresh weight
GAs	Glycoalkaloids
GC	Gas Chromatography
GD	Gestational day
GOT	Glutamic oxaloacetic transaminase
GPT	Glutamic pyruvic transaminase
h	hour
HDL	High-density lipoprotein
HGB	Haemoglobin
HMGR	3-hydroxy-3-methylglutaryl coenzyme A reductase
HPLC	High-performance liquid chromatography
HPTLC	High performance thin layer chromatography
HTC	Haematocrit
IPCS	International Programe on Chemical Safety
i.p.	Intraperitoneal injection
i.v.	Intravenous injection
IFN-γ	Interferon-gamma
IL-1β	Interleukin 1 beta
JECFA	Joint FAO/WHO Expert Committee on Food Additives
K_i	Inhibitory constants
LB	Lower bound
LC	Liquid chromatography
LD_{50}	Lethal dose, median
LDL	Low-density lipoproteins
LOEL	Lowest-observed-effect level
LOAEL	Lowest-observed-adverse-effect level
LOD	Limit of detection
LOQ	Limit of quantification
LPS	Lipopolysaccharide
M	male

MOE	Margin of exposure
MS	Mass spectrometry
MS/MS	Tandem mass spectrometry
MTHFR	Methylenetetrahydrofolate reductase
nAChR	Specific acetylcholine-receptor
$NaHSO_3$	Sodium bisulfite
NAK	Nederlandse Algemene Keuringsdienst (Dutch General Inspection Service)
NOEL	No-observed-effect level
NOAEL	No-observed-adverse-effect level
NPD	Nitrogen Phosphorous Detector
NTD	Neural tube defects
OECD	The Organisation for Economic Co-operation and Development
OFCs	Orofacial clefts
PAD	Pulsed amperometric detector
PCE	Polychromatic erythrocytes
p.o.	per oral
RASFF	Rapid Alert System for Food and Feed
RBC	Red blood cell
RPC	Raw primary commodity
s.c.	subcutaneous
SGT	Solanidine glycosyltransferase
SMT	Sterol C24-methyltransferase
SOPs	Standard operational procedures
SPE	Solid-phase extraction
SS	Squalene synthase
TGA	Total glycoalkaloids
TLC	Thin layer chromatography
TNF-α	Tumor necrosis factor-alpha
UB	Upper bound
UK	United Kingdom
USA	United States of America
UV	Ultraviolet
UV-vis	Ultraviolet-visible
VLDL	Very low-density lipoproteins
Voedingscentrum	The Netherlands Nutrition Centre Foundation
VSP	Vetispiradiene cyclase
WBC	White blood cells
WHO	World Health Organization
ww	Wet weight

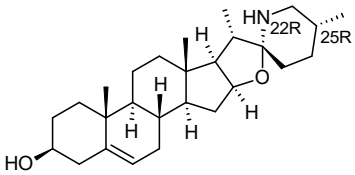
5125

Appendix A - Major glycoalkaloids and their aglycones present in Solanum species

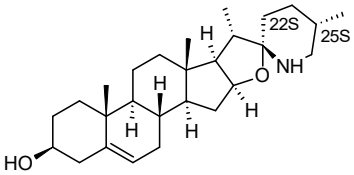
A. The main aglycones found in *Solanum* species considered in this Scientific Opinion



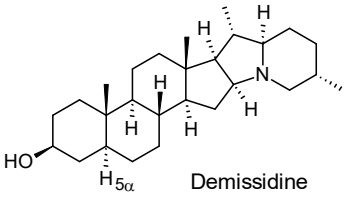
Solanidine



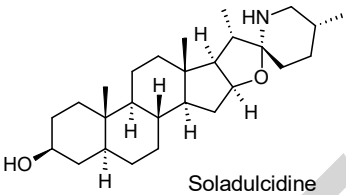
Solasodine



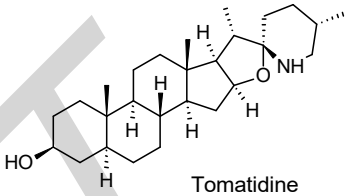
Tomatidenol



Demissidine

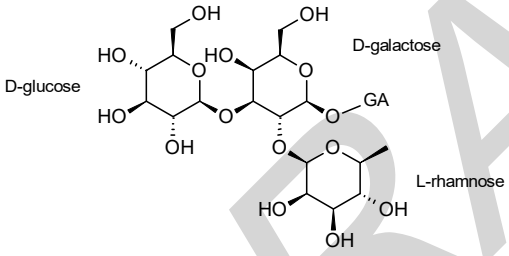


Soladulcidine

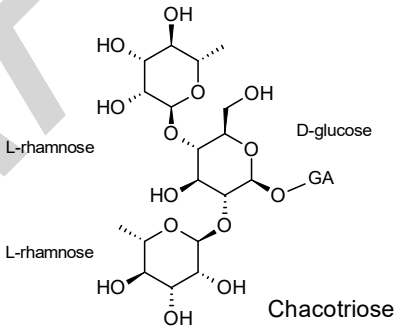


Tomatidine

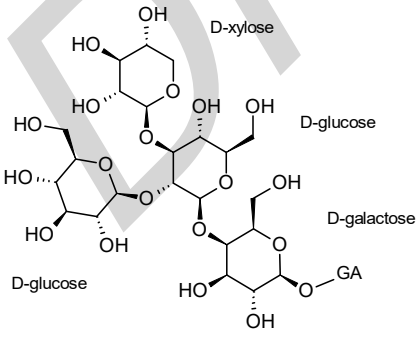
B. The main sugar moieties found in *Solanum* species considered in this Scientific Opinion



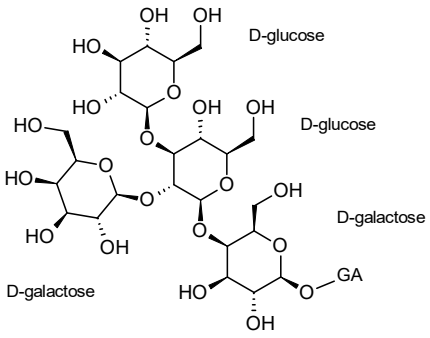
Solatriose



Chacotriose

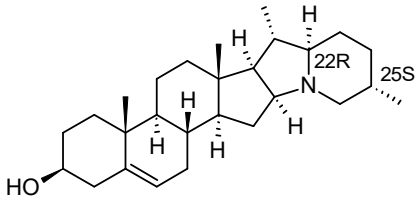
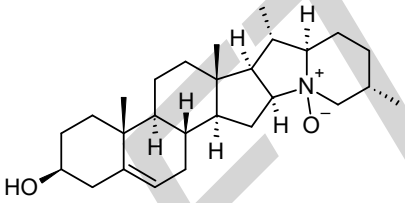
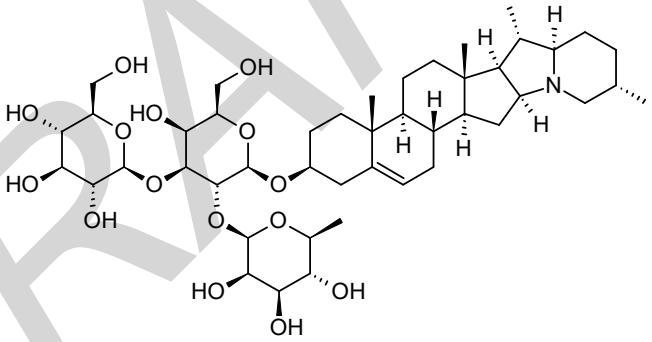
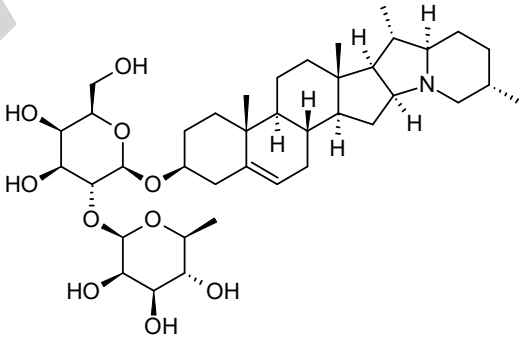
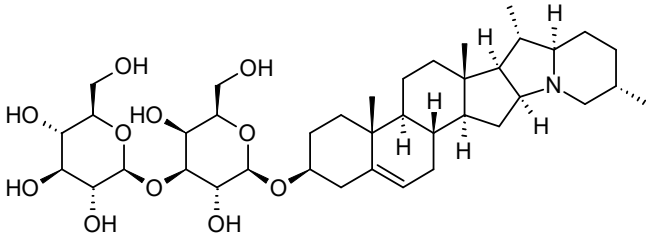


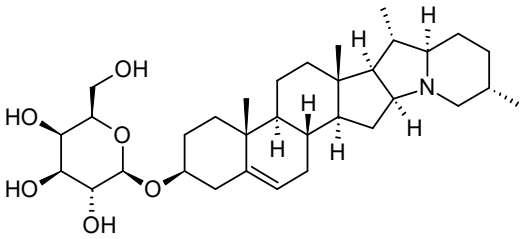
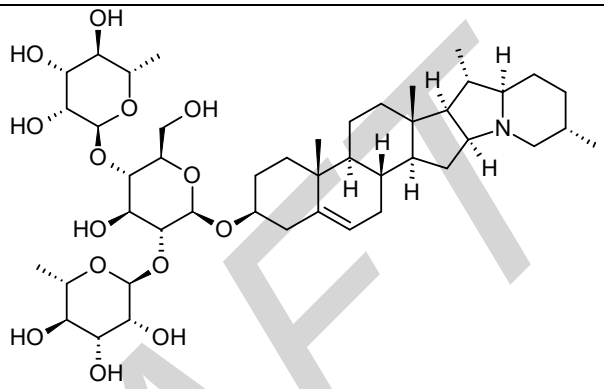
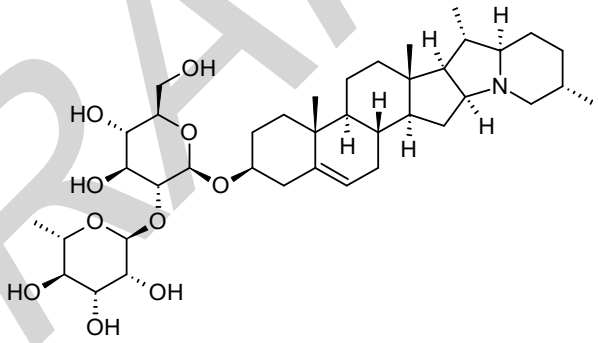
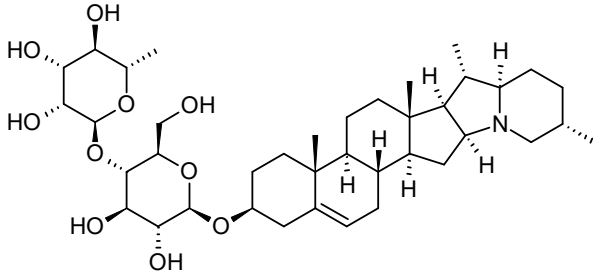
Lycotetraose

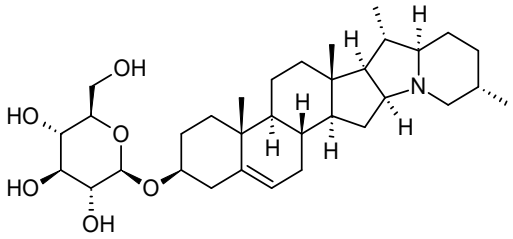
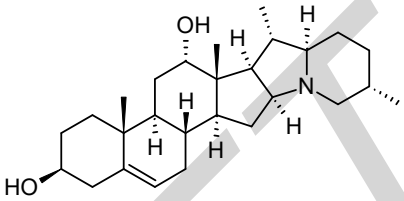
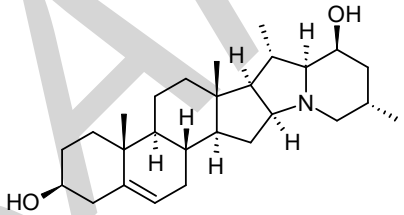
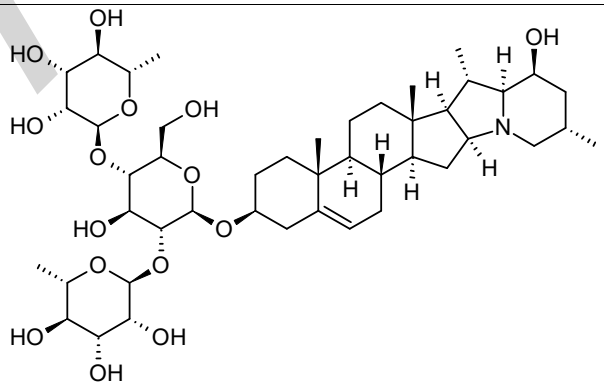


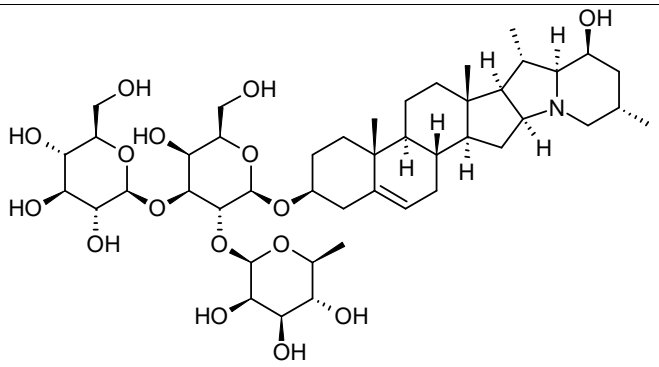
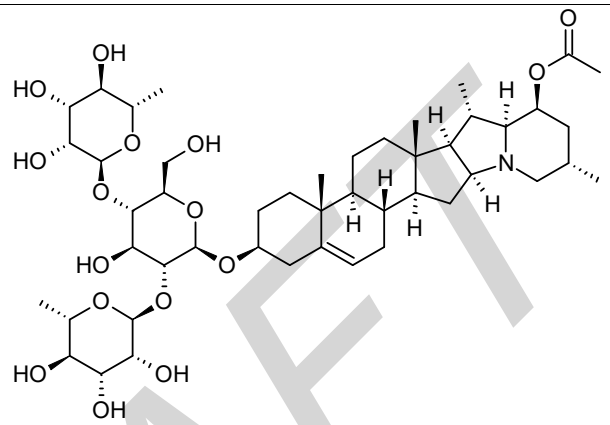
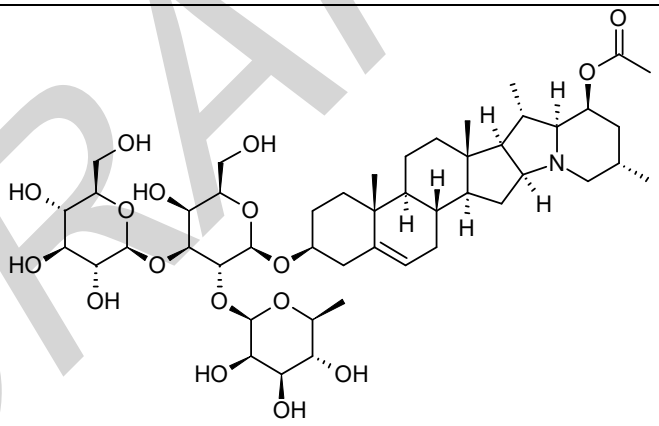
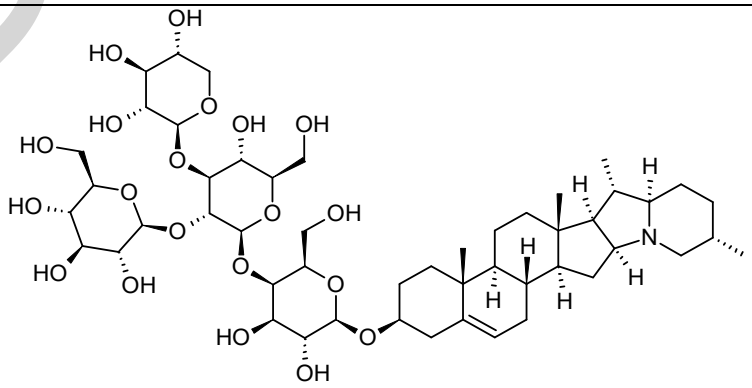
Commertetraose

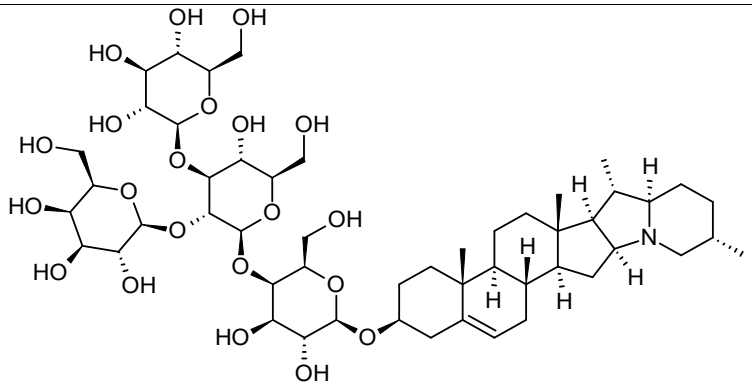
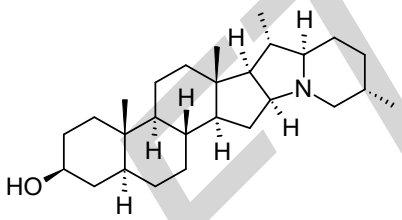
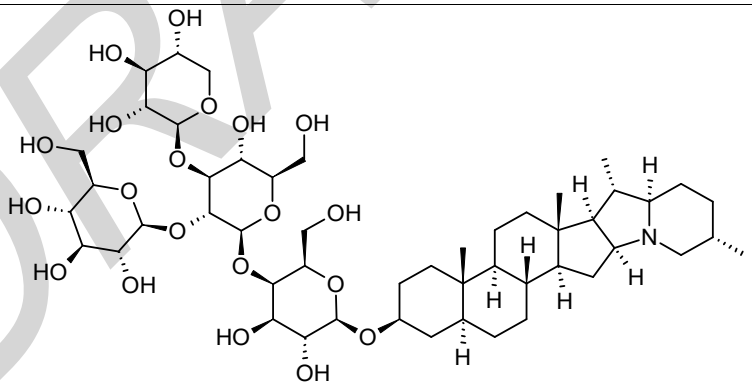
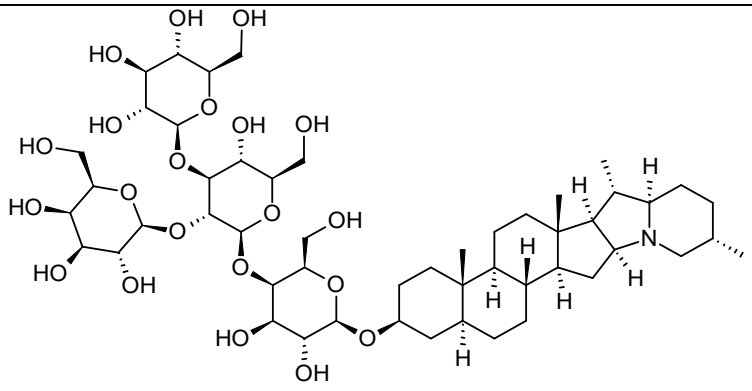
C. Glycoalkaloids considered in this Scientific Opinion: structures and physico-chemical properties

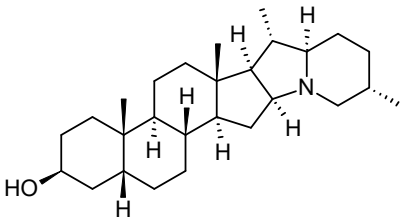
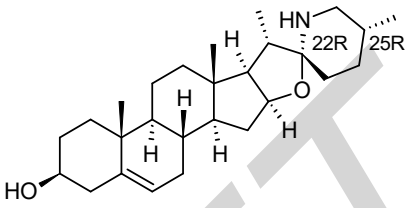
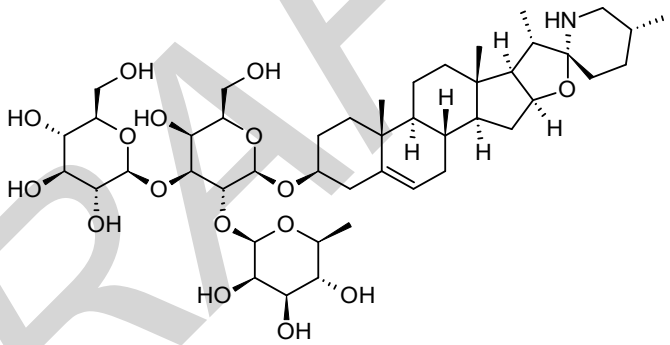
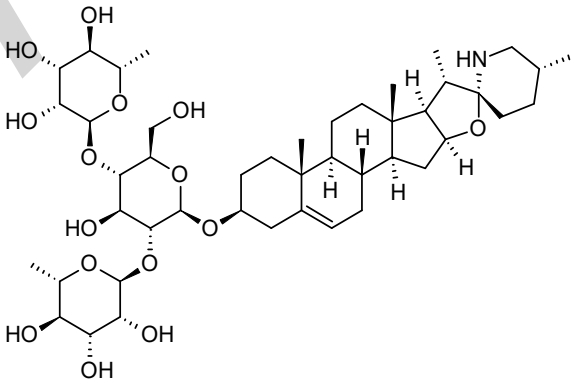
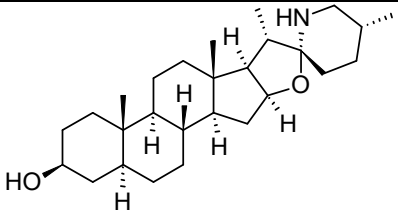
Solanidine Synonym: solatubine CAS No: 80-78-4 Sugar moiety: none, aglycone $C_{27}H_{43}NO$ MW: 397.647 LogP (est): 6.1 Solubility (est): 0.0004 g/L	
Solanidine N-oxide Sugar moiety: none, aglycone $C_{27}H_{43}NO_2$ MW: 413.636 LogP (est) : 5.5	
α-Solanine Synonym: solatunine CAS No: 20562-02-1 Sugar moiety: solatriose $C_{45}H_{73}NO_{15}$ MW: 868.071 LogP (est): 1.8 Solubility (est): 0.62 g/L	
β_1-Solanine CAS No: 142287-76-1 Sugar moiety: galactose-rhamnose $C_{39}H_{63}NO_{10}$ MW: 705.930 LogP (est): 3.4	
β_2-Solanine CAS No: 61877-94-9 Sugar moiety: galactose-glucose $C_{39}H_{63}NO_{11}$ MW: 721.929	

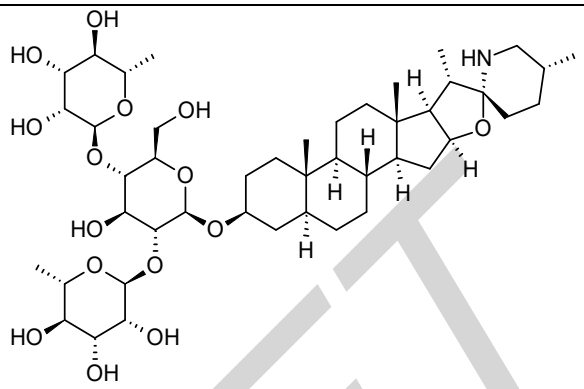
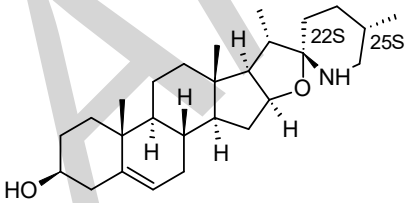
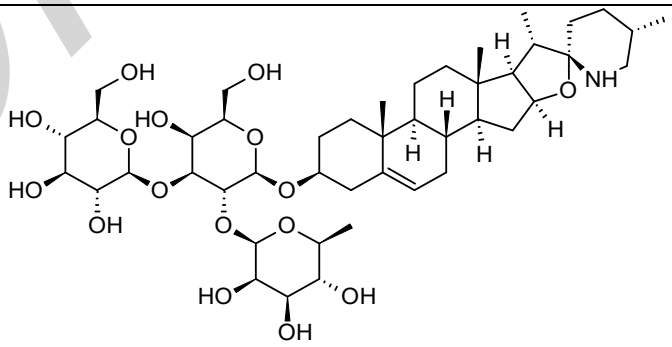
LogP (est): 2.9 Solubility (est): 0.25 g/L	
γ-Solanine CAS No: 511-37-5 Sugar moiety: galactose $C_{33}H_{53}NO_6$ MW: 559.788 LogP (est): 4.5 Solubility (est): 0.021 g/L	
α-Chaconine CAS No: 20562-03-2 Sugar moiety: chacotriose $C_{45}H_{73}NO_{14}$ MW: 852.072 LogP (est): 1.8 Solubility (est): 0.29 g/L	
β₁-Chaconine CAS No: 472-51-5 Sugar moiety: rhamnose-glucose $C_{39}H_{63}NO_{10}$ MW: 705.930 LogP (est): 3.4 Solubility (est): 0.092 g/L	
β₂-Chaconine CAS No: 469-14-7 Sugar moiety: rhamnose-glucose $C_{39}H_{63}NO_{10}$ MW: 705.930 LogP (est): 2.8 Solubility (est): 0.088 g/L	

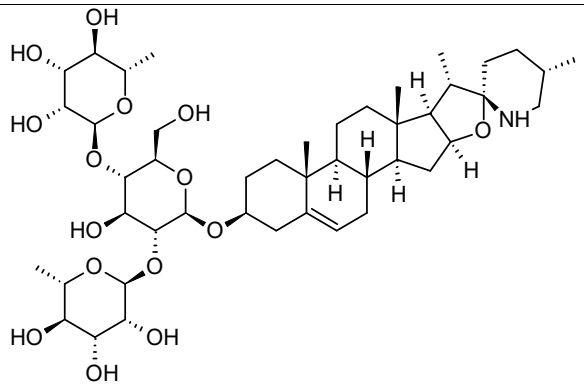
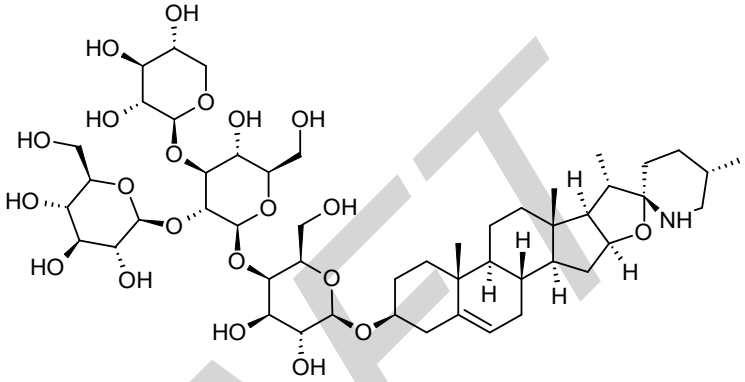
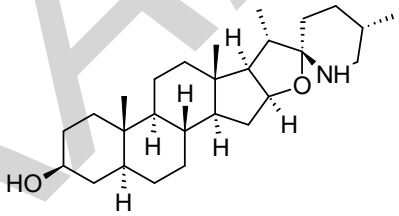
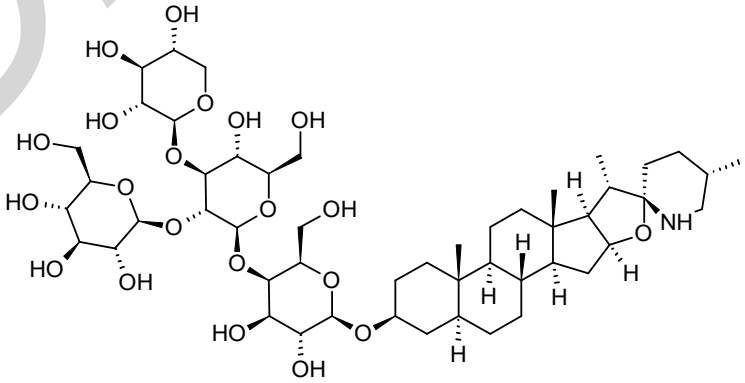
γ-Chaconine CAS No: 511-36-4 Sugar moiety: glucose $C_{33}H_{53}NO_6$ MW: 559.788 LogP (est): 4.5 Solubility (est): 0.021 g/L	
Rubijervine Synonym: 12α-hydroxysolanidine CAS No: 79-58-3 Sugar moiety: none, aglycone $C_{27}H_{43}NO_2$ MW: 413.646 LogP (est): 4.7	
Leptinidine Synonym: 23β-hydroxysolanidine CAS No: 24884-17-1 Sugar moiety: aglycone $C_{27}H_{43}NO_2$ MW: 413.646 LogP (est): 5.1	
Leptinine I CAS No: 101009-59-0 Sugar moiety: chacotriose $C_{45}H_{73}NO_{15}$ MW: 868.071 LogP (est): 0.8	

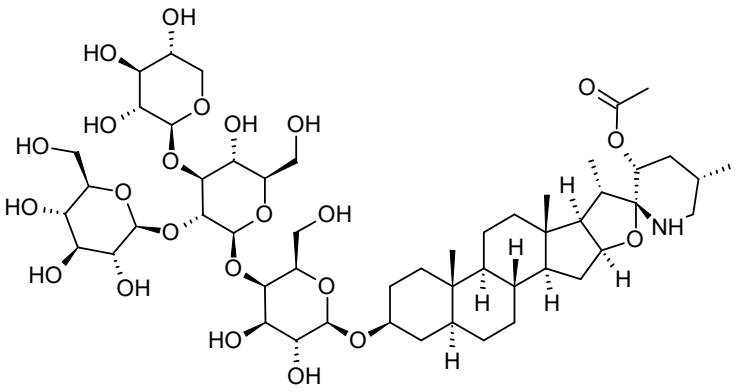
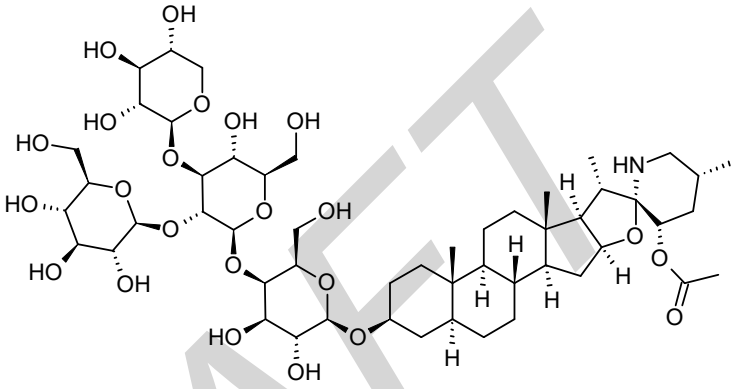
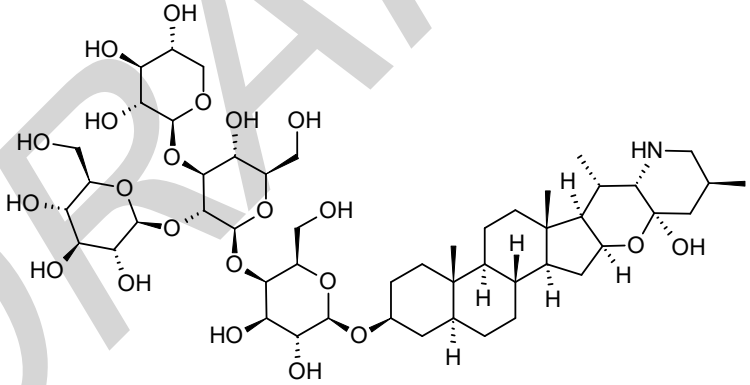
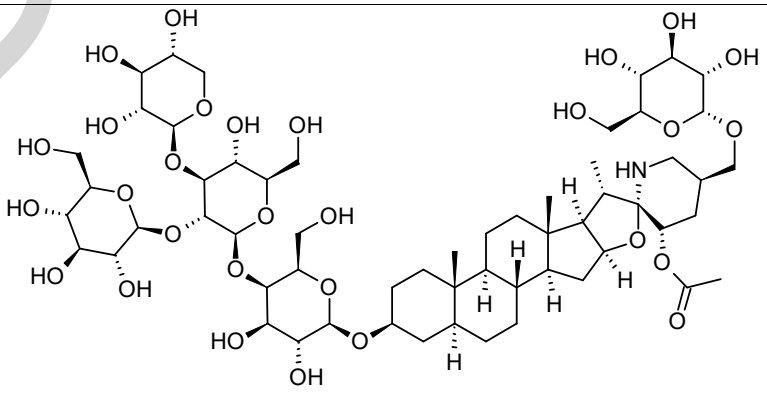
Leptinine II CAS No: 100994-57-8 Sugar moiety: solatriose $C_{45}H_{73}NO_{16}$ MW: 884.070 LogP (est): 0.8	
Leptine I CAS No: 101030-83-5 Sugar moiety: chacotriose $C_{47}H_{75}NO_{16}$ MW: 910.108 LogP (est): 1.4	
Leptine II CAS No: 101054-39-1 Sugar moiety: solatriose $C_{47}H_{75}NO_{17}$ MW: 926.107 LogP (est): 1.4	
Dehydrodemissine CAS No: 195433-57-9 Sugar moiety: lycotetraose $C_{50}H_{81}NO_{20}$ MW: 1016.185 LogP (est): -0.8 Solubility (est): 1.69 g/L	

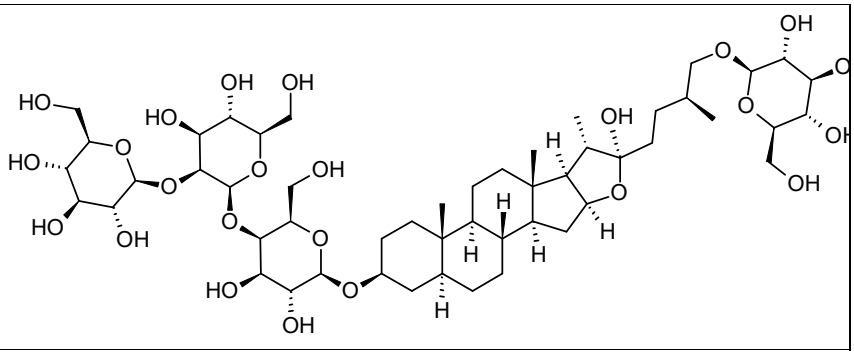
Dehydrocommersonine CAS No: 65428-74-2 Sugar moiety: commertetraose $C_{51}H_{83}NO_{21}$ MW: 1046.211 LogP (est): -0.8 Solubility (est): 1.69 g/L	
Demissidine Synonym: 5α,6-dihydrosolanidine, 5α-Solanidan-3β-ol CAS No: 474-08-8 Sugar moiety: none, aglycone $C_{27}H_{45}NO$ MW: 399.663 LogP (est): 6.9 Solubility (est): 0.00022 g/L	
Demissine CAS No: 6077-69-6 Sugar moiety: lycotetraose $C_{50}H_{83}NO_{20}$ MW: 1,018.201 LogP (est): 0.0 Solubility (est): 1.63 g/L	
Commersonine CAS No: 60776-42-3 Sugar moiety: commertetraose $C_{51}H_{85}NO_{21}$ MW: 1,048.227 LogP (est): 0.0 Solubility (est): 1.63 g/L	

5β-solanidan-3β-ol Sugar moiety: none, aglycone $C_{27}H_{45}NO$ MW: 399.663	
Solasodine Synonym: purapuridine CAS No: 126-17-0 Sugar moiety none: aglycone $C_{27}H_{43}NO_2$ MW: 413.646 LogP (est): 5.4 Solubility (est): 0.00084 g/L	
α-Solasonine Synonym: purapurine, γ-solanigrine CAS No: 19121-58-5 Sugar moiety: solatriose $C_{45}H_{73}NO_{16}$ MW: 884.070 LogP (est): 1.1 Solubility (est): 0.41 g/L	
α-Solamargine Synonym: δ-solanigrine CAS No: 20311-51-7 Sugar moiety: chacotriose $C_{45}H_{73}NO_{15}$ MW: 868.071 LogP (est): 1.1 Solubility (est): 0.2 g/L	
Soladulcidine Synonym: solasodan-3β-ol, dihydrosolasodine CAS No: 511-98-8	

Sugar moiety: none aglycone $C_{27}H_{45}NO_2$ MW: 415.662 LogP (est): 6.2 Solubility (est): 0.00019 g/L	
Soladulcine A CAS No: 156555-52-1 Sugar moiety: chacotriose $C_{45}H_{75}NO_{15}$ MW: 870.087 LogP (est):	
Tomatidenol Synonym: dehydrotomatidine CAS No: 546-40-7 Sugar moiety: none, aglycone $C_{27}H_{43}NO_2$ MW: 884.07 LogP (est): 5.4 Solubility (est): 0.00084 g/L	
α-Solamarine CAS No: 20318-30-3 Sugar moiety: solatriose $C_{45}H_{73}NO_{16}$ MW: 884.07 LogP (est): 1.1 Solubility (est): 0.38 g/L	

β-Solamarine CAS No: 3671-38-3 Sugar moiety: chacotriose $C_{45}H_{73}NO_{15}$ MW: 868.071 LogP (est): 1.1 Solubility (est): 0.16 g/L	
α-Dehydrotomatine CAS No: 157604-98-3 Sugar moiety: lycotetraose $C_{50}H_{81}NO_{21}$ MW: 1032.184 LogP (est): -1.5 Solubility (est): 1.82 g/L	
Tomatidine CAS No: 77-59-8 Sugar moiety: none, aglycone $C_{27}H_{45}NO_2$ MW: 415.662 LogP (est): 6.2 Solubility (est): 0.00019 g/L	
α-Tomatine CAS No: 17406-45-0 Sugar moiety: lycotetraose $C_{50}H_{83}NO_{20}$ MW: 1034.188 LogP (est): -0.7 Solubility (est): 1.71 g/L	

Lycoperoside A Syn: 23R-Acetoxytomatine CAS No: 176181-33-2 Sugar moiety: lycotetraose $C_{52}H_{85}NO_{23}$ MW: 1092.224 LogP (est): -1.1	
Lycoperoside B Syn: 23S-Acetoxysoladulcidine CAS No: 176300-86-0 Sugar moiety: lycotetraose $C_{52}H_{85}NO_{23}$ MW: 1092.224 LogP (est): -1.1	
Lycoperoside H CAS No: 675828-28-1 Sugar moiety: lycotetraose $C_{50}H_{83}NO_{22}$ MW: 1050.188 LogP (est): 1.2	
Esculeoside A CAS No: 532387-86-3 Sugar moiety: lycotetraose and glucose $C_{58}H_{95}NO_{29}$ MW: 1270.376 LogP (est): -3.9	

Tomatoside A CAS No: 75557-22-1 Sugar moiety: triose and glucose $C_{51}H_{86}O_{24}$ MW: 1084.210 LogP (est): -1.4	

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Appendix B - Identification and selection of evidence relevant for the risk assessment of glycoalkaloids in feed and food

In addition to the literature search outsourced by EFSA (see University of Chemistry and Technology Prague, 2019 for further details), some specific searches were performed to identify the scientific literature.

B.1 Literature search for hazard identification and characterisation

Adverse effects of glycoalkaloids in humans

A literature search was performed to retrieve data on the adverse effects of glycoalkaloids in humans. Three databases were used: Web of Science, Pubmed and Scopus; the search was performed without limits regarding the year/type of publication, and language. All the references were uploaded in Endnote. After removal of duplicates, the final number of hits is 330.

Solanine

Web of science

TOPIC: (solanin* OR 20562-02-1 OR 243-879-8) AND TOPIC: (adverse effect* OR epidemiolog* OR "case control" OR case report* OR case stud* OR clinical trial* OR clinical stud* OR cohort OR "cross sectional" OR human stud* OR human volunteer*)
Results: 276

Pubmed

(((solanin[All Fields] OR solanina[All Fields] OR solaninase[All Fields] OR solaninaza[All Fields] OR solaninbestimmung[All Fields] OR solanine[All Fields] OR solaninlosung[All Fields] OR solaninneubildung[All Fields] OR solaninom[All Fields] OR solaninum[All Fields] OR solaninvergiftung[All Fields]) OR "20562-02-1"[All Fields])) AND ("Epidemiologic Studies"[Mesh] OR "Drug-Related Side Effects and Adverse Reactions"[Mesh] OR "Volunteers"[Mesh] OR "Human Experimentation"[Mesh] OR "Clinical Study" [Publication Type] OR "Clinical Studies as Topic"[Mesh] OR "Case Reports" [Publication Type] OR epidemiolog*[tiab] OR "case control"[tiab] OR case stud*[tiab] OR cohort[tiab] OR "cross sectional"[tiab] OR adverse effect*[tiab] OR "human volunteers"[tiab] or human stud*[tiab])
Results: 11

Scopus

(TITLE-ABS-KEY ((solanin* OR "20562-02-1" OR "243-879-8")) AND TITLE-ABS-KEY ((human AND adverse AND effect*) OR (epidemiolog*) OR (case AND control) OR (case AND report*) OR (case AND stud*) OR (clinical AND trial*) OR (clinical AND stud*) OR (cohort) OR (cross AND sectional) OR (human AND stud*) OR (human AND volunteer*)))
Results: 144

All the retrieved references were uploaded in Endnote. Final number of hits after removal of duplicates: 356

Chaconin

Web of science

TOPIC: ((chaconin* OR 20562-03-2 OR 472-51-5)) AND TOPIC: (adverse effect* OR epidemiolog* OR case control OR case report* OR case stud* OR clinical trial* OR clinical stud* OR cohort OR cross sectional OR human stud* OR human volunteer*)
Results: 172

Pubmed

(((chaconin[All Fields] OR chaconine[All Fields] OR chaconinverhaltnisses[All Fields]) OR "20562-03-2"[All Fields] OR "472-51-5"[All Fields])) AND ((("Epidemiologic Studies"[Mesh] OR "Drug-Related Side Effects and Adverse Reactions"[Mesh] OR "Volunteers"[Mesh] OR "Human Experimentation"[Mesh] OR "Clinical Study" [Publication Type] OR "Clinical Studies as Topic"[Mesh] OR "Case Reports" [Publication Type] OR epidemiolog*[tiab] OR "case control"[tiab] OR case stud*[tiab] OR cohort[tiab] OR "cross sectional"[tiab] OR adverse effect*[tiab] OR "human volunteers"[tiab] or human stud*[tiab])))
Results: 7

Scopus

(TITLE-ABS-KEY (chaconin* OR 20562-03-2 OR 472-51-5) AND TITLE-ABS-KEY ((human AND adverse AND effect*) OR (epidemiolog*) OR (case AND control) OR (case AND report*) OR (case AND stud*) OR (clinical AND trial*) OR (clinical AND stud*) OR (cohort) OR (cross AND sectional) OR (human AND stud*) OR (human AND volunteer*)))
Results: 79

All the retrieved references were uploaded in Endnote. Final number of hits after removal of duplicates: 206

Glycoalkaloids

Web of science

TOPIC: (glycoalkaloid*) AND TOPIC: (adverse effect* OR epidemiolog* OR case control OR case report* OR case stud* OR clinical trial* OR clinical stud* OR cohort OR cross sectional OR human stud* OR human volunteer*)
Results: 497

Pubmed

((glycoalkaloid[All Fields] OR glycoalkaloidal[All Fields] OR glycoalkaloidic[All Fields] OR glycoalkaloids[All Fields] OR glycoalkaloidsalpha[All Fields])) AND ((("Epidemiologic Studies"[Mesh] OR "Drug-Related Side Effects and Adverse Reactions"[Mesh] OR "Volunteers"[Mesh] OR "Human Experimentation"[Mesh] OR "Clinical Study" [Publication Type] OR "Clinical Studies as Topic"[Mesh] OR "Case Reports" [Publication Type] OR epidemiolog*[tiab] OR "case control"[tiab] OR case stud*[tiab] OR cohort[tiab] OR "cross sectional"[tiab] OR adverse effect*[tiab] OR "human volunteers"[tiab] or human stud*[tiab])))
Results: 18

Scopus

(TITLE-ABS-KEY (glycoalkaloid*) AND TITLE-ABS-KEY ((human AND adverse AND effect*) OR (epidemiolog*) OR (case AND control) OR (case AND report*) OR (case AND stud*) OR (clinical AND trial*) OR (clinical AND stud*) OR (cohort) OR (cross AND sectional) OR (human AND stud*) OR (human AND volunteer*)))

Results: 167

All the retrieved references were uploaded in Endnote. Final number of hits after removal of duplicates: 541

B.2. Literature search for hazard identification and characterisation

Toxicity of glycoalkaloids

A extra search was conducted including all the relevant glycoalkaloids from the mandate. The search included the search strings listed in the tables below, 3 different search databases were used. The glycoalkaloids were divided into three groups for the search:

- Tomato alkaloids: rubijervin, tomatidine, tomatine, tomatidenol, dehydrotomatine.
- Solanine alkaloids: solasodine, solasonine, solamargine, soladulcine, soladulcidine, solamarine, solanidine.
- Leptin alkaloids: leptinin, leptidine, leptinine I and II, leptine I and II

Web of science

TOPIC: (rubijervin*) OR TOPIC: (tomatidin*) OR TOPIC: (tomatidenol*) OR TOPIC: (tomatin*) OR TOPIC: (dehydrotomatin)) AND (TOPIC: (toxic*) OR TOPIC: (adverse) OR TOPIC: (terato*))
Results: 78

Pubmed

((((((rubijervin*) OR tomatidin*) OR tomatin*) OR tomatidenol) OR dehydrotomatin*)) AND (((toxic*) OR adverse) OR terato*)
Results: 101

Scopus

((TITLE-ABS-KEY (rubijervin*) OR TITLE-ABS-KEY (tomatidin*) OR TITLE-ABS-KEY (tomatin*) OR TITLE-ABS-KEY (tomatidenol*) OR TITLE-ABS-KEY (dehydrotomatin*))) AND ((TITLE-ABS-KEY (toxic*) OR TITLE-ABS-KEY (adverse) OR TITLE-ABS-KEY (terato*)))
Results: 109

a) <u>Solanine alkaloids</u>

Web of science

(TOPIC: (solasodin*) OR TOPIC: (solasonin*) OR TOPIC: (solamargin*) OR TOPIC: (soladulcin*) OR TOPIC: (solamarin*) OR TOPIC: (solanidin*)) AND (TOPIC: (toxic*) OR TOPIC: (adverse) OR TOPIC: (terato*))
Results: 103

Pubmed

(((((((((solasodin*) OR solasonin*) OR solamargin*) OR soladulcidin*) OR soladulcin*) OR solamarin*) OR soladinin*)) AND (((toxic*) OR adverse) OR terato*)
Results:87

Scopus

((TITLE-ABS-KEY(solasodin*) OR TITLE-ABS-KEY (solasonin*) OR TITLE-ABS-KEY (solamargin*) OR TITLE-ABS-KEY (soladulcidin*) OR TITLE-ABS-KEY (soladulcin*) OR TITLE-ABS-KEY

(solamarin*) OR TITLE-ABS-KEY (soladinin*))) AND ((TITLE-ABS-KEY (toxic*) OR TITLE-ABS-KEY (adverse) OR TITLE-ABS-KEY (terato*)))
Results: 107

b) Leptin alkaloids

Web of science
TOPIC: (leptinin*) OR TOPIC: (leptinidin*) OR TOPIC: (leptin*) AND TOPIC: (toxic*) OR TOPIC: (adverse) OR TOPIC: (terato*) NOT TS= (fat OR adipos* OR adipocyt* OR diabetes OR obese OR obesity OR insulin* OR hormon* OR disease*)
Results:335

Pubmed
(((((leptinin*) OR leptinidin*) OR leptin*)) AND (((toxic*) OR adverse) OR terato*)) NOT (((((((((fat) OR adipos*) OR adipocyt*) OR diabetes) OR obese) OR obesity) OR insulin*) OR hormon*) OR disease*)
Results:294

Scopus
((TITLE-ABS-KEY (toxic*) OR TITLE-ABS-KEY (adverse) OR TITLE-ABS-KEY (terato*))) AND ((TITLE-ABS-KEY (leptinin*) OR TITLE-ABS-KEY (leptinidin*) OR TITLE-ABS-KEY (leptin*))) AND NOT ((TITLE-ABS-KEY (fat) OR TITLE-ABS-KEY (adipos*) OR TITLE-ABS-KEY (adipocyt*) OR TITLE-ABS-KEY (diabetes) OR TITLE-ABS-KEY (obese) OR TITLE-ABS-KEY (obesity) OR TITLE-ABS-KEY (insulin*) OR TITLE-ABS-KEY (hormon*) OR TITLE-ABS-KEY (disease*)))
Results: 321

All the retrieved references were uploaded in Endnote. Final number of hits after removal of duplicates were

- **Tomato alkaloids**: 183 articles
- **Solanine alkaloids**: 187 articles
- **Leptin alkaloids**: 655 articles

The articles were screened for their relevance, exclusion criteria:

- **Tomato alkaloids**: analytics/detection of compounds, synthesis of compounds, pest resistance, positive/pharmaceutical effects, plant defence mechanisms
- **Solanine alkaloids**: analytics/detection of compounds, pest resistance, positive/pharmaceutical effects/clinical trials, toxicity on snails
- **Leptin alkaloids**: synthesis of compounds, insects (e.g. *Leptinotarsa*), plant defence mechanism, pest resistance

Appendix C – Details of the study design of the toxicokinetic studies

5286 **Table C.1.** Studies on the toxicokinetics of glycoalkaloids (GAs) in experimental animals.

Test compound	Study design (species, doses, administration routes, study duration)	Parameters	Reference
[^{3}H]-α-chaconine (tritiated in the aglycone moiety at carbon atoms adjacent to the double bond and to the nitrogen atom), activity	Male Sprague-Dawley rats (200–300 g bw) Dosage by gavage (5 mg/kg bw) or i.p. injection (5, 10, 15 or 25 mg/kg bw) Rats housed in metabolic cages. 2 rats/treatment group euthanized at each time point Timepoints: 3, 6, 12, 24, 72 or 100 h following treatment	Radioactivity distribution and elimination. Metabolism studied by means of thin layer chromatography following ether extraction	Norred et al. (1976)
[^{3}H]-α-chaconine (no information on labelling position)	Female Swiss-Webster mice (20 g bw) Experiments: • Subcellular distribution: oral dose of 10 mg/kg bw, 4 mice/timepoint (3, 6, 14, 72 and 120 h), • Accumulation in subcellular fractions: 5 mice/group, oral doses of 1, 3 or 10 mg/kg bw, • Enzyme induction: 4 mice group orally exposed to 10 mg/kg bw [^{3}H]-α-chaconine and by i.p. injection either to 100 mg/kg bw phenobarbital, 100 mg/kg bw Aroclor, or 0.5 mg/kg bw dibenzo[a]pyrene.	Subcellular distribution Accumulation in subcellular fractions Metabolism (enzyme induction)	Sharma et al. (1983)
[^{3}H]-α-chaconine (randomly labelled)	Male Golden hamsters (130–150 g bw) Gavage or i.p. injection at a dose of 10 mg/kg bw 3 animals/time point euthanised at regular time intervals (3, 12, 24, 72 and 168 h following treatment)	Tissue distribution Excretion Subcellular distribution	Alozie et al. (1979a)
[^{3}H]-α-chaconine	Study design described in Aloize et al. (1979a)	Metabolite identification	Alozie et al. (1979b)

(randomly labelled)	Collected urine and feces from animals orally exposed were repeatedly extracted with chloroform. The water soluble and chloroform soluble extracts were subject to TLC analysis for the identification of metabolites.		
[^{3}H]-α-solanine (no information on the labelling position)	Male rats (strain SPF Riv:TOX rats; 240–260 g bw; 12–13 weeks of age, implanted with a Colonisation Resistant Factor mouse flora) Male hamsters (SPF Charles River/Wiga Syrian golden hamsters; 130–160 g bw, 12–15 weeks of age) Dosage either by gavage (in 4 rats and 5 hamsters) at a dose of 170 µg/kg bw, or via i.v. injection (in 5 rats and 5 hamsters) at a dose of 54 µg/kg bw, respectively Animals were housed in metabolic cages for 7 days following the treatment. Urine, feces and exhaled air. Blood was regularly collected over the period after the treatment and analysed for the radioactivity level of unchanged α-solanine	Absolute bioavailability Kinetics in blood and plasma Excretion	Groen et al. (1993)
[^{3}H]-α-solanine (tritiated in the aglycone moiety at carbon atoms adjacent to the double bond and to the nitrogen atom)	Male Fischer rats (180–250 g bw for rats subject to oral treatment, 95–170 g bw for rats treated by i.p. injection) Dosage by gavage (5 mg/kg bw) or i.p. injection (5, 10, 15 or 25 mg/kg bw). Number of rats/treatment group was not reported Rats were housed in metabolic cages and sacrificed after 3, 6, 12, 24, 48, 72 or 96 hours after treatment Metabolites in urine and feces were studies by TLC analysis	Tissue distribution Metabolic fate Excretion	Nishie et al. (1971)

bw: body weight. i.p.: intraperitoneal. i.v.: intravenous. TLC: thin-layer chromatography.

5292 **Table C.2.** Details of studies reporting on the toxicokinetics of glycoalkaloids (GAs) in humans.

Test compound	Study design (subjects, doses, administration routes, study duration)	Parameters	Reference
α-solanine, α-chaconine and solanidine	7 healthy male adults Treatment with a portion of peeled mashed potatoes containing 200 mg TGAs/kg (82 and 118 mg/kg for α-solanine and α-chaconine, respectively), adjusted to reach a dose of 1 mg TGAs/kg bw The study subjects were instructed to refrain from eating potatoes and potato products as from 48 h before and up to 25 h after the exposure Blood was sampled prior to the exposure and 1, 2, 3, 4, 5, 8 and 25 h following the exposure Levels of α-solanine, α-chaconine and solanidine in serum were measured via HPLC analysis	Kinetic in blood serum (Cmax, Tmax, $t_{1/2}$)	Hellenäs et al. (1992)
α-solanine, α-chaconine and solanidine	6 male and 8 female healthy volunteers (age range: 18–45 years old) Dietary rules to refrain from other products containing GAs were followed as from 72 h prior to the treatment Treatments: • 0.30, 0.50 or 0.70 mg TGA/kg bw in 200 mL aqueous solutions (2 subjects/dose group). The solutions contained 50% α-solanine and 50% of α-chaconine • 0.95, 1.10 or 1.25 mg total TGA/kg bw in portions of a mashed potato meal (3, 3 and 2 subjects/dose group, respectively). The GA content of the potato meal was 51% for α-solanine and 49% for α-chaconine. Blood was withdrawn 1 h and 30 min before treatment and 1, 2, 3, 4, 5, 6, 7, 8, 10, 12, 16, 20, 24, 32, 48, 56, 72 and 96 h after treatment Serum α-solanine, α-chaconine and solanidine levels were measured by HPLC analysis	Kinetic in blood serum (Cmax, Tmax, $t_{1/2}$)	Mensinga et al. (2005)

[^{3}H]-solanidine (tritiated in the aglycone moiety at carbon atoms adjacent to the nitrogen atom)	Two male and one female volunteers Treatment via a single i.v. injection (doses not reported) Blood was sampled at various times up to 150 h Urine was collected at 2, 4, 8 and 12 h Feces were collected as individual voidings The distribution of solanidine in liver was studied in five post-mortem liver samples. Four solanidine fractions were extracted: • Free-solanidine was extracted with dichloromethane from liver homogenates. • Beta-glucuronide conjugates were extracted following treatment with β-glucuronidase subsequent extraction with dichloromethane • Glycoside fraction was isolated following treatment with mixed glycosidases and subsequent extraction with dichloromethane • Finally, the residual aqueous fraction was treated with hydrochloric acid and extracted with dichloromethane (acid-hydrolysed fraction)	Kinetic in plasma, urine and feces Distribution of solanidine in liver	Claringbold et al. (1982)
Solanidine	34 human donors (7 males and 27 females), randomly selected from a hospital clinic Development of a radioimmunoassay method to quantify solanidine in human plasma. Serum from rabbits immunized using a solanine-BSA conjugated was used. 100% cross reactivity was found for solanidine, α-solanine, α-chaconine and desmissidine. A chloroform extraction procedure showed allowed the specific quantification of solanidine	Determination of solanidine in plasma	Matthew et al. (1983)
Solanidine	57 volunteers (30 males, age range: 18–44 years old; 27 females, age range: 16–62 years old) Solanidine levels in plasma were analysed using the radioimmune assay developed by Matthew et al. (1983) The study subjects were asked to maintain their standard diet during winter (November-December), with the exclusion of two male volunteers who maintained a	Determination of solanidine in plasma	Harvey et al. (1985a)

	potato-free diet for several weeks while their serum was regularly analysed for solanidine levels The consumption of potatoes and potato products of 33/57 subjects was recorded for one month.		
Solanidine, TGA	Two groups of volunteers: • From UK: 18 males (18–41 years old), and 15 females (19–63 years old) • from Sweden: 5 males (31–41 years), and 5 females, 61–67 years) GAs levels were analysed using a radioimmuno method adapted from Matthew et al. (1983). Exposure to GAs through normal diet (during summer), with the exclusion of three Swedish volunteers (2 m and 1 f), who had daily consumption of two potato cultivars known to be high in GAs ('Ulster Chieftain' and 'SV72118') for a week before serum and saliva sampling. Blood and saliva samples were collected from all volunteers and analysed for the presence of solanidine (upon chloroform extraction) or total GA levels.	Solanidine and total GA levels in blood and saliva	Harvey et al. (1985b)

HPLC: high performance liquid chromatography.

Appendix D – Comparison of developmental toxicity of single dose studies

5296 **Table D.1.** Comparison of developmental toxicity of GAs and their aglycones from single dose studies.

Compound	Dose (mmol/kg bw) *mmol/kg diet	Dose (mg/kg bw)	Abnormal litters (%)	Abnormal fetuses (%)	Reference
Solanidine N-oxide	0.14	58	0	0	Gaffield and Keeler (1996)
Solanidine	0.29	115	19	2.3	Gaffield and Keeler (1996)
Solanidine N-oxide	0.29	120	0	0	Gaffield and Keeler (1996)
Solanidine	0.43	175	50	16	Gaffield and Keeler (1996); other study
Solanidine	0.44	176	50	24	Gaffield and Keeler (1993)
Demissidine	0.44	176	14	3	Gaffield and Keeler (1996)
Demissidine	0.44	176	14	3	Gaffield and Keeler (1993)
Solanidine N-oxide	0.43	178	50	16	Gaffield and Keeler (1996)
α-Chaconine	0.19	165	63	23	Renwick et al. (1984)
α-Chaconine	0.21	179	20	2.7	Gaffield and Keeler (1996)
α-Chaconine	0.21	180	88	18	Renwick et al. (1984)
α-Solanine	0.23	200	59	23	Renwick et al. (1984)
α-Solanine	0.28	243	21	5.5	Gaffield and Keeler (1996)
α-Chaconine	0.29	247	Not measurable, maternal lethality	Not measurable, maternal lethality	Gaffield and Keeler (1996)
Solasodine	3.4	1,400	75	29	Gaffield and Keeler (1993)
Dihydrosolasodine	3.4	1,400	45	6	Gaffield and Keeler (1993)

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Appendix E - Inhibition of cholinesterases by GAs

5298 **Table E.1**. Inhibition of cholinesterases by GAs. Note: In older papers, e.g. Orgell (1963) existance of BuChE was not known, data were reported as AChE
5299 activity with no discrimination made.

Compound	Species	AChE		BuChE		Citation
		dose	% inhibition	dose	% inhibition	
α-Solanine	Bovine erythrocyte	5 µM	27.7			Roddick (1989)
α-Solanine	Bovine erythrocyte	10 µM	44.3			
α-Chaconine	Bovine erythrocyte	5 µM	25.9			
α-Chaconine	Bovine erythrocyte	10 µM	54.4			
α-Solanine + α-chaconine	Bovine erythrocyte	5+5 µM	37,3			
α-Solasonine	Bovine erythrocyte	50 µM	-12			
α-Solasonine	Bovine erythrocyte	100 µM	16.7			
α-Solamargine	Bovine erythrocyte	50 µM	-3.6			
α-Solamargine	Bovine erythrocyte	100 µM	13			
Solanindine (0 h)	Bovine erythrocyte	100 µM	11.1			Roddick (1989)
Solanidine (1 h)	Bovine erythrocyte	100 µM	5.7			
Solanidine (2 h)	Bovine erythrocyte	100 µM	2.0			
Solanidine (0 h)	Human erythrocyte	100 µM	-6.2			
Solanidine (1 h)	Human erythrocyte	100 µM	3.6			
Solanidine (2 h)	Human erythrocyte	100 µM	14.1			
Tomatidine (0 h)	Bovine erythrocyte	100 µM	15.6			
Tomatidine (1 h)	Bovine erythrocyte	100 µM	-7.7			
Tomatidine (2 h)	Bovine erythrocyte	100 µM	-4.5			
Tomatidine (0 h)	Human erythrocyte	100 µM	-13.8			
Tomatidine (1 h)	Human erythrocyte	100 µM	-13.8			
Tomatidine (2 h)	Human erythrocyte	100 µM	-10.6			
Solasodine (0 h)	Bovine erythrocyte	100 µM	8.9			
Solasodine (1 h)	Bovine erythrocyte	100 µM	-15			
Solasodine (2 h)	Bovine erythrocyte	100 µM	-1.5			
Solasodine (0 h)	Human erythrocyte	100 µM	-8.3			
Solasodine (1 h)	Human erythrocyte	100 µM	-2.4			

Solasodine (2 h)	Human erythrocyte	100 µM	0.6			
α-Chaconine	Eel	33.8 ppm (39.6 µM)	26.8			Bushway et al. (1987)
β-Chaconine	Eel	33.8 ppm (40 µM)	23.3			
α-Solanine	Eel	33.4 ppm (38.5 µM)	26.3			
Dehydrocommersonine	Eel	33.7 ppm (32.2 µM)	18.5			
Commersonine	Eel	34.5 ppm (32.9 µM)	20.2			
Demissine	Eel	34.3 ppm (33.6 µM)	21.6			
α-Tomatine	Eel	34 ppm (32.8 µM)	4.2			
Solanidine	Eel	33.8 ppm (85.1 µM)	15.4			
Demissidine	Eel	40 ppm (100 µM)	11.3			
Tomatidine	Eel	41.3 ppm (99.5 µM)	4.2			
Potato peel	Human serum (unusual phenotype)	(85 g potato peel extracted by 300 mL water)	77–83			Harris and Whittaker (1959)
Potato peel	Human serum (intermediate phenotype)	(85 g potato peel extracted by 300 mL water)	53–67			
Potato peel	Human serum (atypical phenotype)	(85 g potato peel extracted by 300 mL water)	16–24			
α-Solanine	Human serum (unusual phenotype)	2.88 µM	86.2			Harris and Whittaker (1962)
α-Solanine	Human serum (intermediate phenotype)	2.88 µM	65.8			
α-Solanine	Human serum (atypical phenotype)	2.88 µM	21			
Solanidine	Human serum (unusual phenotype)	3.14 µM	80			

Solanidine	Human serum (intermediate phenotype)	3.14 µM	53.2			
Solanidine	Human serum (atypical phenotype)	3.14 µM	0.8			
α-Chaconine	Rat plasma isoenzyme 1	100 µM	0			Alozie et al. (1978)
α-Chaconine	Rat plasma isoenzyme 2	100 µM	0			
α-Chaconine	Rat plasma isoenzyme 3	100 µM	78.5			
α-Chaconine	Rat plasma isoenzyme 4	100 µM	81.7			
α-Chaconine	Rat plasma isoenzyme 5	100 µM	90			
α-Chaconine	Rat plasma isoenzyme 6	100 µM	100			
α-Chaconine	Rat erythrocyte isoenzyme 1	100 µM	0			
α-Chaconine	Rat erythrocyte isoenzyme 2	100 µM	42			
α-Chaconine	Rat erythrocyte isoenzyme 3	100 µM	42			
α-Chaconine	Rat erythrocyte isoenzyme 4	100 µM	100			
α-Chaconine	Rat erythrocyte isoenzyme 5	100 µM	100			
α-Chaconine	Rat erythrocyte isoenzyme 6	100 µM	100			
α-Chaconine	Rat brain isoenzyme 1	100 µM	23			
α-Chaconine	Rat brain isoenzyme 2	100 µM	27.1			
α-Chaconine	Rat brain isoenzyme 3	100 µM	15.6			
α-Chaconine	Rat brain isoenzyme 4	100 µM	19.5			
α-Chaconine	Rat brain isoenzyme 5	100 µM	67.3			

α-Chaconine	Rat brain isoenzyme 6	100 µM	100			
α-Chaconine	Human purified	100 µM	67.3	100 µM	92.8	McGhee et al. (2000)
α-Solanine	Human purified	100 µM	76.8	100 µM	91.5	
α-Chaconine (0/10/30 min)	Human serum			2.88 µM	68.3/70.2/67.3	Nigg et al. (1996)
α-Solanine (0/10/30 min)	Human serum			2.88 µM	50/50/50	

5300 **Table E.2.** Determined Ki values.

Compound	Species	AChE	BuChE	Citation
		K_i (µM)	K_i (µM)	
α-Solanine	Human plasma	5		Orgell (1963)
α-Tomatine	Human plasma	100		
Chaconine (TLC preparation)	Purified bovine cholinesterase	8,3		Alozie et al. (1978)
	Horse serum cholinesterase	4		
α-Chaconine	equine BuChE		1.39	Benilova et al. (2006)
α-Solanine	equine BuChE		3.33	
α-Tomatine	equine BuChE		1.66	
α-Chaconine	human BuChE		0.17	
α-Solanine	human BuChE		0.22	
α-Tomatine	human BuChE		1.3	
α-Solanine	Human serum (D/D; wildtype, homozygous)		2.9	Sternfeld et al. (1997)
α-Solanine	Human serum (D/G; heterozygous)		3.6	
α-Solanine	Human serum (G/G; homozygous)		165	
α-Solanine	Human recombinant (D/D; wildtype, homozygous)		2.0	
α-Solanine	Human recombinant (D/G; heterozygous)		3.5	
α-Solanine	Human recombinant (G/G; homozygous)		171	

α-Solanine	Human, purified	14	0.17	McGehee et al. (2000)
α-Chaconine	Human, purified	17	0.066	
α-Solanine	Human wildtype		3.3	Loewenstein-Lichtenstein et al. (1996)
α-Solanine	Human atypical (D70G)		78	
α-Solanine	Various artificial mutations		0.3–78	

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Appendix F - Rapid Alert System for Food and Feed (RASFF) reports on the presence of *S. nigrum* in food products

5303 **Table D.1.** Rapid Alert System for Food and Feed (RASFF) reports on the presence of Solanum nigrum in food products

product category	date	reference	product type	notification type	notification basis	notified by	countries concerned	subject	action taken	distribution status	risk decision
dietetic foods, food supplements, fortified foods	9-5-2018	2018.1276	food	information for follow-up	official control on the market	Poland	Cyprus (D), Germany (D), Greece (D), India (O), Italy (D), Latvia, Poland (D), Romania (D), Slovenia (D), Spain	Solanum nigrum and unauthorised novel food ingredients Cassia occidentalis, Achillea millefolium and Tamarix gallica in food supplement from India, via Latvia and via Spain		distribution to other member countries	undecided
dietetic foods, food supplements, fortified foods	9-5-2018	2018.1278	food	information for follow-up	official control on the market	Poland	Austria (D), Bulgaria (D), Cyprus (D), France (D), Germany (D), Greece (D), India (O), Italy (D), Latvia, Lithuania (D), Poland (D), Portugal (D), Romania (D), Slovakia (D), Slovenia (D), Spain (D), United Kingdom (D)	Solanum nigrum and unauthorised novel food ingredients Cassia occidentalis, Achillea millefolium and Tamarix gallica in food supplement from India, via Latvia and via Spain		distribution to other member countries	undecided
dietetic foods, food supplements, fortified foods	6-3-2015	2015.AKE	food	border rejection	border control - consignment detained	Portugal	India (O), Portugal	unauthorised placing on the market (Solanum nigrum, Sida cordifolia Operculina turpethum and Nepeta	import not authorised	product not (yet) placed on the market	undecided

								hindostana) of food supplements from India			
fruits and vegetables	7-6-2013	2013.0804	food	alert	official control on the market	Germany	Germany (D), Netherlands (O)	Solanum nigrum (black nightshade) in young green beans from the Netherlands	recall from consumers	distribution restricted to notifying country	serious
fruits and vegetables	18-2-2004	2004.086	Food	alert	consumer complaint	France	Belgium (O), France (D)	foreign body (50 green solanum nigrum in 2 samples) in frozen green beans	product recall or withdrawal		undecided
fruits and vegetables	6-11-1985	1985.15	Food	alert		Germany	Germany, Netherlands (O)	Solanum nigrum (Berries of solanum nigrum) in beans - canned			undecided
fruits and vegetables	13-10-1982	1982.07	Food	alert		Italy	Belgium (O), Italy	Solanum nigrum in Green beans - quick-frozen			undecided

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Appendix G - Studies on the toxicity of Glycoalkaloids not considered in the risk assessment

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Excluded studies	Reason for exclusion
Baker DC, Keeler RF and Gaffield WP, 1988**.** Mechanism of death in Syrian hamsters gavaged potato sprout material. Toxicologic Pathology, 16, 333-339.	In the study no GA concentrations were determined.
Bell DP, Gibson JG, McCarroll AM and McClean GA, 1976**.** Embryotoxicity of solanine and aspirin in mice. Journal of Reproduction and Fertility, 46, 257-259.	i.p. study. Although the authors call it solanine, in fact a-solanine was used (standard from Sigma)
Caldwell KA, Grosjean OK, Henika PR and Friedman M, 1991. Hepatic ornithine decarboxylase induction by potato glycoalkaloids in rats. Food Chemistry and Toxicology, 29, 531-535.	Excluded because additional application of dexamethasone in all treatment groups, no proper control.
Candeias MF, Abreu P, Pereira A and Cruz-Morais J, 2009. Effects of strictosamide on mouse brain and kidney Na^+, K^+-ATPase and Mg^{2+}-ATPase activities. Journal of Ethnopharmacology, 12, 117-22.	Strictosamide is not a potato GA.
Chaube S, Swinyard CA and Daines RH, 1973. Failure to induce malformations in fetal rats by feeding blighted potatoes to their mothers. Lancet, 1, 329-330.	In the study no GA concentrations were determined.
Crawford L and Myrh B, 1995. A preliminary assessment of the toxic and mutagenic potential of steroidal alkaloids in transgenic mice. Food Chemistry and Toxicology, 33, 191-194.	A preliminary study without proper controls and without value for the mutagenicitiy assessement.
Friedman M, Rayburn JR and Bantle JA, 1992. Structural relationships and developmental toxicity of Solanum alkaloids in the frog embryo teratogenesis assay-*Xenopus*. Journal of Agricultural and Food Chemistry, 40, 1617-24.	Studies in Xenopus are not informative for the risk assessment.
Gupta AK, Ganguly P, Majumder UK and Ghosal S, 2009. Antidiabetic and antihyperlipidaemic effects of *Solanum Xanthocarpum* total extract in alloxan induced diabetic rats. Pharmacologyonline, 1, 484-97.	In the study no GA composition and concentration were determined
Hellenäs KE, Cekan E, Slanina P and Bergman K, 1992. Studies of embryotoxicity and the incidence of external malformations after continuous intravenous infusion of α-chaconine in pregnant rats. Pharmacology and Toxicology, 70, 381-3.	i.v. excluded from the consideration in the experimental animal section, except in the acute toxicity for comparison with the oral route.
Hornfeldt CS and Collins JE, 1990. Toxicity of nightshade berries (*Solanum dulcamara*) in mice. Clinical Toxicology, 28, 185-92.	In the study no GA composition and concentration were determined.
Ji Y, Sun J and Lang L, 2011. Analysis of testicular toxicity of solanine in mice. Advanced Materials Research, 282, 195-200.	i.p. administration.
Patel VB, Rathod IS, Patel JM and Brahmbhatt MR 2010. Anti-urolithiatic and natriuretic activity of steroidal constituents of *Solanum xanthocarpum*. Der Pharma Chemica, 2, 173-6.	No proper methodical description of the experiments and no proper description of isolation and purification procedure. Purity of the tested substances is highly questionable.
Peičev P, Nikiforov N and Rusev R, 1967. Pharmacological properties of glycoalkaloids, obtained from Solanum tuberosum L. Experimental Medicine and Morphology. VI, 1.	Extract used of Herba Solani (leaves of *S. tuberosum*), puritiy

	of isolated GA is not described, documentation is incomplete.
Poswillo DE, Sopher D and Mitchell S, 1972. Experimental induction of foetal malformation with ''blighted'' potato: a preliminary report. Nature, 239, 462-4.	In the study no GA composition and concentration were determined
Poswillo DE, Sopher D, Mitchell SJ, Coxon DT, Curtis RF and Price KR, 1973. Further investigations into the teratogenic potential of imperfect potatoes. Symposium on Potatoes and Birth defects, 8, 339-47.	Only TGA content reported. Reported TGA content is much lower than generally found in (healthy) potato tubers, indicating an error in the analytical methodology used.
Schwarz A, Pinto E, Haraguchi M, de Oliveira CA, Bernardi MM and de Souza Spinosa H, 2007. Phytochemical study of *Solanum lycocarpum* (St. Hil) unripe fruit and its effects on rat gestation. Phytotherapy Research, 21, 1025-1028.	Extracts of *S. lycocarpum* were tested, which is not a food plant.
Sharma RP, Willhite CC, WU MT and Salunkhe DK, 1978. Teratogenic potential of blighted potato concentrate in rabbits, hamsters and miniature swine. Teratology, 18, 55-62.	In the study no GA composition and concentration were determined
Wang XG, 1993. Teratogenic effect of potato glycoalkaloids. Zhonghua Fu Chan Ke Za Zhi, 28, 121-2.	Abstract only. Intra-abdominal administration. Article in Chinese.
Yoon DJ and Kirkowski A, 1979. Effects of dietary Solanaceous potatoes on the serum, calcium, phosphorus, magnesium and hydroxyproline levels in rats. Korean Journal of Animal Science, 21, 401-6.	In the study no GA composition and concentration were determined
Allen et al 1977. Teratogenicity studies on late blighted potatoes in nonhuman primates (Macaca mulatta and Saguinus labiatus). Teratology, 15, 17-23	Only TGA content reported. Reported TGA content is much lower than generally found in (healthy) potato tubers, indicating an error in the analytical methodology used.
Rao MV, 1988. Effects of alcoholic extract of Solanum xanthocarpum seeds in adult male rats. Indian Journal of Experimental Biology, 26, 95–98.	Extract of *Solanum xanthocarpum* seeds. In the study no GA composition and concentration were determined.
Mali PC, Chaturvedi M and Dixit VP, 1996. Antispermatogenic activity of Solanum xanthocarpum S and W (50 percent EtOH-extract) in rats. Journal of Phytological Research, 9, 13–17.	Extract of *Solanum xanthocarpum* root. In the study no GA composition and concentration were determined.

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Appendix H – Additional scenario for the human risk characterisation

Table H.1. Summary statistics of the % of survey days with an intake of potato TGAs above the LOAEL of 1 mg/kg bw calculated only for days of potato consumption. The corresponding 95% confidence intervals (95% CI) are given in the brackets.

Age class	Range of mean % of days above the LOAEL of 1 mg/kg bw per day [(a)]	
	Minimum	**Maximum**
Infants	<0.1 (0 - <0.1)	0.8 (0.4 - 1.3)
Toddlers	0.3 (0.1 - 0.5)	1.7 (0 - 5.0)
Children	0.1 (0 - 0.4)	1.3 (0 - 3.2)
Adolescents	<0.1 (0 - 0.2)	0.7 (0 - 1.4)
Adults	<0.1 (0 - 0.1)	0.4 (0.1 - 0.7)
Elderly	<0.1 (0 - <0.1)	0.3 (0 - 1.1)
Very elderly	<0.1 (0 - 0.2)	0.3 (0 - 1.6)

(a) The mean percentage of days above the LOAEL of 1 mg/kg bw per age group for each survey was calculated by averaging the percentage of days above the reference for each subject and then averaged over all subjects within the age group for each survey. The process was iterated 1,000 times as for the exposure estimates, and a mean of the mean percentage of days above the reference per age group was calculated together with a 95% confidence interval.

Annex A – Occurrence data in food and feed submitted to EFSA and dietary exposure assessment for humans

Annex A can be found as a separate document.

Description: this Annex is an excel file which presents tables on GAs on occurence data in food and feed, and dietary exposure assessment for humans.